

**CONCENTRATION OF HEAVY METALS (Cd and Cr) IN
VEGETABLES COLLECTED FROM DIFFERENT
MARKETS OF DHAKA CITY AND
HEALTH RISK ASSESSMENT**

MD. RIAZ MAHMUD



DEPARTMENT OF AGRICULTURAL CHEMISTRY

**SHER-E-BANGLA AGRICULTURAL UNIVERSITY
DHAKA-1207**

JUNE-2018

**CONCENTRATION OF HEAVY METALS (Cd and Cr) IN
VEGETABLES COLLECTED FROM DIFFERENT
MARKETS OF DHAKA CITY AND
HEALTH RISK ASSESSMENT**

BY

MD. RIAZ MAHMUD

REGISTRATION NO. 12-04781

A Thesis

*Submitted to the Faculty of Agriculture,
Sher-e-Bangla Agricultural University, Dhaka
in partial fulfillment of the requirements
for the degree of*

MASTER OF SCIENCE (MS)

IN

AGRICULTURAL CHEMISTRY

SEMESTER: JANUARY- JUNE, 2018

APPROVED BY:

Dr. Rokeya Begum
Professor
Supervisor

Dr. Md. Sirajul Islam Khan
Associate Professor
Co-supervisor

Dr. Md. Sirajul Islam Khan
Chairman
Examination Committee



DEPARTMENT OF AGRICULTURAL CHEMISTRY
Sher-e-Bangla Agricultural University
Sher-e-Bangla Nagar, Dhaka-1207

CERTIFICATE

This is to certify that the thesis entitled “CHEMICAL COMPOSITION OF SESAME (Sesamum Indicum L.) UNDER DIFFERENT RATES OF MICRONUTRIENTS APPLICATION” submitted to the Faculty of Agriculture, Sher-e-Bangla Agricultural University, Dhaka, in partial fulfillment of the requirements for the degree of Master of Science in Agricultural chemistry, embodies the result of a piece of bona fide research work carried out by Md. Nazmul Islam Shekh, Registration number: 12-04842 under my supervision and guidance. No part of the thesis has been submitted for any other degree or diploma.

I further certify that any help or source of information, received during the course of this investigation has duly been acknowledged.



Dated: June, 2018
Place: Dhaka, Bangladesh

Dr. Rokeya Begum
Professor
Supervisor



**DEDICATED
TO
MY BELOVED PARENTS**

ACKNOWLEDGEMENTS

First of all, I express my best gratitude to Almighty Allah for His never-ending blessing to complete this work successfully. It is a great pleasure to express profound thankfulness to my respected parents, who entitled much hardship inspiring for prosecuting my studies, thereby receiving proper education.

*I would like to express my earnest respect, sincere appreciation and enormous indebtedness to my reverend Supervisor **Prof. Dr. Rokeya Begum**, Department of Agricultural Chemistry, Sher-e-Bangla Agricultural University, Dhaka-1207, for his scholastic supervision, helpful commentary and unvarying inspiration throughout the research work, and preparation of the thesis.*

*I wish to express my gratitude and best regards to my respected Co-Supervisor, **Dr. Md. Sirajul Islam Khan**, Associate Professor, Department of Agricultural Chemistry, Sher-e-Bangla Agricultural University, Dhaka-1207, for his continuous direction, constructive criticism, encouragement and valuable suggestions in carrying out the research work and preparation of this thesis.*

*I am also highly grateful to my honorable, **Md. Khairul Alam**, Scientific Officer in Soil Science Division, BARI, Joydebpur, Gazipur-1701, for his valuable teaching, direct and indirect advice, and encouragement and co-operation during the study period.*

I would like to thank Md. Eunuse Ali Hera, Riaz Mahmud, Parves Sarder, Md Rafiqul Islam, Md. Tanvir zubayer, Md. dip and Farzana Akter Laboni who have helped me so much throughout the research work and preparation of the thesis.

I feel to express my heartfelt thanks to all the teachers of the Department of Agricultural Chemistry, Sher-e-Bangla Agricultural University, Dhaka-1207, for their valuable suggestions and encouragement during the period of the study.

I also wish to extend sincere gratitude to the Ministry of Science and Technology, Government of the People's Republic of Bangladesh for providing me financial support to conduct the research.

I found no words to thank my father Md. Nurul Islam Shekh, mother Morsheda Begum, Sister Sumaiya Islam Soma and Brother Md. Zahidul Islam Shekh for their unquantifiable love and continuous support, their sacrifice never ending affection, immense strength and untiring efforts for bringing my dream to proper shape. They are my constant source of inspiration, zeal and enthusiasm in the critical moment of my studies.

***June, 2018
SAU, Dhaka***

The Author

**CHEMICAL COMPOSITION OF SESAME (*Sesamum indicum* L.)
UNDER DIFFERENT RATES OF MICRONUTRIENTS
APPLICATION**

BY

MD. NAZMUL ISLAM SHEKH

ABSTRACT

A field experiment entitled “Chemical composition of sesame (*sesamum indicum* L.) under different rates of micronutrients application” was conducted during Kharif-1 2017 at central farm of SAU, Dhaka. Sesame (*Sesamum indicum*.L) is one of the most valuable oilseed crops whose seeds are used as traditional health food, with high quality oil, protein and natural antioxidants. They contained high amount of protein (26-28%), considerable amount of oil (45-55%), characterized by an elevated level of unsaturated fatty acids (79.50-83.40% of UFA) and high content of stearic acid (7.3-8.6%), and remarkable content of dietary fibers. Seed oil was also found to be rich in total phenolic content, chlorophyll content and carotenoid content. It was showed that sesame seeds can be considered as a good source of natural antioxidant, particularly after roasting. Due to their distinguishable and favorable properties, sesame seeds could be useful for food, industrial and pharmaceutical purposes. The experiment was laid out in a Randomized Block Design with ten treatments and replicated thrice. The treatments were comprising of T1: control (no micronutrients), T2: B (100 ppm), T3: Zn (100 ppm), T4: Mo (50 ppm), T5: Cu (100 ppm), T6: Fe (100 ppm), T7: Mn (100 ppm), T8: Co (50 ppm), T9: (combination of above micronutrients) and T10: commercial mixture of micronutrients.. Observations on plant height, number of branches, TDM, growth analysis parameters, chlorophyll content of leaf at various stages of crop, yield and yield attributes, oil content and N, P and K uptake by plant and plant parts recorded during the course of investigation were statistically analyzed. The results revealed that most of the putative morphological traits like plant height, number of branches, TDM of plant and plant parts increased with foliar application of different micronutrients but significantly the highest increase was found when micronutrients were applied in combination followed by individual application of Zn, B and Mo. Physiological growth parameters such as LAI, LAD, RGR, NAR, CGR and chlorophyll content of leaf increased significantly by foliar application of different micronutrients either in single or in combination. Seed yield in sesame increased significantly over control by foliar application of different micronutrients used in single or in their combination. In view of such beneficial effects of foliar application of micronutrients on sesame, it may be asserted that improvement in

productivity of sesame can be achieved under a depleted soil condition through supplementary application of micronutrients like B, Zn, Mo, Cu, Fe, Mn, Co, either in single or their combination along with normal dose of N, P and K.

LIST OF CONTENTS

Chapter	Title	Page	
	Certificate	i	
	Acknowledgement	Iii	
	Abstract	v	
	List of contents	vi	
	List of tables	x	
	List of plates	xi	
	List of figures	xi	
I	Introduction	1-3	
II	Review of literature	4-13	
	2.1	Growth and development	4
	2.2	Yield and yield attributes	7
	2.3	Chlorophyll content	9
	2.4	N, P, K uptake	11
	2.5	Oil content	12
	2.6	Biochemical composition	12
III	Materials and Methods	14-31	
	3.1	Location of the experimental site	15
	3.1.1	Soil	15
	3.1.2	Climate	16
	3.1.3	Experimental details	16

		3.1.4	Design of experiment	18
		3.1.5	Collection of seeds	18
		3.1.6	Description of variety	18

LIST OF CONTENTS (contd.)

Chapter	Title			Page
		3.1.7	Details of treatments	19
	3.2	Intercultural operation		19
		3.2.1	Thinning	19
		3.2.2	Weeding	19
		3.2.3	Irrigation	19
		3.2.4	Foliar application	19
	3.3	Pre harvest observations		20
		3.3.1	Sampling techniques	20
		3.3.2	Growth observations	20
		3.3.2.1	Plant height	21
		3.3.2.2	Number of branches per plant	21
		3.3.2.3	Leaf area	21
	3.4	Physiological growth parameters		21
		3.4.1	Leaf Area Index (LAI)	21
		3.4.2	Leaf Area Duration (LAD)	21
		3.4.3	Leaf area ratio (LAR)	21
		3.4.4	Specific leaf weight (SLW)	22
		3.4.5	Specific Leaf Area (SLA)	22
		3.4.6	Relative Growth Rate (RGR)	22
III				

		3.4.7	Net Assimilation Rate (NAR)	22
		3.4.8	Crop Growth Rate (CGR)	23

LIST OF CONTENTS (contd.)

Chapter	Title		Page	
	3.5	Chemical constituent	23	
		3.5.1	Estimation of chlorophyll content	23
		3.5.2	Saponification value	24
		3.5.3	Acid value	24
		3.5.4	Estimation of fatty acid composition	25
		3.5.5	Estimation of total protein content by Microkjeldhal method	26
	3.6	Estimation of Chemical content		28
		3.6.1	Estimation of seed nitrogen content of different plant parts	28
		3.6.2	Estimation of phosphorus and potassium contents of plant sample	28
		3.6.3	Estimation of phosphorus present in plant sample	29
	III		3.6.4	Estimation of potassium present in plant sample
3.6.5			Estimation of oil content	29
3.7		Post-harvest studies		30
		3.7.1	Capsules per plant	30
		3.7.2	Shelling percent	30

		3.7.3	Thousand seed weight	30
		3.7.4	Seed yield	30
		3.7.5	Harvest index	30

LIST OF CONTENTS (contd.)

Chapter	Title		Page	
	3.8	Statistical analysis	31	
IV	Results and Discussion		32-58	
	4.1	Morphological Growth Parameters	32	
		4.1.1	Plant height and branches per plant	32
		4.1.2	Dry matter accumulation and its partitioning	34
	4.2	Physiological Traits	34	
		4.2.1	LAI, SLA, SLW and LAR	34
		4.2.2	RGR, NAR, CGR and LAD	37
		4.2.3	Chlorophyll index and total chlorophyll content	39
	4.3	Uptake of N, P, K in Plant	41	
	IV		4.3.1	N Content and Uptake
		4.3.2	P content and uptake	41
		4.3.3	K content and uptake	43
4.4		Yield, Yield Attributes and Oil Content	47	
4.5		Chemical Composition	50	
4.6		Morphological growth	51	
		4.6.1	Dry matter accumulation	52

		4.6.2	Physiological growth parameters	52
		4.6.3	Chlorophyll index and total chlorophyll content	54
	4.7	Nutrient uptake by plant and plant parts		54
		4.7.1	N content and uptake	54

LIST OF CONTENTS (contd.)

Chapter	Title			Page
		4.7.2	P content and uptake	55
		4.7.3	K content and uptake	55
	4.8	Yield and yield attributes		56
	Summary And Conclusion			59-63
V	5.1	Summary		59
	5.2	Conclusion		63
VI	References			64-69
	Appendix			70

LIST OF TABLES

Table No.	Title	Page
1	Physical composition of the soil (0-15 cm)	14
2	Chemical composition of the soil (0-15cm)	15
3	Meteorological data of Dhaka from March to June 2018	16
4	Experimental Design	18

5	Calendar of field operation	20
6	Effects of different micronutrients on plant height (cm) and number of branches per plant at different growth stages.	33
7	Effect of different micronutrients on Dry matter accumulation (g/plant) and its partitioning.	35
8	Effect of different micronutrients on LAI, SLA, SLW, LAR at different growth stages	36
9	Effect of different micronutrients on RGR, NAR, CGR and LAD at different growth period	38
Table No.	Title	Page
10	Effect of different micronutrients on chlorophyll index and total chlorophyll content at different growth stages	40
11	Effect of different micronutrients on Nitrogen content and uptake	42
12	Effect of different micronutrients on Phosphorous content and uptake	45
13	Effect of different micronutrients on Potassium content and uptake	46
14	Effect of different micronutrients on seed yield and its components	48
15	Proximate composition (%) of various parts of <i>Sesamum indicum</i> .	50
16	Mineral compositions (%) of various parts of <i>Sesamum indicum</i>	51

LIST OF PLATES

Plate No.	Title	Page
1	Some photographs show the experimental works	70

LIST OF FIGURES

Plate No.	Title	Page
1	Experimental Plot Layout	17

CHAPTER I

INTRODUCTION

Sesame (*Sesamum indicum L.*) belongs to the family Pedaliaceae is one of the significant oil crops. It is grown for seed and oil both for human ingestion and has been grown for thousand of years and today its major production areas are the tropics and the subtropics of Asia, Africa, East and Central America. In Bangladesh, it is locally known as til and is the second vital edible oil crop (Mondal *et al.*, 1997). Sesame is one of the significant oilseed crops in Bangladeshi agriculture. Sesame seeds are rich source of food, nutrition, edible oil and bio-medicine. Sesame oil has superb nutritional, medicinal, cosmetic and cooking qualities for which it is known as ‘the queen of oilseeds’. Due to the manifestation of potent antioxidants, sesame seeds are called as ‘the seeds of immortality’. Sesame cake or meal attained as a byproduct of the oil milling industry is rich in protein, vitamin (Niacin) and minerals (Ca and P). The crop is grown for its seeds, which contain 50-60% oil, 8% protein, 5.8% water, 3.2% crude fiber, 18% carbohydrate, 5.7% ash and it is very opulent in minerals such as Ca, P and vitamin E. Also, sesame oil has a very high level of unsaturated fatty acids, which is presumed to have decreasing effect on plasma cholesterol, as well as on coronary heart disease. In 2016-2017, the crop covered an area of 8530 acres in Bangladesh with the production 2970 Metric tons (BBS, 2017).

Sesame is the second most important oil-producing crop in the country. From ancient times, sesame is being cultured in the Indian subcontinent and in China. The plants may attain a height of about one metre, usually with side branches. In Bangladesh it is grown in almost all districts but grows well in greater Khulna, Faridpur, Pabna,

Barisal, Rajshahi, Jessore, Comilla, Dhaka, Rangpur, Sylhet, and Mymensingh districts (BBS, 2017).

Generally this crop is grown by the farmer with short dose of fertilizer under rainfed condition but application of nutrients at a right time is important in gaining good yields. Application of nutrients in split will fulfill the nutrient demands of plant at acute stages of the plant growth and development. Foliar fertilization is gaining more importance in recent years due to the accessibility of soluble fertilizers and is of great significance in rainfed areas under changing climatic environments. Many research reports indicated the positive effect in increasing the crop yield and quality of oilseed crops

Although micronutrients are needed in relatively small quantities for optimum plant growth and production, their deficiencies induce a great disruption in different physiological and metabolic processes in the plant. The situation has been forced with the introduction of high yielding crop varieties and intensive cropping system. As the demands of nutrients for higher yields proliferation and plant requirements for major nutrients are only met micronutrient deficiencies are likely to become acute. In Dhaka soil, the deficiency of micronutrients like Zn, B, and Mo have been reported both by farmers, extension and research workers. Low seed yield, due to deficiency of above micronutrients, have been well renowned due to several reasons such as flower and fruit drop, low harvest index and poor vegetative growth.

Phytochemical studies conducted on this plant revealed the presence of lignans, phenolics acids, flavonoids, saponins and alkaloids (Hassan and Umar 2004). The Proximate composition and mineral analysis of sesame seeds both whole and decupled at different part of the world has been determined (Hui 1996; Obiajunwa *et al.* 2005; Uaboi *et al.* 2008).

The role of different micronutrients has been well recognized in plant metabolism. Zinc (Zn), as micronutrients, is tangled in the biosynthesis of auxins, indole -3- acetic acid. It participates in the metabolism of plant as an activator of several enzymes. Boron (B) is involved in the carbohydrate transport within the plant which helps in translocation of sugar and DNA synthesis in meristems. Also it has been concerned in cellular differentiation and development, nitrogen metabolism, fertilization, active salt absorption, hormone metabolism, water relations, fat metabolism, phosphorus

metabolism and photosynthesis (Mortvedt *et al.*, 1999). Therefore, the steady amount of macro nutrients and Zn was found to increase stem height and nodes for capsule development in sesame. Major micronutrients like nitrogen, phosphorus and potassium along with micronutrients such as zinc and manganese are influencing the growth and yield of sesame.

Cobalt (Co) being a beneficial element is known to cause irreversible damage to a number of vital metabolic ingredients of plant cell and cell membrane. However it has been known for many years that Cobalt is an essential element for humans, animals and prokaryotes, but the physiological function for this element in higher plants has not been identified. The Cobalt-containing vitamin B12 does not occur in plants deficient with cobalt.

Therefore, the present study aimed to provide information on the chemical structure, available nutrients and the physicochemical properties of the oil extracted from the seed of *S. indicum*. Keeping all the above facts into consideration, present enquiry has been undertaken to study the effect of micronutrients applied as foliar spray, on metabolism, growth and yield of sesame crop with the following objectives.

OBJECTIVES

1. Effect of micronutrients on the chemical composition.
2. Effect of micronutrient on growth and yield attributes.
3. Effect of micronutrient on uptake of major nutrient by plant and oil content of seed.

CHAPTER II

REVIEW OF LITERATURE

Role of micronutrients in crop nutrition and eventually plant growth and development has been well recognized. In recent years their position on maximization of production and quality of oilseed crops has attracted the devotion of scientists all over the world. The micronutrients like iron, manganese, zinc, boron, copper, molybdenum and cobalt which are integral parts of several photosynthetic and respiratory enzymes have limiting role on absorption of crop plants and hence growth and development. Iron is essential for protein amalgamation and chloroplast development. Manganese is for cell division and expansion, zinc is essential for RNA and protein synthesis, at the same time boron is important for carbohydrate metabolism and translocation. Here attempts have been made to review the effect of micronutrients on growth, yield and quality of oilseed crop with due emphasis on sesame.

2.1 Growth and development

Boron

Sing and Sing (1996) conducted a pot culture experiment on tomato with dissimilar boron concentration 0.5, 1.0, 1.5 and 2.0 ppm and distilled water as control. They showed that boron plays an essential role in the development and growth of new cells in the plant meristem. Number of flowers and fruits increased significantly at all levels of B application and the element had positive effects on plant height, number of branches, leaves, flowers and fruit set resulting in improved number of fruits per plant and the total yield.

Nath and Biswas *et al.*, (2002) reported that foliar application of boron 100 ppm twice at monthly interval produced the maximum height of plant, increased the number of leaves per clump resulting in improved yield of spikes per plot. Foliar spray of micronutrients causes more somatic growth and increases the final plant height (Sepehr *et al.*,2002).

Bilen *et al.*, (2011) also reported that appropriate dose of boron increase plant height and higher level of boron lower urease enzyme action which was observed in 9 kg ha⁻¹ boron at sowing time that reduces plant height.

Zinc

Rajput *et al.*, (2003) in *Tagetes minuta* observed that application of boron independently or in combination with zinc and sulphur increased the plant height and fresh weight of plant significantly over the control, Individual submission of boron increased the dry matter yield significantly by 69.8 per cent and 71.1 percent over the control.

Jauhari *et al.*, (2005) found that foliar spray of zinc sulphate at 0.2 percent recorded extreme plant height, spike length, number of florets per spike and corms yield per m². Corm yield per square meter was maximum with the application of zinc sulphate at 0.4 per cent. It was also perceived that higher concentration of zinc sulphate (beyond 4 percent) had negative effects on plant growth, flowering and corm yield, revealing that 0.4 per cent zinc sulphate is the ideal concentration in gladiolus for better performance.

Molybdenum

Gupta and Vyas (1994) observed that dry weight of soybean plant was increased due to application of zinc, iron, molybdenum. Iron, zinc and molybdenum are the metallic compounds of one or more enzymes which are involved in various physiological purposes and there by increased the leaf area index, crop growth rate; relative growth rate leading to the development and productivity of plant.

Copper

The favorable effect of Zn and Cu on fruit yield in brinjal plant could be attributed to their effect in maintaining an optimum balance of nutrients in the plant for better growth and developments (Dhakshinamoorthy and Krishnamoorthy, 1989).

Iron

Zinc foliar application increases tallness, branch number per plant and dry weight of stem of grass pea. Also zinc, magnesium and iron foliar application increases growth parameters, yield and plant parts significantly (Thalooth *et al.*, 2006).

Manganese

Micronutrient play an indispensable role in cell division and development of meristematic tissues, stimulate photosynthesis; respiration, energy and nucleotide transmission reactions and fasten the plant maturity (Marschner *et al.*, 1998).

Mixture of micronutrient

Micronutrients are involved in all metabolic and cellular functions. Plants differ in their need for micronutrients; boron (B), iron (Fe), zinc (Zn), copper (Cu), chloride (Cl), manganese (Mn), molybdenum (Mo) and nickel (Ni). These elements are active that kinds them essential as catalytically active cofactors of enzymes, others have enzyme activating functions, and yet others fulfill an organizational role in stabilizing proteins. Improvement in growth characters due to micronutrient application might basically be due to enhanced photosynthetic and other metabolic activities related to cell division and elongation as opined by Hatwar *et al.*, 2003.

Ibrahim *et al.* (2007) in faba bean found that foliar application of both amino acids and micronutrients significantly increased plant height, number of branches, leaf area and number of pods per plants and subsequently the faba bean seed yield.

Yadav *et al.* (2009) studied on “effect of micronutrients in combinations with organic manures on invention and net returns of sesame (*sesamum indicum*)” in bundelkhand tract of Uttar Pradesh and found that the treatments applied with 100% of the

recommended NPKS fertilizer along with micronutrients and organic manure recorded expressively higher growth and yield attributing characters, seed yield and net returns over half of the recommended dose of fertilizer (20:10:10:15 kg NPKS/ha) coupled with micronutrients and organic manure.

Salwa *et al.* (2010) studied on “Amelioration productivity of sandy soil by using amino acids, sulphur and micronutrients for sesame production” found that there were significantly rise in the whole plant weight with increasing application of sulphur as soil application and micronutrients as foliar spray. Generally, a combined submission of amino acid with micronutrient Fe, Zn, Mn in the presence of elemental sulphur expressively increased the sesame yield; improved nutrition and increased seed quality.

2.2 Yield and yield attributes

Boron

Sakal *et al.* (1994) evaluated the direct and residual effect of varying levels of B (0, 8, 16, 32 and 64 kg Borax ha⁻¹ and FYM (0.25 and 5.0 t ha⁻¹) alone and in combinations on crops in maize-lentil cropping system. Increasing levels of B up to 16 kg borax ha⁻¹ significantly increased and higher levels declined the yield of first crop. Application of 16 kg Borax ha⁻¹ in conjunction with 5 t FYM ha⁻¹ was an ideal combination which considerably enhanced the cumulative grain yield response as well as sustained the productivity.

Talashilkar and Chavan (1996) observed that pod production of groundnut was enhanced significantly with the addition of B by 44 percent. The maximum pod and haulm yields were recorded in the treatment receiving B through super phosphate along with application of FYM, N and P. Vyakaranahal *et al.* (2001) reported that significant affect on 100-achene weight. This may be due to the B role in translocation of photo conforms from vegetative to reproductive parts of sunflower.

Zinc

Singaravel *et al.* (2002) described that the combined application of Zn and Mn in soil produced the highest number of capsules per plant by 59% and number of seeds per capsules by 57% over control. They also reported that application of Zn alone as foliar and soil increased the yield from 592 to 611 kg ha⁻¹. Zn and Mn increased

growth and yield components as well as an increased availability and better uptake of these nutrients.

Kobraee *et al.* (2011) reported that Zinc foliar application enhanced soybean yield by influencing the number of seeds per plant and seed weight. Grain yield increased significantly with each increment of zinc (Pable and Patil, 2011).

Molybdenum

Micronutrients in soybean are useful to improve productivity as well as seed quality parameters. Among the micro nutrients Mn, Zn, B, and Mo are important for increasing the productivity of soybean crop (Devarajan and Palaniappan, 1995).

Liu *et al.* (2003) studied the effects of Mo and B, alone or in mixture, on seed quality of soybean cultivars Zhechun 3, Zhechun 2, and 3811 grown in pot. Application of Mo and/or B increased the content of protein, in dispensable amino acids, total amino acids (excluding proline), N, P, K and decrease the content of Ca and oil.

Manganese

Moussavi-Nik *et al.* (1997) reported that application of iron, zinc and manganese have a significant and positive effect on wheat biomass production and yield components. Micronutrient applications significantly increased yield components, viz. number of capsules per plant and number of seeds per capsule. The combined application of Zn and Mn in soil produced the highest number of capsules per plant and number of seeds per capsules. The valuable influence of micronutrients might be due to the activation of various enzymes and the efficient utilization of applied nutrients resulting in increased yield components (Tiwari *et al.*, 2000; Shanker *et al.*, 1999).

Foliar application of micronutrients particularly Zn as well as Mn in small amounts had significant positive effect on 1000 seed weight, plant height, biological yield, grain yield, harvest index and oil content of sunflower (Babaeian *et al.*, 2011).

Shehu *et al.* (2014) studied on Manganese and Zinc nutrition of Sesame (*Sesamum indicum*) in Mubi, Northern Guinea Savannah Zone of Nigeria. And found that there were no significant ($P=0.05$) effect of Mn and Zn on stem height, number of leaves, number of branches and capsules. The different grain yield responses were associated

to their differences in Mn and Zn concentrations. Sesame seed yield increased by 2 and 5% from the application of 0.5 kg Mn ha⁻¹ and 0.5 kg Zn ha⁻¹, respectively. Dry matter increased by 13.2 and 2% from 1 kg Mn ha⁻¹ and 0.5 kg Zn ha⁻¹ rates, respectively.

Mixture of micronutrients

Yilmaz *et al.* (1997) reported that the use of micronutrients be able to increase the 1000 seed weight. Micronutrients increases photosynthesis rate and improves leaf area duration thus seed yield will be increased (Cakmak, 1999).

Kohnaward *et al.* (2012) studied on “Effect of foliar application of micronutrients on yield and yield components of safflower under conservative and ecological cropping systems” and found that significant effect of cropping system and foliar application on the number of seeds per head and number of heads per plant, and however significant interaction effect on 1000 seed weight, seed yield and biological yield. The highest 1000 seed weight (36.37 g), seed yield (4115.6 kg/ha) and biological yield (8175 kg/ha) were obtained from plants treated by zinc at high input cropping system. In all foliar sprays, there was a descending trend from along with reducing inputs, so the minimum 1000 seed weight (20.76 g), seed yield (1540.3 kg/ha) and biological yield (3112.5 kg/ha) were gained from ecological system. In conclusion, use of micronutrients, foliar application caused to improve the yield of safflower; especially in medium and low input systems.

2.3 Chlorophyll content

Schlegel *et al.* (1987) revealed that Photosynthetic pigments such as chlorophyll ‘a’, chlorophyll ‘b’ and total chlorophyll contents of *Sesamum indicum* reduced with increasing Co level in the soil. The increased chlorophyll content at lesser level of Co was obviously due to better growth.

Hemantaranjan *et al.* (2000) lead a field experiment on soybean. Plant height, root length, chlorophyll-b content, total dry matter production, seed yield of soybean were higher at 50 than 100 ppm B. However, chlorophyll-a content was higher at 100 ppm B. The collective application of S and Fe considerably increased shoot height, root length, number of leaves per plant, leaf area per plant, total dry matter production as well as seed oil content, with 80 + 20 mg per kg soil giving the best results.

Iron plays essential roles in the metabolism of chlorophylls. Exterior application of Fe increased photosynthesis, net assimilation relative growth in seawater stressed rice (Sultana *et al.*, 2001).

Iron plays an important role in the amalgamation of chlorophyll and plant growth regulators. Iron improves photosynthesis and assimilates transportation to sinks and finally enlarged seed yield (Alvarez - Fernandez *et al.*, 2006).

Iron is an essential micro element for plant development; it plays an important role in the formation of chlorophyll A and chlorophyll B, carbohydrate production, cell respiration and chemical reduction of nitrate (Mousa *et al.*, 2010).

Iron plays an important role in the stimulation of chlorophyll and in the synthesis of many heme proteins such as different cytochrome, which participate in different functions in the plant metabolism (Al -Bamarny *et al.*, 2010).

Chlorophyll and carotenoids are important quality parameters because they have a relationship with color, which is a basic attribute for evaluating oil superiority. Their magnitude depends on different factors, such as fruit ripeness, cultivar, climate conditions, type of soil, and extraction procedures. chlorophylls ranging from 0.53 to 7.57 mg/kg of oil which are responsible for the yellow color of the seed oil.

2.4. N, P, K uptake

Ramamoorthy and Sudrasan (1992) found that B application considerably increased the protein contents due to highest uptake of nitrogen by seed and N might have been incorporated in the protein molecule.

Huang *et al.* (2000) revealed that Zn deficiencies increases P content of plant shoot. They found Zn to play definite role in the regulative mechanism of the genes encoding phosphate transporter proteins in the plasma membranes of sugar beet and barley. Thus, measured uptake of P is lost under low or deficient levels of Zn leading to high concentration of P in plants.

Singaravel *et al.* (2002) recorded significant increase in K uptake from additions of Mn and Zn to NPK on Vertisols. Combined application of Zn and Mn in soil recorded the highest NPK uptake in seed and shoot. The soil or foliar application of Fe, Cu and B significantly increased N and P uptake from soil and increased their content in flax plants (El-Nagdy *et al.*, 2010).

Potarzycki and Grzebisz (2009) reported that zinc exerts a great stimulus on basic plant life processes, such as (i) nitrogen metabolism– uptake of nitrogen and protein quality; (ii) photosynthesis– chlorophyll synthesis, carbon anhydrase activity.

Application of boron at sowing time, increase the seed yield of sunflower. This might be due to applying of boron through soil, which improves the facility of crop to absorb micronutrient, leading to favorable in direct effect on uptake of nitrogen, phosphorous, potassium and yield of crop increased (Silva *et al.*, 2011).

Devi *et al.* (2012) studied the effect of sulphur, boron fertilization on yield, quality and nutrient uptake by soybean under upland condition and found useful for obtaining maximum yield attributes.

Shehu *et al.* (2014) studied on Effects of Manganese and Zinc Fertilizers on Shoot content and Uptake of N, P and K in Sesame (*Sesamum indicum* L.) on Lithosols and found that stem height, number of branches, leaves, and capsules were not significantly influenced by Mn and Zn. Seed yield was depressed at 1 kg Mn ha⁻¹, 1 kg Mn + 1 kg Zn

2.5. Oil content

In China, on a clayey soil with 0.7 mg B kg⁻¹, application of B fertilizer to *Brassica napus* L. improved plant height, pod-bearing branches and pod number per plant, seed

number per pod, seed yield and oil content (Hu *et al.*, 1994). Renukadevi and Savithri *et al.* (2003) reported that enhanced uptake of B in the seed had a significant effect on oil contents in sunflower. Zinc also plays an important role in the production of biomass, grain yield, quality and extent of oil (Kaya and Higgs, 2002; Cakmak, 2008).

Dehnavi (2008) studied the possessions of foliar application of zinc and manganese on oil content and fatty acid profiles in some safflower cultivars under drought conditions, also found that the zinc and manganese foliar applications significantly increased palmitic and oleic acids, whereas these foliar applications declined linoleic acid percent.

2.6. Biochemical composition

The protein content of the 13 Moroccan cultivars grown in different areas varied from 26.77 to 27.93%, with an average of 27.4%. The protein content depends on the climatic conditions and the stage of development of the plant. Sesame seeds protein contains high amounts of aspartic acid, glutamic acid and arginine (Prakash V *et al.*, 1975) presenting a great reservoir of amino acids. Also sesame seeds are rated among the highest organic sources containing cysteine (Rangarajan N *et al.*, 2012). Also, sesame seeds are an important source of dietary fiber. The Moroccan cultivars contain high amounts of dietary fiber, ranging from 17.55 to 20.84% of dry matter, with an average of 19.2%. The amount of insoluble dietary fibers varied from 12.46 to 15.78%, while that of soluble dietary fibers varied from 4.50 to 5.87%. Average of the latter was 5.21%, which is higher than that of cereals and its derivatives, such as corn, wheat bran, oat bran and rice bran, having a fiber content ranging from 0.4 to 4.1% (Abdul-Hamid A *et al.*, 2000).

Polyphenols, which are secondary metabolites distributed in the plant, are considered to be very important antioxidants, due to their ability to give a hydrogen atom or an electron to form an intermediate stable radical, and consequently avoid the oxidation of different biological molecules. The polyphenol content of Moroccan sesame seeds ranged from 3.75 to 3.92 mg/g. The content of flavonoids, which are also lesser metabolites, with an important role in the plant defense system, and which function as hydroxyl radical sensors and peroxides, ranged from 0.13 to 0.14 mg/g and no significant differences were detected between the cultivars. Actually, the content of

primary and secondary metabolites is strongly influenced by plant stress when exposed to severe scarcity conditions (Vishwanath et al., 2011)

Sesame is also considered as a rich source of minerals such as calcium, magnesium (which plays an important role for the support of the respiratory system), iron, potassium and selenium which is detected in sesame at doses useful to health (Ensminger AH *et al.*,1986)

Calcium, phosphorus and potassium are the most rich minerals in seeds of the different cultivars. Magnesium, which is an essential mineral for enzyme activity, is also found in significant amount in sesame seeds. Like calcium, magnesium plays a role in regulating acid-alkaline balance in the body, a significant role in photosynthesis. Also, the potassium is an essential nutrient and has an key role in the synthesis of amino acids and proteins (Fallon S. *et al.*, 2001)

CHAPTER III

MATERIALS AND METHODS

A field experiment entitled “Chemical composition of sesame (*sesamum indicum* L.) under different rates of micronutrients application ” was conducted at central farm (SAU), Dhaka during kharif-1 season 2016-2017, in order to assess the effect of foliar application of certain micronutrient names on the growth and yield of sesame. The materials used and the methods of different experimental techniques employed during the course of investigation are mentioned in the foregoing pages.

3.1 Location of the experimental site

The research work was carried out at the research field of Sher-e- Bangla Agricultural University Dhaka during the kharif-1 period from March to June 2017. The experimental field was located at 90° 33' E longitude and 23° 71' N latitude at a height of 9 m above the sea level. The land belongs to the Agro-ecological zone "Madhupur Tract' (AEZ-28) of Nodda soil series.

3.1.1 Soil

The soil of the experimental site was well drained and medium high. The physical and chemical properties of soil of the experimental site was examined prior to experimentation from 0-15 cm depth. The soil was clay loam in texture and having soil pH varied from 5.41-5.63. Organic matter content was very low (0.83%). The physical composition such as sand silt and clay content were 40%, 40% and 20%, respectively. The chemical properties of soil of experimental field have been presented in Table 1 and Table 2.

Table 1. Physical composition of the soil (0-15 cm)

Physical composition	Percentage composition (Air dry basis)	Methods adopted.
Sand coarse sand	51.3	Bouyoucos hydrometer method
Fine sand	30.2	
Silt	8.1	
Clay	10.3	
Texture class	Sandy loam	

Source: BARC, SRDI

Table 2. Chemical composition of the soil (0-15cm)

Chemical composition	Amount present (oven dry basis)	Amount present (oven dry basis)
pH	5.4	Digital PH meter with 1:2.5, soil: water
Organic carbon %	0.43	Walkley and black method
Available N (kg/ha)	204.5	Alkaline Potassium permanganate (KMO4) distillation method
Available P (kg/ha)	20.0	Brays – I method
Available K (kg/ha)	110.7	Flame photometer method

Source: BARC, SRDI

3.1.2 Climate

Sher-e-Bangla Agricultural University, Dhaka comes under the 28th agro-climatic region of the country where organic matter content and fertility level is low i.e. characterized by high temperature and heavy rainfall during kharif-1 season. The average annual rainfall of Dhaka is 1854 mm. out of which about 80% rainfall occurs from May to September and the rest is received within October to May. The average maximum temperature ranges from 27°-35°C during May to June while the minimum temperature varies from 15-18°C during December to January. The relative humidity varies between 50% in summer to 90% in rainy season. The details of the weather condition during the period of investigation from March to June 2017 have been presented in the Table 3.

Table 3. Meteorological data of Dhaka from March to June 2018

Year	Month	Monthly mean temp. in °C		Monthly mean Relative humidity		Total rainfall in mm	Sunshine hours
		Max.	Min.	Morning	Afternoon		
2017	March	32	20	83	63	65	7
2017	April	34	24	85	64	155	6
2017	May	33	25	87	65	340	5
2017	June	34	27	89	66	335	3

Source: Bangladesh meteorological department (<http://www.bmd.gov.bd/>)

3.1.3 Experimental details

Preparation of experimental plot

The experimental field was ploughed thoroughly earlier in the season and FYM was applied @ 5t/ha and was incorporated thoroughly in to the soil. Recommended dose of N P and K 30:20:20 kg/ha was applied to the experimental plots before sowing and mixed thoroughly. Half of N and all P and K were applied as basal dose and rest of N was applied as top dressing after first weeding and before flowering. Then the plots were laid out as per the plan.

REPLICATION-1	REPLICATION-2	REPLICATION-3
T ₁	T ₅	T ₄

T ₂	T ₈	T ₁
T ₃	T ₁₀	T ₃
T ₄	T ₂	T ₉
T ₅	T ₃	T ₆
T ₆	T ₁	T ₈
T ₇	T ₆	T ₅
T ₈	T ₉	T ₂
T ₉	T ₄	T ₇
T ₁₀	T ₇	T ₁₀

Figure 1. Experimental Plot Layout.

3.1.4 Design of experiment

The experiment was carried out in randomized block design with 12 treatments and replicated thrice.

Table 4. Experimental Design.

Design	RCBD
No. of Replication	Three
Plot size (Gross)	4.0m x 2.5m
Date of Sowing	20.03.2018

Spacing	30 cm. x 15 cm
Seed Rate	7 Kg/ha

3.1.5 Collection of seeds

The variety of sesame used for the present study was BARI Til-3. The seeds of this variety were collected from Bangladesh Agricultural Research Institute (BARI), Gazipur. Before sowing, the seeds were tested for germination in the laboratory and the percentage of germination was found to be over 90%. The important characteristics of this variety is mentioned below

3.1.6 Description of variety

BARI Til-3

Plants are of average 100-110 cm in medium height. Leaves are darker green and rough. Days to maturity medium (85 days). Stem is branched and contains 3-5 branches. The number of capsule per plant is 60-65 and seeds per capsule is 50-55. Maximum yield is 1200-1400 kg/ha. Seeds contain 42-50% oil and 25% protein. 1000 seed weight is 3.0 g.

3.1.7 Details of treatments

- T₁:** Controlled (RDF N P K 30:20:20 kg/ha)
- T₂:** T₁ + foliar spray of Boron @ 100ppm (Na₂B₄O₇.10H₂O)
- T₃:** T₁ + foliar spray of Zinc @ 100ppm as (ZnSO₄)
- T₄:** T₁ + foliar spray of Mo @ 50ppm as (NH₄Mo₇O₂₄. 4H₂O)
- T₅:** T₁ + foliar spray of Cu @ 100ppm as (CuSO₄.5H₂O)
- T₆:** T₁ + foliar spray of Fe @ 100ppm as (FeSO₄.7H₂O)
- T₇:** T₁ + foliar spray of Mn @ 100ppm as (MnSO₄.H₂O)

T₈: T₁ + foliar spray of Co @ 50ppm as (Co(NO₃)₂)

T₉: T₁ + Mixture of all above micronutrient

T₁₀: T₁ + foliar spray of Commercial micronutrient mixture (5 ml/2lt H₂O)

3.2 Intercultural operation

3.2.1 Thinning

To maintain optimum plant population in the experimental field, thinning was done after 15 days of seedling emergence to keep plant to plant distance 15 cm in each line.

3.2.2 Weeding

Weeding was done with thinning to keep the plots weed free through hand weeding. Generally, 3 weeding were adequate for this purpose and the second dose of nitrogen was top dressed after the last weeding.

3.2.3 Irrigation

Irrigation was given as when needed. From sowing till harvest six number of irrigation were applied to the crop.

3.2.4 Foliar application

Solutions of different concentrations of micronutrients as per treatments were prepared in deionized water and were sprayed twice at 45 and 60 DAS.

Table 5. Calendar of field operation

Date	Field operation
11.03.2017	Collection of soil sample
13.03.2017	Ploughing
17.03.2017	Application of FYM and Fertilizer
19.03.2017	Layout and leveling
20.03.2017	Sowing

21.03.2017	Irrigation
03.04.2017	2nd Irrigation
08. 04.2017	Thinning
09. 04.2017	Hoeing and weeding
10. 04.2017	Top dressing
11. 04.2017	Earthing Up
13. 04.2017	3rd irrigation
23. 04.2017	4th irrigation
05.05.2017	1st Foliar application
10.05.2017	Sampling of plant
13.05.2017	5th irrigation
20.05.2017	2nd Foliar application
27.05.2017	Sample collection
23.06.2017	6th irrigation
25.06.2017	Harvesting

3.3 Pre harvest observations

3.3.1 Sampling techniques

Plants were sampled from a unit area of 0.30 m² 5 no. of plants at random from each plot leaving sufficient border rows around it. The plant parts were separated and sun dried. Then dried in a hot air oven at 800 for 72 hrs. till constant weight. The dry weight of different plant parts were recorded.

3.3.2 Growth observations

The following observations were recorded at 45, 60 and 90 DAS.

3.3.2.1 Plant height

Height of the main shoot was measured from the base of the main shoot up to the tip of the top most leaf, averaged out and expressed in cm.

3.3.2.2 Number of branches per plant

Primary branches developing from the main shoot were considered as the criteria of observation.

3.3.2.3 Leaf area

Systronics leaf area meter model 211 was used for measuring leaf area. For this all the leaflets from each of the sample plants were detached and their area (one side) was recorded to compute leaf area per plant and expressed (cm²/plant).

3.4 Physiological growth parameters

3.4.1 Leaf Area Index (LAI)

LAI is expressed as the ratio of the leaf area (A) (only one side) to the ground area (P) occupied by the crop.

$$\mathbf{LAI = A / P}$$

3.4.2 Leaf Area Duration (LAD)

LAD is defined as the leaf area index integrated over time

$$LAD = \frac{A_1 + A_2}{2} \times (t_2 - t_1)$$

Where, A₁ = Leaf area index at the start of test period

A₂ = Leaf area index at the end of test period

t₂ - t₁ = Period in days between initial and final observation

3.4.3 Leaf area ratio (LAR)

LAR is defined as the ratio between leaf area (A) and total plant dry weight (W). It reflects the leafiness of a plant.

$$\mathbf{LWR = W_L / W}$$

3.4.4 Specific leaf weight (SLW)

SLW is the ratio between leaf dry weight (W_L) and leaf area (A)

$$SLW = W_L / A$$

3.4.5 Specific Leaf Area (SLA)

SLA is the ratio between leaf area (A) and leaf dry weight (W_L).

$$SLA = A / W_L$$

3.4.6 Relative Growth Rate (RGR)

Relative Growth Rate (RGR) is defined as the total plant dry weight increase in a time interval in relation to the initial weight or Dry matter increment per unit biomass per unit time or milligrams of dry weight increase per gram of dry weight increased per gram of original weight per unit time and expressed as unit dry weight / unit dry weight / unit time (mg/ g/day).

$$RGR = \frac{\ln W_2 - \ln W_1}{T_2 - T_1}$$

Where, W_1 = whole plant dry weight at t_1

W_2 = whole plant dry weight at t_2

$T_2 - T_1$ = time interval in days

3.4.7 Net Assimilation Rate (NAR)

NAR is defined as dry matter increment per unit leaf area per unit time and expressed as mg/cm²/day.

$$NAR = \frac{(W - W_1)}{(t_2 - t_1)} \times \frac{(\ln L_2 - \ln L_1)}{(L_2 - L_1)}$$

Where, W_1 = dry weight of whole plant at time t_1

W_2 = dry weight of whole plant at time t_2

L_1 = leaf area at t_1

L_2 = leaf area at t_2

$t_1 - t_2$ = time interval in days between initial and final observations

3.4.8 Crop Growth Rate (CGR)

The CGR explains the dry matter accumulated per unit land area per unit time ($\text{g/m}^2/\text{day}$).

$$\text{CGR} = \frac{(W_2 - W_1)}{\rho (t_2 - t_1)}$$

Where, W_1 = whole plant dry weight at time t_1

W_2 = whole plant dry weight at time t_2

$t_2 - t_1$ = time interval in days

ρ = ground area on which W_1 and W_2 were recorded.

CGR of a species are usually closely related to interception of solar radiation.

3.5 Chemical constituent

3.5.1 Estimation of chlorophyll content

Chlorophyll a, b and total chlorophyll content in the leaves were firm by using the method stated by Arnon (1949). The second leaf from the top was sampled for the purpose. The collected leaf samples were nearly kept in moist polythene bags to keep them turgid. Exactly 100 mg of fresh leaf was taken from the middle portion of the leaf and were cut into minor pieces. The leaf discs were then put in 80 % v/v acetone solution and kept in dark for 24 hours. Then the content was filtered by Whatman No.1 filter paper and the filtrate was used to record the absorbance (OD) at 645 nm and 663 nm. The respective chlorophyll content was calculated using the following formula and expressed as mg g^{-1} FW leaf.

$$\text{Chlorophyll-a} = (12.7 \times \text{OD}_{663} - 2.69 \times \text{OD}_{645}) \times \frac{V}{1000 \times W_f}$$

$$\text{Chlorophyll-b} = (22.9 \times \text{OD}_{645} - 4.68 \times \text{OD}_{663}) \times \frac{V}{1000 \times W_f}$$

$$\text{Total Chlorophyll} = (20.2 \times \text{OD}_{645} - 8.02 \times \text{OD}_{663}) \times \frac{V}{1000 \times W_f}$$

Where, OD_{645} = OD value at 645 nm

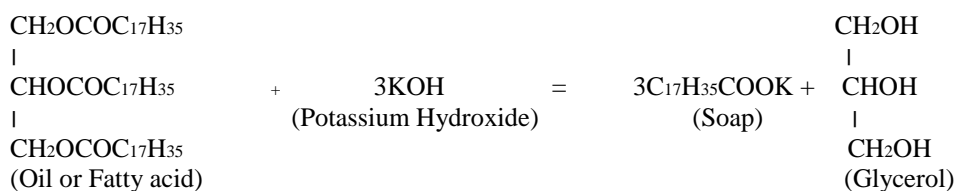
OD_{663} = OD value at 663 nm

V = Total volume of extract (ml)

W_f = Fresh weight of leaf (g)

3.5.2. Saponification value

Saponification value is the number of milligrams of KOH required to entirely saponify 1 g of oil. The method is based upon the principle that fat, on conduct with excess of alcoholic KOH is used up. The excess of KOH left unused may then be found by titrating it against a standard acid.



Reagents

1. Hydrochloric acid 0.5N
2. Alcoholic solution of Potassium hydroxide. 28 g potassium hydroxide are taken and dissolved it in very little water. Make up to one liter by adding rectified spirit ($\text{C}_2\text{H}_5\text{OH}$) of specific gravity 0.81.

Procedure

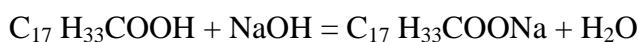
1. Weight accurately 2 g of fat in 250 ml conical flask. Add 25 ml of 0.5 N alcoholic potash solutions and fit the flask with a cork and a long air condenser.
2. Reflux the contents of the flask for about 30 minutes by heating on boiling water bath so that the contents just simmer. Cool the flask and add 1 ml of 1% solution of phenolphthalein and titrate the excess of the alkali against standard N/2 acid (a). At the same time and under similar conditions carry out a blank expt. (b) without fat (25 ml of the same alcoholic KOH heated in a similar way is titrated, against .05 N acid). 1 ml of 0.5 N HCL was equivalent to 0.02805g of KOH.

Calculation

$$\text{Saponification value} = \frac{(b-a) \times 0.02805 \times 1000}{\text{Wt. of substance in g}}$$

3.5.3 Acid value

Acid value of oil is determined by titration of a known weight of it against N/4 sodium hydroxide using phenolphthalein as the indicator.



Reagents

1. Phenolphthalein: 1 percent solution in alcohol neutralized with 0.1 N NaOH.
2. Denatured alcohol (Neutral): Mix 10 volumes of ethyl alcohol with 1 volume of methyl alcohol and neutralize with N/40 NaOH using phenolphthalein as indicator.

Procedure

Weigh 5-7 g of oil in 250 ml conical flask and add 50 ml denatured alcohol (neutral) and shake well. Now add 2 ml of phenolphthalein as indicator and titrate against N/4 NaOH with vigorous shaking after each addition till a permanent light pink color is produced which persists for at least 1 minute (a).

$$\text{Acid value} = \frac{a \times 0.00561}{\text{Wt. of oil}} \times 1000$$

3.5.4 Estimation of fatty acid composition

Seed sample of sesame were received from ORC, BARI, Joydebpur, Gazipur. Fatty acid composition was determined by Gas-liquid chromatographic method (Uppstrom *et al.*, 1978).

Reagent

1. Ethylate reagent (Petroleum ether / 0.02M sodium hydroxide in ethanol (2/3))
2. A Salt solution (80 g NaCl and 3g Sodium hydrogen Sulphate in 1 litre water)

Procedure

1. About 12 mg of oil or equivalent amount of oil seeds was taken (seed was crushed in an oil paper and then transferred into a test tube).
2. The sample was extracted and transesterified at the same time with 5 ml ethylated reagent and shaken.
3. The samples were kept for overnight at room temperature.
4. 10 ml salt solution was added and shaken. As soon as the two layers were separated, the benzene phase was transferred to small test tubes.
5. A Philips PU 4500 chromatograph instrument was used with flame ionization detector (FID).

).

3.5.5 Estimation of total protein content by Microkjeldhal method

The protein content of food stuff is obtained by estimating the nitrogen content of the material and multiplying the nitrogen value by 6.25 (according to the fact that nitrogen constitutes on average 16% of a protein molecule). This is referred to as crude protein content, since the non-protein nitrogen (NPN) present in the material is not taken in consideration. The estimation of nitrogen is done by Kjeldhal method (AOAC 1984) which depends upon the fact that organic nitrogen when digested with sulphuric acid in the presence of catalyst selenium oxide, mercury or copper sulfate is converted into ammonium sulphate. Ammonia liberated by making the solution alkaline is distilled into a known volume of a standard acid which is then back titrated.

Equipments

1. Balance
2. Microkjeldhal (Mkj) digestion set
3. Mkj distillation set.

Reagents

1. Digestion mixture: 100 g of potassium sulphate (K_2SO_4) was thoroughly mixed with 20 g of copper sulfate ($CuSO_4 \cdot 5H_2O$) and 2.5 g selenium dioxide (SeO_2) was added with it.
2. 60% Sodium hydroxide solution: 600 g sodium hydroxide and 50 g sodium thiosulphate were dissolved in distilled water, cooled and made the volume up to 1 liter.
3. Boric acid: 40 g of boric acid was dissolved in water and made up to 1 liter.
4. Double indicator: 200 mg each methyl red and bromocresol green was dissolved separately in 100 ml of 70% ethanol. One part of methyl red and five parts of bromocresol green were mixed before use.
5. Hydrochloric acid (0.02 N HCl): 8.5 ml concentrated hydrochloric acid was added to 5 liter of distilled water. Standardized to 0.02 N acids by titrating it against standard sodium carbonate (0.02 N) solution.

Procedure

A known quantity of the finely mustard ground sample (100 mg) weighted out in an Mkj digestion flask. About 2 g digestion mixture was added with it 2 ml of concentrated sulphuric acid was dispensed into the flask. Then it was digested for about 2 hrs in Mkj digestion set and was cooled the clear digest. The digest was dissolved in minimum amount of distilled water and carefully transferred to an Mkj distillation set. 10 ml of sodium hydroxide solution was added and distilled it. The distillate was collected for 5 min into 5 ml boric acid containing 2 drops of mixed indicator in a 50 ml conical flask, till the color of solution was changed. The distillate was titrated against a standard hydrochloric acid and noted the titer value (TV).

Calculation

$$N \% = \frac{(14.007) \times (\text{normality of the acid}, 0.02) \times (\text{TV})}{\text{Weight of sample (mg)}} \times 100$$

Where 14.007 is the equivalent weight of nitrogen. Nitrogen % is converted into protein by multiplying with a factor 6.25 for cereals and pulses.

Viscosity measurements

A rheometer as described by Nzikou *et al.* (2007) was used to measure the different oil viscosities. By this procedure, a concentric cylinder system is submerged in the oil and the force necessary to overcome the resistance of the viscosity to the rotation is measured. The viscosity value, in mPa.s, is automatically calculated on the basis of the speed and the geometry of the probe. Temperature (20°C) was controlled with a water bath connected to the rheometer. The experiment was carried out by putting 3 ml of sample in a concentric cylinder system using 100 sG1 as shear rate.

3.6. Estimation of Chemical content

3.6.1. Estimation of seed nitrogen content of different plant parts

Nitrogen content of different plant parts at harvest were estimated by following the procedure of A.O.A.C, (1970). 200mg of dry and powdered plant samples were taken in 100ml digestion tube. About 200mg of digestion mixture (K_2SO_4 : $CUSO_4$ = 5:1) and 4ml of concentrated sulphuric acid (H_2SO_4) were added. These tubes were kept as such for about 1hr and transferred the digestion tubes into digestion unit, then heated slowly till frothing occurred. To check the frothing, two crystals of sodium thiosulphate were added to each digestion tube. Thereafter, digestion was continued

until the contents of the tubes became completely cleared blue syrupy liquid without any bubbling. The tubes were cooled and 10 ml of distilled water was added to it and mixed thoroughly. The digestion tubes were cooled by running tap water over the outside surface of tube and transferred the content to 50 ml volumetric flask. The digestion tubes were rinsed with about 5 ml distilled water twice and transferred to the same volumetric flask and volume was made up to 50 ml and kept it for sample analysis. 10ml of diluted sample extract was transferred in to a microkjeldahl distillation unit.

Calculation

$$\% \text{ of nitrogen in the sample} = \frac{(\text{sample titer} - \text{blank titer}) \times N \text{ of HCL} \times 14 \times 100 \times 5}{\text{Sample wt. (g)} \times 1000}$$

3.6.2 Estimation of phosphorus and potassium contents of plant sample

Digestion of plant sample for determination of phosphorus and potassium

1g powdered plant sample was taken in 100ml conical flask separately. To each flask 10ml of ternary acid mixture ($\text{HNO}_3 + \text{H}_2\text{SO}_4 + \text{HClO}_4 = 5:1:2$) was added and pre-digested slowly for 15 minutes on a hotplate. Gradually heat was increased and digestion was continued till content became clear and white fumes encircle inside the flask. Then content was cooled and 30 ml distilled water was added. The contents were transferred to 50ml volumetric flask and volume was made up to 50ml with distilled water and the aliquot was filtered through whatmann no. 1 filter paper. The filtrate was kept for the estimation of phosphorus and potassium present in plant sample.

3.6.3 Estimation of phosphorus present in plant sample

Phosphorus present in plant sample was estimated by adopting the procedure of Jackson 1973.

Sample analysis

Standards of 0, 2, 4, 6, 8, 10 and 12ml of 25ppm phosphorus solution and 2ml of digested sample extracts were taken in 25ml volumetric flask separately. 5ml of 2N HNO_3 solution was added to each flask. Then required amount of distilled water was

added to each flask to make the final volume 15 ml. thereafter 2.5 ml molybdate vanadate solution was added. Final volume was made up to 25 ml with distilled water and flasks were shaken well thoroughly. Absorbance was measured by a spectrophotometer at 420 nm after 20 mins of shaking. The phosphorus content of plant sample was calculated in percentage by using the standard curve.

3.6.4 Estimation of potassium present in plant sample

1ml of digested sample extract was taken in a 25 ml volumetric flask and the volume was adjusted to 25 ml with distilled water. Similarly 1,2,3,4,5 ppm standard solution were prepared by diluting 1,2,3,4,5 ml of 100 ppm of K solution (i.e 0.1907g of KCL per litre) in 100ml volumetric flask with water. The readings for standards and samples were taken in a digital flame photometer. As per the standard curve, the ppm of potassium present in the extracted solution was calculated. Then the percentage of potassium presents in plant and seed samples were estimated.

3.6.5 Estimation of oil content

The oil percentage in seed sample was determined by soxhlet apparatus. 2g of crushed seed was taken in a thimble put in a soxhlet extraction apparatus connected with a pre weight extraction flask containing 250ml petroleum ether. Then the flask was heated for 1hr the solvent petroleum ether circulated through the condenser of soxhlet apparatus over the entire refluxing period. Then the receiving flask was detached and again attached with another condenser to recollect the pure petroleum ether in another flask. The remained petroleum ether with oil in the previous flask was heated to make the oil free from petroleum ether. Then the weight of the extraction flask with oil was taken and the % of oil content was determined.

Calculation

$$\text{Oil content (\%)} = \frac{W_0}{W_s} \times 100$$

W_0 = weight of oil extracted in grams

W_s = weight of seed in grams

3.7 Post harvest studies

3.7.1 Capsules per plant

Matured capsule from ten sample plants were detached, counted and averaged per plant.

3.7.2 Shelling percent

The weight of seed obtained from 100 gm dry pods from each treatments replication wise was recorded to calculate shelling %.

3.7.3 Thousand seed weight

One thousand seeds were selected randomly replication wise from each treatment and the weight was taken.

3.7.4 Seed yield

Seeds were collected from 10 samples and weight recorded to compute seed yield per plant. Plants were cut from base at maturity from each plot and sundried. Seeds were collected after threshing and seed yield per plant and per hectare was calculated.

3.7.5 Harvest index

$$\text{HI} = \frac{\text{Seed yield}}{\text{Total dry matter of plant}} \times 100$$

3.8. Statistical analysis

Analysis of variance

The data collected from the experiment on various aspects of growth, yield and yield attributing characters of groundnut were arranged in appropriate tables according to the treatments and were subjected to statistical analysis appropriate to the design (panse and sukhatme, 1985). Values represented are the means and standard

deviations for three replicates. Statistical analysis was carried out by Excel Version 8.0 software. Significance was defined at $P < 0.05$. The treatment variations were tested for significance of F test. The standard error of mean {SE (m)} and critical difference (CD) at 5% were calculated as

$$SE (m) = \sqrt{(EMS \div r)}$$

Where EMS = error mean sum of square

R = number of replication

$$CD (0.05) = SE (m) \times \sqrt{2} \times t (0.05) \text{ at error d.f.}$$

CHAPTER IV

RESULTS AND DISCUSSION

Field experiment was conducted at central farm SAU, Dhaka, during Kharif-1 2017 with 10 treatments replicated thrice in a Randomized Block Design to study the

“Chemical composition of sesame (*Sesamum indicum* L.) under different rates of micronutrients application”. The morphological, physiological and biochemical observations taken at various growth stages during the course of investigation were recorded, statistically analyzed and presented in this chapter with appropriate tables and figures.

4.1 Morphological Growth Parameters

4.1.1 Plant height and branches per plant

Plant height and branches per plant recorded at 45, 60 and 90 DAS (table 6) revealed that height and number of branches increased with age of the plant up to 90 DAS. Irrespective of treatments maximum rate of elongation and formation of branches occurred during 45 to 60 DAS as compared to 60 to 90 DAS.

Plant height varied among the treatments at all the 3 stages of crop growth. Foliar application of micronutrients significantly increased the plant height which varied a minimum height of 61, 91.3 and 95.2 cm in control (T₁) to a maximum of 76.9, 108.8 and 109.1 cm in (T₉) at 45, 60 and 90 DAS respectively. Foliar application of all the micronutrients in combination (T₉) registered the highest plant height followed by foliar application of Zn (T₃), B (T₂), and Mo (T₄) among the treatments at all the growth stages. Application of micronutrients as foliar spray significantly increased number of branches per plant as recorded at different growth stages (table 6). Minimum number of branches per plant was recorded in control (T₁) and the maximum number of branches per plant was recorded in combined application of micronutrients (T₉). Foliar spray of all the micronutrients alone or in combination as well as their commercial mixture significantly increased number of branches over control except application of Cu (T₅). Significantly the highest number of branches (8) per plant was recorded in plants sprayed with combined micronutrients (T₉) followed by Zn (T₃), B (T₂) and Mo (T₅)

Table 6. Effects of different micronutrients on plant height (cm) and number of branches per plant at different growth stages.

Treatment	Plant height (cm)			No. of branches /plant		
	45DAS	60DAS	90DAS	45DAS	60DAS	90DAS
T1	61.0	91.3	95.2	2.0	3.3	4.0
T2	70.6	104.5	106.3	4.6	6.6	7.3
T3	73.5	104.9	107.1	5.3	6.0	7.3
T4	69.3	101.4	106.0	4.0	5.3	6.6
T5	62.9	95.5	96.7	2.6	4.0	6.0
T6	66.4	96.7	98.5	3.3	5.3	6.0
T7	68.1	100.1	100.5	4.0	4.6	6.6
T8	67.0	98.8	99.0	3.3	4.6	6.0
T9	76.9	108.8	109.2	6.0	8.0	8.0
T10	66.2	101.4	102.6	4.0	5.3	7.3
SE(m)±	0.586	0.749	1.499	0.225	0.307	0.372
LSD(0.5)	1.94	2.82	4.85	0.87	0.99	1.36

4.1.2 Dry matter accumulation and its partitioning

Dry matter accumulation and its partitioning in to different plant parts were recorded at 45, 60, and 90 DAS were presented in table 7. Data revealed that there was concomitant increase in total dry matter (TDM) accumulation till 90 DAS but it's partitioning to different parts like leaf, stem and pod increased up to 60 DAS but at 90 DAS partitioning of TDM was less to the leaf compared to stem and pod.

Foliar application of all the micronutrients and their combination significantly increased leaf dry matter (LDM), stem dry matter (SDM), pod dry matter (PDM) and total dry matter (TDM) of the plant recorded at 45, 60 and 90 DAS over control except foliar application of Cu (T₅), Fe (T₆) which found at par with control with respect to (TDM) and its partitioning to different plant parts at various growth stages. Significantly the highest accumulation of dry matter in leaf , stem, pod as well as in whole plant recorded at all the growth stages in plant applied with foliar spray of micronutrients in combination (T₉) followed by foliar application of Zn (T₃) and B (T₂). The pattern of partitioning of TDM at 45 and 60 DAS are same i.e. stem > leaf > pod whereas pattern of partitioning of TDM was in a sequence of pod > stem > leaf at 90 DAS.

4.2 PHYSIOLOGICAL TRAITS

4.2.1 LAI, SLA, SLW and LAR

Growth parameters like LAI, SLA, SLW and LAR determined at 45 and 60 DAS were presented in table-8. Data indicated that higher value of LAI and SLW were found at 60 DAS where as higher value of SLA and LAR were noticed at 45 DAS in all the treatments.

Leaf area index (LAI) increased significantly due to foliar application of micronutrients which was ranged a minimum of 0.91 (T₁) to a maximum of 1.35 (T₉) at 45 DAS and 1.08 (T₁) to 2.24 (T₉) at 60 DAS. In both the stages (45 and 60 DAS) significantly the highest LAI was recorded in plants sprayed with combined micronutrients (T₉) followed by application of Zn (T₃), B (T₂), Mo (T₄) and commercial mixture (T₁₀).

Table 7. Effect of different micronutrients on Dry matter accumulation (g/plant) and its partitioning.

Treatments	45DAS				60 DAS				90DAS			
	LDM	SDM	PDM	TDM	LDM	SDM	PDM	TDM	LDM	SDM	PDM	TDM
T1	0.95	0.90	0.08	1.95	1.93	3.08	2.80	7.81	1.71	4.31	4.42	10.44
T2	1.51	1.68	0.32	3.52	3.27	5.50	5.50	14.27	2.95	7.01	10.27	20.23
T3	1.59	1.69	0.25	3.54	3.90	5.56	5.30	14.76	3.73	7.63	10.14	21.50
T4	1.49	1.64	0.22	3.37	3.12	4.57	4.40	12.09	2.83	5.88	8.21	16.92
T5	1.09	1.11	0.10	2.30	2.11	3.97	3.30	9.38	1.96	4.37	5.90	12.23
T6	1.27	1.23	0.11	2.62	2.67	4.13	3.50	10.30	1.77	4.79	6.59	13.15
T7	1.33	1.50	0.19	3.03	2.83	4.31	3.60	10.74	2.56	5.50	7.63	15.69
T8	1.32	1.48	0.12	2.93	2.80	4.29	3.50	10.59	2.64	5.18	7.14	14.96
T9	1.92	1.72	0.36	4.01	4.26	7.16	6.90	18.32	4.09	8.40	13.40	25.89
T10	1.49	1.55	0.20	3.24	2.88	4.34	4.30	11.52	2.77	5.85	7.59	16.21
SE(m)±	0.014	0.003	0.015	0.183	0.171	0.268	0.303	0.690	0.158	0.341	0.479	0.979
LSD(0.5)	0.04	0.008	0.04	0.54	0.50	0.79	0.90	2.05	0.47	1.01	1.42	2.90

Table 8. Effect of different micronutrients on LAI, SLA, SLW, LAR at different growth stages

Treatments	LAI		SLA(cm ² /g)		SLW(mg/cm ²)		LAR(cm ² /g)	
	45DAS	60 DAS	45DAS	60 DAS	45DAS	60 DAS	45DAS	60 DAS
T1	0.91	1.08	428.8	252.8	2.33	3.95	210.4	62.4
T2	1.08	1.77	324.2	244.4	3.08	4.09	139.0	56.0
T3	1.13	2.10	319.0	243.2	3.13	4.11	143.7	64.2
T4	1.09	1.65	329.2	239.1	3.03	4.18	146.4	61.7
T5	0.98	1.33	405.0	284.3	2.46	3.51	191.6	63.9
T6	1.00	1.38	353.6	233.6	2.82	4.28	171.7	60.5
T7	1.06	1.63	360.1	259.7	2.77	3.84	158.3	68.4
T8	0.98	1.32	335.3	213.0	2.98	4.69	151.7	56.3
T9	1.35	2.24	317.3	237.4	3.15	4.21	151.9	55.2
T10	1.07	1.64	321.9	257.0	3.10	3.89	148.2	64.2
SE(m)±	0.052	0.053	0.641	0.606	0.082	0.001	0.475	3.382
LSD(0.5)	0.15	0.15	1.90	1.79	0.24	0.002	1.41	NS

Specific leaf area (SLA) was recorded at 45 and 60 DAS differed significantly among the treatments. At 45 DAS the highest SLA 428.8 cm²/g was recorded in control (T₁) and lowest 317.3 cm²/g in T₉ where as at 60 DAS the SLA was highest 284.3 cm²/g in T₅ and the lowest 213 cm²/g recorded in T₈. Though significant difference in SLA was observed among the treatments in both the stages but no definite trend in influence of micronutrient spray on SLA was observed at both stages.

Specific leaf weight (SLW) recorded at 45 and 60 DAS differed significantly among the treatments. Foliar application of most of the micronutrients alone or in combination significantly increased SLW over control. At 45 DAS the maximum SLW 3.15 mg/cm² recorded in plants applied with foliar spray of combined micronutrients (T₉) followed by Zn (T₃), commercial mixture (T₁₀) and B (T₂) where as at 60 DAS, the highest SLW 4.69 mg/cm² was recorded in Co (T₈) followed by Fe (T₆) and T₉. The lowest value of SLW was recorded in control (T₁) at 45 DAS and in Fe (T₅) at 60 DAS.

4.2.2 RGR, NAR, CGR and LAD

RGR, NAR, CGR and LAD determined between 45 and 60 DAS were depicted in table-9. Relative growth rate (RGR) increased with foliar spray of most of the micronutrients over control but significant increase was recorded (101.3 mg/g/day) in combined micronutrients sprayed (T₉) followed by Zn (T₃). Net assimilation rate (NAR) significantly influenced by foliar application of micronutrients. The significant increase in NAR over control was observed in case of foliar spray of all the micronutrients except Mn (T₇) which was at par with control. The highest NAR 1.2 mg/cm²/day was recorded in T₉ followed by B (T₂) and Zn (T₃).

Table 9. Effect of different micronutrients on RGR, NAR, CGR and LAD at different growth period

Treatments	RGR(mg/gm/day)	NAR(mg/cm ² /day)	CGR(g/m ² /day)	LAD
	45-60 DAS	45-60 DAS	45-60 DAS	45-60 DAS
T1	92.5	0.87	8.7	14.97
T2	93.2	1.13	15.9	21.49
T3	95.0	1.05	16.6	24.31
T4	85.2	0.95	12.9	20.66
T5	93.5	0.91	10.5	17.37
T6	91.2	0.96	11.4	17.90
T7	84.4	0.85	11.4	20.24
T8	85.5	0.98	11.3	17.36
T9	101.3	1.20	21.1	27.01
T10	84.4	0.91	12.2	20.36
SE(m)±	0.763	0.001	0.760	0.596
LSD(0.5)	2.26	0.002	2.58	1.77

Crop growth rate (CGR) was significantly increased by application of different micronutrients as foliar spray. Among the treatments the maximum CGR 21.1 g/m²/day was registered in plants applied with combined micronutrients (T₉) followed by Zn (T₃), B (T₂) and Mo (T₄). The lowest CGR was recorded in control plant (T₁) where no micronutrient was applied.

Leaf area duration (LAD) was influenced significantly by foliar application of micronutrients. Among the treatments the highest LAD 27.01 was registered in T₉ followed by application of Zn (24.31), B (21.49), and Mo (20.66). The lowest LAD was recorded in control plant without micronutrients (T₁).

4.2.3 Chlorophyll index and total chlorophyll content

Chlorophyll index and total chlorophyll content of leaf were recorded at 45 and 60 DAS presented in table 10. The data revealed that, the chlorophyll index and total chlorophyll content of leaf were more at 60 than 45 DAS.

The chlorophyll index differed significantly between control and micronutrient applied plants. The index varied with the lowest value of 8.16 (T₁) to highest value of 13.39 (T₉) at 45 DAS and lowest value of 8.99 (T₁) to highest value of 17.74 (T₉) at 60 DAS among the treatments. Foliar application of all the micronutrients alone or in combination as well as their commercial mixture significantly increased the chlorophyll index over control except application of Cu (T₅), which was at par with control. Among the treatments, the highest chlorophyll index was recorded in T₉ followed by Zn (T₃), B (T₂), and Mo (T₄) at both of the growth stages.

Like chlorophyll index similar trend was also observed for total chlorophyll content of leaf at both 45 and 60 DAS, which varied with the lowest value of 1.27 and 1.61 mg/g fresh wt. in T₁ and highest value of 1.67 and 1.93 mg/g fresh wt. in T₉ at 45 and 60 DAS respectively. Almost all the treatments of micronutrients showed significantly greater chlorophyll content over control except application of Cu (T₅) which was found at par with control. Among the treatments, T₉ showed the maximum chlorophyll content followed by Zn (T₃), B (T₂) and Mo (T₄) at 45 and 60 DAS.

Table 10. Effect of different micronutrients on chlorophyll index and total chlorophyll content at different growth stages

Treatments	chlorophyll index		total chlorophyll content(mg/g fresh weight)	
	45 DAS	60 DAS	45 DAS	60 DAS
T₁	8.16	8.99	1.27	1.61
T₂	12.91	16.6	1.57	1.90
T₃	13.19	17.2	1.65	1.91
T₄	11.94	14.58	1.56	1.86
T₅	9.54	10.11	1.35	1.69
T₆	10.83	13.11	1.48	1.79
T₇	11.15	13.33	1.49	1.85
T₈	10.22	12.06	1.47	1.76
T₉	13.39	17.74	1.67	1.93
T₁₀	11.16	14.21	1.53	1.85
SE(m)±	0.653	0.548	0.047	0.003
LSD(0.5)	1.93	1.62	0.14	0.009

4.3 Uptake of N, P, K in Plant

4.3.1 N Content and Uptake

N content in different plant parts and its uptake were estimated at harvest and presented in table 11. N content (%) in various plant parts varied among the treatments and ranged a maximum of 1.4 to a minimum of 0.52 % in leaf, 0.17 to 0.10 % in stem and 1.96 to 1.08 % in seed. Among the treatments the highest N content was recorded in T₉ and lowest in T₁. Irrespective of treatments N content in different plant parts were found in the order of seed N content >leaf>stem. Significant increase in N content was observed in all the plant parts with foliar application of micronutrients over control except in seed where application of Cu (T₅) and Mn (T₇) were at par with control. Among the treatments foliar application of combined micronutrients (T₉) registered significantly the highest N content in leaf, stem and seed followed by B (T₂), commercial mixture (T₁₀) , Zn (T₃) and Mo (T₄) in that order.

N uptake by plant and plant parts differed among the treatments and varied from a minimum of 8.97, 4.52, 32.67,46.1 mg in (T₁) to a maximum of 57.26, 14.7, 125.6 and 197.5 mg in (T₉) by leaf, stem, seed and whole plant respectively. The result indicated that partitioning of N uptake by plant was in the order of uptake in seed > leaf > stem. The lowest uptake of N was recorded in T₁ (control) while all the foliar application of micronutrients increased N uptake significantly. Among all the treatments T₉ registered the highest N uptake followed by Zn (T₃), B (T₂), commercial mixture (T₁₀) in leaf, Zn(T₃), B(T₂), Mo(T₄) in stem, B(T₂), commercial mixture (T₁₀), Zn (T₃) in seed and B(T₂), Zn (T₃) and commercial mixture (T₁₀) in whole plant respectively.

4.3.2 P content and uptake

The P content and uptake by plant parts and whole plant at harvest were presented in table 12. Irrespective of treatments, P content in different plant parts were found in the order of seed P content > leaf > stem. P content (%) in various plant parts varied among the treatments and ranging from 0.37 to 0.48 % in leaf, 0.23 to 0.34 % in stem and 0.49 to 0.57% in seed. The data revealed that there was significant increase in P content of

Table 11. Effect of different micronutrients on Nitrogen content and uptake

Treatments	Nitrogen content (%)			Nitrogen uptake(mg/plant)			
	Leaf	Stem	Seed	Leaf	Stem	Seed	Total
T₁	0.52	0.10	1.08	8.97	4.52	32.67	46.1
T₂	1.22	0.15	1.36	36.13	10.51	80.20	126.8
T₃	0.98	0.14	1.36	36.55	10.68	74.14	121.3
T₄	0.87	0.14	1.36	24.76	8.23	69.51	102.5
T₅	1.01	0.15	1.22	19.89	6.55	48.32	74.7
T₆	1.15	0.16	1.36	20.44	7.66	55.16	83.2
T₇	0.94	0.15	1.19	24.19	8.25	52.70	85.1
T₈	1.05	0.14	1.29	27.72	7.25	53.41	88.3
T₉	1.4	0.17	1.96	57.26	14.7	125.62	197.5
T₁₀	1.05	0.14	1.68	29.08	8.19	77.73	115.0
SE(m)±	0.059	0.008	0.072	1.741	0.496	3.884	6.102
LSD(0.5)	0.17	0.02	0.21	5.17	1.47	11.53	18.12

stem over control by foliar application of all micronutrients except Co (T₈) and combined micronutrients (T₉). Whereas, P content in leaf and seed were found in significant among the treatments.

P uptake by plant parts and whole plant showed variation among the treatments at harvest. The P uptake varied from 7.52 to 16.98 mg in leaf, 10.33 to 29.82 mg in stem, 15.82 to 32.86 mg in seed, and 33.67 to 69.23 mg in whole plant. The result indicated that P uptake by plant partitioned in order of P uptake by seed > stem > leaf. Foliar application of all the micronutrients and its combination significantly increased P uptake in leaf over control except Cu (T₅) and Fe (T₆). Among the treatments, the highest uptake of P in leaf was recorded by application of Zn (16.98mg) followed by T₉ (15.31mg), T₂ (14.21mg), T₄ (13.11mg). Foliar application of all the micronutrients significantly increased P uptake in stem over control. Significantly the highest P uptake was recorded 29.82mg in T₂ followed by T₃, T₄, T₉ and T₁₀ which were at par among themselves. P uptake by seed and whole plant showed a similar trend in response to foliar application of micronutrients. Maximum P uptake by seed (32.86 mg) and (69.23 mg) per plant in T₉ which followed closely by T₂, T₃ and T₄.

4.3.3 K content and uptake

The K content (%) and uptake by different plant parts and whole plant were presented in table 13. The data revealed that the K content varied from 1.06 to 1.31% in leaf, 1.75 to 1.97% in stem and 0.25 to 0.34% in seed, which indicated that K content in stem > leaf > seed. Among the treatments the highest K leaf content 1.31% was recorded in T₈ and the lowest in T₇. However no significant change in leaf K content was observed among the treatments. Significant increase in K content of stem was recorded due to foliar application of B (T₂), Fe (T₆), Cu (T₅) and combination of micronutrients (T₉) over control but the rest of the treatments found non-significant as compared to control. K content in seed didn't differed significantly over control however the highest K content was recorded in T₁₀ followed by T₉ and the least was recorded in T₃.

The uptake of K by plant parts and whole plant (table-13) revealed that it varied from a minimum of 19.23mg (T₁) to maximum of 53.17 mg (T₉) in leaf, 75.4mg (T₁) to 157.5 mg (T₉) in stem, 9.03mg(T₁) to 21.63mg (T₉) in seed and 103.6mg (T₁) to 232.3

mg (T₉) in whole plant. This indicated that the partitioning of K uptake among the plant parts in the sequence of K uptake in stem > leaf > seed. All the foliar application of micronutrients enhanced K uptake in different plant parts as well as per plant as compared to control. Foliar application of all the micronutrients in combination (T₉) registered the highest K uptake followed by Zn (T₃), B (T₂) and commercial mixture (T₁₀) in leaf, B (T₂), Zn (T₃), Mo (T₄) in stem, commercial mixture (T₁₀), Mo (T₄) and B (T₂) in seed and Zn (T₃), B (T₂) and Mo (T₄) in whole plant. However foliar application of Cu (T₅) and Fe (T₆) didn't register any significant change in K uptake per plant as well as in leaf.

Table 12. Effect of different micronutrients on Phosphorous content and uptake

Treatments	Phosphorous content (%)			Phosphorous uptake(mg/plant)			
	Leaf	Stem	Seed	Leaf	Stem	Seed	Total
T1	0.44	0.23	0.52	7.52	10.33	15.82	33.67
T2	0.48	0.34	0.51	14.21	29.82	30.44	68.31
T3	0.45	0.29	0.52	16.98	22.13	28.74	67.86
T4	0.46	0.36	0.56	13.11	21.10	28.82	63.04
T5	0.46	0.33	0.53	8.99	14.26	21.05	44.31
T6	0.46	0.28	0.55	8.25	13.69	22.14	44.10
T7	0.37	0.36	0.56	9.51	20.34	24.82	54.68
T8	0.37	0.24	0.57	9.79	12.93	23.66	46.39
T9	0.37	0.25	0.51	15.31	21.05	32.86	69.23
T10	0.45	0.34	0.49	12.66	19.76	22.80	55.23
SE(m)±	0.024	0.018	0.025	0.672	1.061	1.478	3.098
LSD(0.5)	NS	0.05	NS	1.99	3.15	4.39	9.20

Table 13. Effect of different micronutrients on Potassium content and uptake

Treatments	Potassium content (%)			Potassium uptake (mg/plant)			
	Leaf	Stem	Seed	Leaf	Stem	Seed	Total
T₁	1.12	1.75	0.30	19.23	75.4	9.03	103.6
T₂	1.28	1.97	0.25	37.98	138.4	14.68	191.1
T₃	1.26	1.75	0.25	47.09	133.5	13.57	194.2
T₄	1.18	1.81	0.30	33.60	106.5	15.59	155.7
T₅	1.25	1.87	0.26	24.50	81.9	10.25	116.7
T₆	1.21	1.97	0.29	21.46	94.6	11.72	127.8
T₇	1.06	1.83	0.28	27.20	101.0	12.40	140.7
T₈	1.31	1.76	0.30	34.65	91.2	12.37	138.3
T₉	1.30	1.87	0.33	53.17	157.5	21.63	232.3
T₁₀	1.25	1.77	0.34	34.62	103.8	15.73	154.2
SE(m)_±	0.082	0.040	0.017	1.921	6.199	0.789	9.081
LSD(0.5)	NS	0.11	NS	5.70	18.17	2.34	26.98

4.4 Yield, Yield Attributes and Oil Content

Yield, yield attributes and oil content were depicted in table 14. Number of capsules per plant differed significantly among the treatments. It varied from a minimum of 25.1 to maximum 35.8 capsules per plant among the treatments. Foliar application of all the micronutrients significantly increased the number of capsules per plant over control except Cu (T₅) and Fe (T₆) whereas highest number of capsules was found in (T₉) followed by B(T₂), Zn(T₃) and Mo(T₄).

Shelling percent varied among the treatments. The highest shelling percent (52.16%) was recorded in T₉ and the lowest (31.85%) in T₁ among the treatments. Foliar application of all the micronutrients showed significant increase in shelling percent over control except Mo (T₄) and Cu (T₅).

Number of seeds per capsule varied among the treatments and ranged a minimum of 45.8 to a maximum of 55.6. Foliar application of all the micronutrients alone or in combination as well as their commercial mixture significantly increased the number of seeds per capsule over control except application of Cu (T₅). Among all the treatments the highest seeds per capsule was obtained in (T₉) followed by B (T₂), Zn (T₃) and Mo (T₄). Variation in seed size was noticed among the treatments. The highest thousand seed weight (3.22g) and the lowest (2.62g) were recorded in T₉ and T₁ respectively. However no significant difference was observed among the rest treatments with respect to thousand seed weight

Table 14. Effect of different micronutrients on seed yield and its components

Treatments	No. of capsule/plant	Shelling (%)	No. of seeds/capsule	1000 seed weight(g)	Seed yield /plant(g)	Yield kg/ha.	HI (%)	Oil (%)
T1	25.1	31.85	45.8	2.62	3.01	662.6	28.84	43.2
T2	35.2	42.80	52	3.21	5.87	1292.6	29.03	48.6
T3	33	46.43	51.6	3.19	5.43	1195.0	25.26	49.3
T4	31.9	37.97	50.2	3.18	5.09	1120.3	30.09	47.4
T5	26.2	33.13	47.8	3.15	3.94	867.8	32.25	44.6
T6	26.4	38.67	48.6	3.15	4.04	889.1	30.73	45.1
T7	28.0	42.00	49.9	3.17	4.42	974.4	28.21	45.3
T8	26.7	42.22	49.2	3.14	4.12	907.4	27.57	44.9
T9	35.8	52.16	55.6	3.22	6.40	1410.0	24.75	50.2
T10	29.1	39.03	50	3.18	4.62	1017.9	28.54	46.5
SE(m)±	0.533	2.254	0.876	0.182	0.274	60.377	1.576	0.837
LSD(0.5)	1.58	6.69	2.60	NS	0.81	179.36	NS	2.48

Seed yield per plant varied among the treatments. Foliar application of micronutrients significantly increased seed yield per plant over control. The highest seed yield (6.40g/plant) was recorded in T₉ followed by T₂ (5.87g), T₃ (5.43g), T₄ (5.09g) per plant. Seed yield per hectare showed similar trend as it was found in seed yield per plant. The highest seed yield 1410 kg per hectare was obtained by combined micronutrients (T₉) which registered 112% higher yield followed by 95% for B (T₂), 80% for Zn (T₃) and 69% for Mo (T₄) over control (T₁).

Harvest index varied in the range between 32.25 and 24.75 but no significant variation was found among the treatments. Variation in oil content was observed among the treatments. Foliar application of micronutrients in combination (T₉) registered the highest oil content (50.2%) followed by Zn (49.3%), B (48.6%), Mo (47.4%) and commercial mixture (46.5%) which were significantly higher over control. However no significant difference in oil content was observed in rest of the treatments.

4.5 CHEMICAL COMPOSITION

Table 15. Proximate composition (%) of various parts of *Sesamum indicum*.

Nutrients	Leaves	Seed	Root	Whole parts
Moisture	5.04 ± 3.37 b	5.60 ± 3.74 b	6.60 ± 4.39 a	4.22 ± 2.81 c
Crude fiber	9.72 ± 6.48 b	6.60 ± 4.39 ac	12.80 ± 8.53 a	8.80 ± 5.87 b
Carbohydrates	56.37 ± 37.59 c	62.23±41.49 ab	67.90 ± 45.26 a	58.86 ± 38.94 b
Crude protein	19.25 ± 12.83 b	16.63 ± 11.09 c	7.88 ± 5.25 d	21.44 ± 14.29 a
Ash	9.62 ± 6.41 a	8.94 ± 0.96 a	4.82 ± 3.22 c	6.68 ± 4.45 b
Fats and Oil	3.88 ± 2.59 b	38.54 ± 25.69 a	2.66 ± 1.77 c	4.54 ± 3.03 b
Dry matter wt.	94.96 ± 63.31 a	94.40 ± 62.93 a	93.40 ± 64.94 a	95.78 ± 68.85 a
Food energy value	337.4±224.93bc	337.40±224.93 bc	327.06 ± 218.1 c	362.06 ± 241.37 a

Test values carrying different letters within a column are significantly different ($P > 0.05$) using the F- test. Data are expressed as mean ± SD (n = 3).

The proximate analysis data of the leaves, seeds, roots and the whole plant are presented in Table 15. The moisture content of root (6.60%) was significantly different to other parts of the plant while that of the whole plant (4.22%) showed the least content. Both the leaves (5.04%) and the seeds (5.60%) compared well with each other.

Table 16. Mineral compositions (%) of various parts of *Sesamum indicum*.

Minerals	Leaves	Seed	Root	Whole parts
Sodium	0.48 ± 0.32 a	0.23 ± 0.23 a	0.33 ± 0.22 a	0.33 ± 0.22 a
phosphorus	0.49 ± 0.38 b	0.55 ± 0.37 a	0.20 ± 0.14 c	0.20 ± 0.14 c
Calcium	1.41 ± 0.94 b	0.90 ± 0.60 c	1.71 ± 1.14 ab	1.71 ± 1.14 ab
Potassium	0.90 ± 0.64 b	1.20 ± 0.80 a	1.03 ± 0.69 a	1.03 ± 0.69 a
Magnesium	0.49 ± 0.38 b	0.24 ± 0.16 c	0.61 ± 0.40 a	0.61 ± 0.40 a

Test values carrying different letters within a column are significantly different ($P > 0.05$) using the F-test. Data are expressed as mean ± SD (n = 3).

The difference in the concentration of the various elements within the different parts may be attributed to preferential absorbability of the plant for the corresponding elements. In addition, the observed results may be linked to the minerals composition of the soil in which the plant grows and climatology conditions. In leaves, seed, root and whole parts sodium concentration have good. Other mineral composition is at various results. Calcium present on root (1.71%) and whole parts (1.71%).

The present investigation under taken in field condition to study the response of *Sesamum* crop to foliar application of different micronutrients, viz. B, Zn, Mo, Cu, Fe, Mn, Co and their combination as well as commercial mixture. The data relating to various characters studied during course of investigation and interpreted in the preceding chapter were discussed in this chapter under the following heads.

4.6 Morphological growth

Plant height and number of branches which are considered important morphological traits of plant growth were significantly influenced by foliar application of different micronutrients applied either individually or in combination presented in table-4. Among all the treatments the height was maximum 77,108 and 109 cm at 45 ,60 and 90 DAS in case of combined micronutrients (T₉) followed by Zn (T₃), B (T₂), Mo

(T₄). *Sesamum* plant applied with all the micronutrients and their combination produced more number of branches as compared to control but significantly a highest number of branches (8/plant) were found in T₉ followed by T₃, T₂ and T₄.

Foliar spray of micronutrients improve metabolic function of plant causing more vegetative growth which manifested increase in height and more number of branches as reported by Ibrahim *et al.*, (2007) in bean and Yadav *et al.*, (2009) in *Sesamum* supported the present finding.

4.6.1 Dry matter accumulation

Dry matter accumulation and its partitioning into different plant parts were influenced by foliar application of micronutrients (table 7). A gradual increase in TDM accumulation was observed up to 90 DAS but the pattern of its partitioning to different plant parts differed at various growth stages. Similar pattern of partitioning of TDM was found at 45 and 60 DAS, which was in the sequence of stem dry matter >leaf>pod, whereas at 90 DAS the TDM partitioning was in the order of pod dry matter>stem> leaf.

Among all the treatments the highest TDM in plant and its plant part like leaf, stem and pod were found in combined micronutrient (T₉) followed by Zn (T₃), B (T₂) and Mo (T₄). Similar increase in dry matter of plant and its plant parts due to foliar application of micronutrients have been reported in *Sesamum* by Salwa *et al.*, (2010) and Yadav *et al.*, (2009) which was corroborated the present result.

4.6.2 Physiological growth parameters

Growth analysis has the advantage of giving integrated measurement of net photosynthetic activity over wide range of condition that reveal in the field. Potential productivity is controlled by the efficient use of solar energy and photosynthetic rate per unit area. Leaf is the ultimate source for increasing biomass production and their translocation through transport network to the sink for such reasons desirable canopy structure must be established. This can be achieved by developing higher leaf area and rapid leaf area development as leaf is the ultimate source. Various physiological parameters related to canopy structure would always be useful which not only through light on crop productivity, but also their manipulation enhances potential productivity. In this regard the role of micronutrients in crop physiology is undisputable to harness

the possibility of such manipulation for achieving higher productivity in different oil seed crops, *Sesamum* in particular.

The present findings in table-6 revealed that there was significant variation among the treatments with respect to LAI, SLA, SLW, LAR determined at 45 and 60 DAS of *Sesamum* plant. In all the treatments, higher value of LAI and SLW found at 60 DAS whereas, SLA and LAR were more at 45 DAS. High value of SLA and lower value of SLW at 45 DAS as compared to 60 DAS in all the treatments indicated a high rate of leaf expansion occurred at 45 DAS beyond that period thickness of leaf was more than its expansion.

Foliar application of different micronutrients and their combination showed higher value of LAI over control both at 45 and 60 DAS. Among the treatments, the highest LAI was found in plants with combined micronutrients (T₉) followed by Zn (T₃), B (T₂), Mo (T₄) and commercial mixture (T₁₀). Similarly the highest SLW (3.15 mg/cm²) was recorded for T₉ at 45 DAS followed by T₃, T₁₀ and T₂ but at 60 DAS the highest SLW (4.69 mg/cm²) was found with application of Co (T₈) followed by T₆ and T₉. Maximum value of SLA and LAR were found in control plant T₁ as compared to plants applied with micronutrients. This might be due to low accumulation of dry matter in leaf in control plants as compared to other treatments. Micronutrients play critical role in plant that lead to increase of LAI and there by increased light absorption and increase the amount of dry matter accumulation and economic yield (Rabi *et al.*, 2008) supported the present findings with respect to influence of micronutrients on LAI.

In plant micronutrients are involved in various physiological function and there by increased LAI, CGR, RGR, NAR leading to the development and productivity of plant in soybean (Gupta and Vyas 1994) and LAD by (Cakmak 1999) were in agreement with the result of present investigation which recorded significant increase in all the above physiological growth parameters by foliar spray of individual micronutrients like Zn, B, Mo and their combination over control.

4.6.3 Chlorophyll index and total chlorophyll content

Chlorophyll index measured by speedometer and total chlorophyll content of leaves showed significant variation among the treatments at both 45 and 60 DAS (table-8). Foliar application of individual micronutrients, their combination and commercial mixture significantly influenced chlorophyll index as well as total chlorophyll in leaves of *Sesamum* plant over control. Among the treatments, the highest chlorophyll index was found 13.39 and 17.74 at 45 and 60 DAS respectively in plants applied with combined micronutrients (T₉) followed by Zn (T₃), B (T₂), Mo (T₄), commercial mixture (T₁₀), Mn (T₇) and Fe (T₆). Similarly total chlorophyll was highest 1.67 and 1.93 mg/g fresh weight at 45 and 60 DAS respectively in T₉ followed by T₃, T₂, T₄, T₁₀, T₇ and T₆.

Fe plays essential role in the chlorophyll synthesis (Sultana *et al.*, 2001), Zn may be required for chlorophyll production (Pandey *et al.*, 2006). Combined application of B and Zn is effective to increase chlorophyll content, N, P and K content in dry matter and seed and oil protein content in seeds of sunflower (Gitte *et al.*, 2005) were in agreement with the present findings.

4.7 Nutrient uptake by plant and plant parts

In order to know the effect of micronutrients application on major nutrient status in sesame plant N, P and K content and their uptake by plant and plant parts were estimated at harvest.

4.7.1 N content and uptake

Nitrogen concentration varied 0.52 to 1.4 in leaf, 0.1 to 0.17 in stem and 0.08 to 1.96 in seed among the treatments (table-9). It indicated that irrespective of treatments, concentration of N in different plant parts was found in the sequence of N content of seed > leaf > stem. Significant increase in N content was observed in all the plant parts due to foliar application of micronutrients (table-9). Among the treatments, combined micronutrients (T₉) registered the highest concentration of N in leaf, stem and seed followed by application of B (T₂), commercial mixture (T₁₀), Zn (T₃) and Mo (T₄). Similarly N uptake by plant and plant parts significantly influenced by foliar application of micronutrients. Among all the treatments, highest uptake of N by plant

and plant parts was found in T₉ followed by T₃ and T₂. The result indicated that partitioning of N uptake by plant was in the order of N uptake by seed > leaf > stem.

4.7.2 P content and uptake

P concentration varied in the range from 0.37 to 0.48 % in leaf 0.23 to 0.34% in stem and 0.49 to 0.57 % in seed among the treatments (table-10), which indicated that concentration of P was maximum in seed and minimum in stem. However no much difference in P concentration of different plant parts was found among the treatments. Uptake of P by plant and plant parts significantly influenced by application of micronutrients over control. Unlike P content partitioning of P uptake of plant in to its plant part was found maximum uptake by seed and minimum by leaf. Among the treatments maximum uptake of P in leaf (16.98 mg) was found due to Zn application, in stem (29.82 mg) due to B but maximum P uptake (32.86 mg) in seed and 69.23 mg/plant was found due to combined application of all micronutrients (T₉) followed closely by T₂, T₃ and T₄. The uptake of P by plant and plant parts was the least in control (T₁).

4.7.3 K content and uptake

Irrespective of the treatments K concentration was maximum in stem and minimum in seed. Foliar application of micronutrients did not show much difference in K content of different plant parts over control except in stem (table-11), where significant increase in K content was found due to foliar spray of B (T₂), Fe (T₆), Cu (T₅), combination of micronutrients (T₉) over control. In contrast K uptake by plant and plant parts significantly influenced by application of micronutrients. The highest uptake of K in plant and plant parts was recorded in T₉ followed by T₃ and T₂ and the least uptake was recorded in T₁. Similar results have been found by (Gette et al 2005) who reported that combined application of B and Zn increase N, P and K content in dry matter in seed. Increased absorption of N, P and K in sesame due to foliar application of Zn (Tirupati *et al.*, 2001). Increase uptake of N, P and K in seed and

stem due to combined application of Zn and Mn to sesame crop (Singervel *et al.*, 2002).

4.8 Yield and yield attributes

Yield of sesame is associated with various yields attributing characters such as number of seeds per capsule, capsules per plant, shelling %, seed weight. The product of these components determines the seed yield per plant, per hectare which very often influence by different factors. In this experiment the effect of different micronutrients spray on yield and yield attributes have been studied (table-12).

Number of seeds per capsule and number of capsule per plant were influenced significantly by foliar application of different micronutrients to sesame plant. Among all the treatments, T₉ (combined application of micronutrients) was the best which produced maximum number of capsule per plant and seeds per capsule followed by T₂, T₃ and T₄. Whereas Cu (T₅) and Fe (T₆) for number of capsule per plant and T₅ for seeds per capsule did not show any significant change over control.

Shelling % increased significantly over control by foliar application of all the micronutrients except Mo (T₄) and Cu (T₅). The highest shelling % (52.16%) was found due to combined application of micronutrients (T₉) followed by T₃, T₂ and T₈.

Seed size measured in terms of 1000 seed weight found maximum (3.22 g) in T₉ and minimum (2.62 g) in T₁. Significant difference in seed size was only observed between T₁ and T₉ but the rest of the treatments did not show any significant difference as compared to control.

Seed yield per plant as well as per hectare showed similar trend among the treatments. Both were increased significantly by the foliar application of micronutrients over control. Whereas highest seed yield 6.4 g per plant and 1410 kg/ha were found due to combine application of micronutrients (T₉) followed by B (T₂), Zn (T₃), Mo (T₄). Which registered an increase in the seed yield to the tune of 112%, 95%, 80% and 69% respectively over control but harvest index (HI) did not show any significant difference among the treatments.

Foliar application of micronutrients increased oil content in seed of *Sesamum*. Among the treatments, the highest oil content (50.2%) was found in plant applied with micronutrients in combination (T₉) followed by Zn (T₃), B (T₂), Mo (T₄) and commercial mixture (T₁₀) which were found significantly higher than control. But the rest of the treatments did not increase oil content significantly over control. Increase in seed yield and seed quality of *Sesamum* due to combined application of micronutrients was reported by (Salwa et al 2010). Micronutrients application significantly increased yield and yield components viz. number of capsules per plant and number of seeds per capsule. The beneficial influence of micronutrients might be due to the activation of various enzymes and the efficient utilization of applied nutrients resulting in increased yield components (Tiwari et al 2000, Shankar *et al.* 1999) are in accordance with the present findings.

Various parts of sesame and mineral composition

The good distribution of nutrients in the leaves may explain the usage of this plant as a forage feed given to the domestic animals. The roots had the highest content of crude fibre ($12.80 \pm 8.53\%$) and carbohydrate ($67.90 \pm 45.26\%$). Crude protein was highest ($21.44 \pm 14.29\%$) in whole plant followed by the leaves ($19.25 \pm 12.83\%$) but lowest in the roots ($7.88 \pm 5.25\%$). The high concentrations of proteins suggest that the whole plant of *S. indicum* can contribute to the daily protein need of 21.44 g for adults near 23.6 g recommended by the National Research Council (1975). These values corroborate with the protein content (21.44) found in non-Nigerian benniseed within the range of 18.00 to 23.18% as reported by Dashak and Fali (1993). The seeds have the maximum oil contents (38.54%), lowest in roots (2.66%). The observed results support the usefulness of sesame seeds as a good source of edible oils which can be used in cooking as well as soap manufacturing industry. The data revealed high food energy value in the seed (662.3%) which may be due to high lipids content. There was no significant different in dry matter weight (93.40-95.78%) of various plant parts investigated in this study. The proximate analysis of the ash contents was significantly high in the leaves ($9.62 \pm 6.41\%$), followed by the seed ($8.94 \pm 5.96\%$), whole plants ($6.68 \pm 4.45\%$) while that of the roots ($4.82 \pm 3.22\%$) was very low. The values of ash and crude fibre content are important in terms of the suitability of food vegetable and digestibility (Bowmen and Russell 2001). The values of crude fibre

contents of this plant as recommended by National Research Council (1975) may contribute to the daily needs of fibre. Recently, the interest in dietary fibre has been stimulated due to its ability to prevent chronic diseases such as cardiovascular disease, cancer and diabetes mellitus (Bowmen and Russell, 2001) to bowel irritation and possibly colon cancer.

The difference in the concentration of the various elements within the different parts may be attributed to preferential absorbability of the plant for the corresponding elements. In addition, the observed results may be linked to the minerals composition of the soil in which the plant grows and climatology conditions. The functional roles of these elements are well documented but must be taken appropriately to meet the daily requirement. Data obtained on elemental concentrations of various parts of *S. indicum* may be useful in deciding the dosage or quantity for consumption as well as herbal remedies of this plant.

CHAPTER V

SUMMARY AND CONCLUSION

A field experiment was conducted during Kharif-1 2017 at central farm SAU, Dhaka to study the “Chemical composition of sesame (*Sesamum indicum* L.) under different rates of micronutrients application”. The experiment was laid out in a Randomized Block Design with twelve treatments and replicated thrice. The treatments were T₁: control (no spray of micronutrients), T₂: B (100 ppm), T₃: Zn (100 ppm), T₄: Mo (50 ppm), T₅: Cu (100 ppm), T₆: Fe (100 ppm), T₇: Mn (100 ppm), T₈: Co (50 ppm), T₉: (combination of above micronutrients) and T₁₀: commercial mixture of micronutrients. Micronutrient treatments were given to the crop at 45 and 60 DAS. Notes were recorded on growth characters, chlorophyll content, yield and yield attributes, N, P and K uptake and oil content of sesame seed during the course of investigation. After statistical analysis of the data, the salient findings are summarized as follows.

- ❖ Plant height of *sesamum* significantly improved by foliar application of different micronutrients as recorded at 45, 60 and 90 DAS. Among the treatments plant height was maximum due to foliar application of joined micronutrients (T₉) at all the growth stages followed by Zn (T₃), B (T₂), and Mo (T₄).
- ❖ Number of branches per plant increased pointedly over control by foliar treatment of micronutrients except Cu (T₅). Among the treatments the highest number of branches produced in plant due to application of combined micronutrients (T₉) followed by Zn (T₃), B (T₂) and Mo (T₄).

- ❖ Significant increase in dry matter accumulation in plant and plant parts due to micronutrient application was found over control at 45, 60 and 90 DAS. Meaningfully the highest accumulation of dry matter in leaf, stem, pod and whole plant at all the three stages was recorded due to application of combined micronutrients T₉ followed by application of Zn and B.
- ❖ LAI increased significantly due to foliar application of micronutrients. At both the stages of 45 and 60 DAS the highest LAI was found in T₉ followed by T₃, T₂ and T₄ over control.
- ❖ SLA was recorded at 45 and 60 DAS differed significantly among the treatments. At 45 DAS the highest SLA was in T₁ and lowest in T₉. However at 60 DAS the SLA was highest in T₅ and the lowest was in T₈. No definite trend was observed amidst the treatments at both the growth stages.
- ❖ SLW recorded at 45 and 60 DAS differed significantly among the treatments. Foliar application of most of the micronutrients alone or in amalgamation significantly increased SLW over control. At 45 DAS maximum SLW was found in combined micronutrients followed by Zn, commercial mixture and B. whereas, at 60 DAS the highest SLW was recorded in Co followed by Fe and combined micronutrients.
- ❖ LAR recorded at 45 and 60 DAS varied among the treatments. The highest and lowest LAR were recorded in T₁ and T₂ respectively at 45 DAS, whereas highest and lowest LAR were recorded in T₇ and T₉ at 60 DAS. No definite influence of micronutrient spray on LAR was observed.
- ❖ RGR increased with foliar spray of micronutrients over control but major increase was noted in combined application of micronutrients (T₉) followed by Zn (T₃).
- ❖ NAR significantly increased by foliar application of all the micronutrients over control excluding Mn (T₇). The highest NAR was recorded in combined micronutrients (T₉) followed by B (T₂) and Zn (T₃).

- ❖ CGR was significantly increased by all the micronutrient treatments over control except Cu (T₅). Among all the treatments the determined CGR was found in T₉ followed by T₃, T₂ and T₄.
- ❖ LAD was increased significantly by foliar application of micronutrients over control. Among all the treatments, the highest LAD was registered in T₉ followed by application of Zn, B and Mo.
- ❖ Chlorophyll index as well as total chlorophyll content of leaf were more at 60 DAS than 45 DAS. Chlorophyll index and total chlorophyll content significantly improved by all the treatments of micronutrients over control except Cu (T₅). Among the conducts the highest value was found in T₉ followed by T₃, T₂ and T₄. Influence of micronutrients on chlorophyll content exposed similar trend at both 45 and 60 DAS.
- ❖ Nitrogen content and uptake of plant and plant parts significantly prejudiced by foliar application of micronutrients. N content and uptake in different plant parts were found in the order of seed > leaf > stem. Among the treatments the highest N content was recorded in T₉ followed by T₂, T₁₀, T₃ and T₄. Among the treatments T₉ registered the highest N uptake followed by T₃, T₂ and T₁₀ in leaf, T₃, T₂ and T₄ in stem, T₂, T₁₀ and T₃ in seed and T₂, T₃ and T₁₀ in whole plant respectively.
- ❖ Irrespective of treatments, P content in dissimilar plant parts were found in the order of seed > leaf > stem. The micronutrients effect on P content was found significant only in stem and non-significant in leaf and seed. The significant increase was found in P content in stem over control by foliar submission of all the micronutrients except T₈ and T₉. Foliar application of all the micronutrients and its combination significantly increased P uptake in leaf over control except Cu and Fe. The highest uptake of P in leaf was found in Zn followed by T₉, T₂ and T₄. The highest P uptake of stem was found in T₂ followed by T₃, T₄ and T₉ and in both the seed and total plant the P uptake was highest in T₉ followed by T₂, T₃ and T₄.

- ❖ Potassium content varied among the plant parts by foliar submission of micronutrients. K content in plant parts was found in the order of stem > leaf > seed. Application of K in leaf and seed did not diverged significantly among the treatments whereas micronutrient treatments significantly inclined K content in stem. The highest K content was found in T₂ followed by T₆, T₅ and T₉ in stem. K uptake enlarged significantly by foliar application of micronutrient. Partitioning of K uptake into plant parts was in the order of stem > leaf > seed. All the foliar application of micronutrients heightened K uptake in different plant parts and per plant as compared to control. Foliar application of all the micronutrients in mixture (T₉) registered the highest K uptake followed by T₃, T₂ and T₄ in leaf, T₂, T₃ and T₄ in stem, T₁₀, T₄ and T₂ in seed and T₃, T₂ and T₄ in whole plant. However foliar application of Cu (T₅) and Fe (T₆) did not register any momentous change in K uptake per plant as well as in leaf.
- ❖ All the micronutrients significantly increased number of capsule per plant over control except Cu and Fe. Among all the treatments highest number of capsules produced per plant due to application of combined micronutrients followed by T₂, T₃ and T₄.
- ❖ Shelling % significantly influenced by foliar submission of micronutrients except T₄ and T₅. Among the treatments the highest shelling % was recorded in T₉ and the lowest was in T₁.
- ❖ Number of seeds per capsule pointedly increased over control by all the micronutrient treatments except Cu (T₅). The highest number of seeds per capsule was obtained in T₉ followed by T₂, T₃ and T₄.
- ❖ Thousand seed weight did not differ significantly among the treatments but highest 1000 seed weight was found in T₉ and lowest in T₁.
- ❖ Seed yield per plant as well as yield per hectare significantly increased by foliar application of micronutrients. Among the treatments foliar application of mutual micronutrients gave highest yield per plant and per hectare and recorded a gain in yield by 112% followed by application of B (95%), Zn (80%) and Mo (69%) over control.

- ❖ Foliar application of micronutrients in combination (T₉) registered the maximum oil content followed by Zn (T₃), B (T₂), Mo (T₄) and commercial mixture (T₁₀), which were significantly higher over control. But the rest of the treatments found insignificant compared to control.

CONCLUSION

In the present study, it was apparent that foliar application of different micronutrients each alone or their combination heightened most of the morpho-physiological traits (plant height, number of branches, capsules per plant, RGR, NAR, CGR and total dry matter accumulation per plant) and bio chemical attributes (chlorophyll content, N, P and K uptake and oil content, biochemical structure), seed yield and yield features over control. Application of micronutrients in combination (T₉) excelled over rest of the treatments in respect of seed yield and oil content. Further application of specific micronutrients like B, Zn and Mo also performed well for growing seed yield and oil content in sesamum next to combined micronutrient treatment. Keeping in view of the above beneficial effect of micronutrient application, from the present effort, it was concluded that foliar spray of individual micronutrients like B, Zn, Mo, Fe, Mn, Cu and Co or their combination can be used as supplementary application laterally with the normal dose of N, P and K in order to improve the productivity of oil seed crop in general and *sesamum* in particular.

CHAPTER VI

REFERENCES

- Arnon DI. 1949. Copper enzymes in isolated chloroplast polyphenol oxidase in Beta Vulgaris. *Plant physiology*.**24**:1-15.
- Abdul-Hamid A, Luan YS. Functional properties of dietary fiber prepared from defatted rice bran. *Food chemistry*. 2000; 15-19.
- Asad A, Blamey FPC and Edwards DG. 2003. Effects of boron foliar applications on vegetative and reproductive growth of sunflower, *Annals of Botany*, **92**: 565-570.
- Alvarez-Fernandez A, Abadia J and Abadia A. 2006. Iron deficiency, fruit yield and quality. In: Barton LL, Abadia J (eds) Iron nutrition in plants and rizospheric microorganisms. Springer, Dordrecht, The Netherlands: 85–101.
- Ahmad W, Niaz A, Kanwal S, Rahmatullah and Khalid R. 2009. Role of boron in plant growth: a review, *Journals of Agricultural Research*, **47**(3): 329-334.
- Al- Bamarny SFA, Salman MA and Ibrahim ZR. 2010. Effect of NAA, KNO₃ and Fe on some characteristics of leaf and fruit of peach (*Prunus persica* L.) cv. early coronet. "World Food System – A Contribution from Europe". Tropentag, September 14 - 16, 2010 in Zurich.
- Bowmen BA, Russel RM. 2001. *Present Knowledge in Nutrition* (8th Edn), International Life Sciences Institute, USA, pp 43-104.
- Babaeian M, Piri I, Tavassoli A, Esmailian Y and Gholami H. 2011. Effect of water stress and micronutrients (Fe, Zn and Mn) on chlorophyll fluorescence, leaf chlorophyll content and sunflower nutrient uptake in Sistan region, *African Journal of Agricultural Research*, **6**(15): 3526-3531.

- Bakkaus E, Gouget B, Gallien JP, Khodja H, Carrot F, Morel JL and Collins R. 2005. Concentration and distribution of cobalt in higher plants: The use of micro-PIXE spectroscopy. *Nucl. Instr. Meth. Phys. Res. B*, **231**: 350-356.
- Balakrishnan V. 2005. Effect of micronutrients on flower yield and xanthophylls content of African marigold (*Tagetes erecta* L.) M. Sc., Thesis submitted to Tamil Nadu Agricultural University, Coimbatore.
- Bilen S, Bilen M and Bardhan S. 2011. The effect of boron management on soil microbial population and enzyme activities, *African Journal of Biotechnology*, **10**(27): 5311-5319.
- Buzetti S, Muraoka T and Sa ME. 1990. Boron rates in soybean under different soil acidity conditions. Dry matter and grain yields and critical level in soil, *Revista Brasileira de Ciencia do Sob*, **14**(2): 157-161.
- BBS. 2017. Bangladesh Bureau of Statistics, Statistical Yearbook of Bangladesh, Statistics Division, Ministry of Planning, GOB.
- Cakmak I, Kalayci M, Ekis H, Brauni J, Kilinc Y and Yilmaz A. 1999. Zn deficiency as a practical problem in Plant and human nutrition in Turkey: a NATO science for stability project, *Field Crop Research*, **60**: 175-188.
- Cakmak I. 2008. Enrichment of cereal grains with zinc: Agronomic or genetic biofortification, *Plant Soil*, **302**: 1-17.
- Cambráia J, Pires DLFJ, Estavo MDM and Oliva MA. 1983. Effects of aluminium on the levels of magnesium, iron, manganese and copper in sorghum, *Rev. Ceres*, **30**: 45-54.
- Chaplot PC, Jain GL and Bansal KN. 1992. Effect of Sulphur and Phosphorus on sesame and its residual effect on wheat, *Indian Journal of Agronomy*, **6**: 167 – 169.
- Dashak DA, Fali CN (1993) Chemical composition of four varieties of Nigeria benniseed. *Food Chemistry* 47, 253-255.
- Dehnavi MM, Mohammad SA, and Sanavy M. 2008. Effects of withholding irrigation and foliar application of zinc and manganese on fatty acid composition and seed oil content in winter safflower, 7th International safflower conference, Wagga Wagga Australia.
- Devarajan R and Palaniappan SP. 1995. Zinc and molybdenum on yield and nutrition of soybean, *The Madras Agricultural Journal*, **82**(3):188-189.
- Devi KN, Singh LNK, Singh MS, Singh SB and Singh KK. 2012. Influence of Sulphur and Boron Fertilization on Yield, Quality, Nutrient Uptake and Economics of Soybean (*Glycine max*) under Upland Conditions, *Journal of Agricultural Science*, **4**(4): 1-10.

- Dhakshinamoorthy M and Krishnamoorthy KK. 1989. Studies the effect of zinc and copper on yield and quality of brinjal, *Madras agricultural journal*, **76** (16): 345- 347.
- Ensminger AH, Ensminger MK. Food for Health: A Nutrition Encyclopaedia Clovis, California: Pegus Press; 1986.
- El-Nagdy GA, Dalia MAN, Eman AE and Gelan SAEL. 2010. Response of flax plant (*Linum usitatissimum* L) to treatments with mineral and Bio-fertilizers from Nitrogen and Phosphorus, *Journal of American science* **6**:10.
- Fallon S, Enig MG. Nourishing Traditions. The Cookbook that Challenges Politically Correct Nutrition and the Diet Dictocrats. Revised 2 Ed., 2001. p: 40-45.
- Gupta PK and Vyas KK. 1994. Effect of phosphorus, zinc and molybdenum on yield and quality of soybean, *Legume Research*, **17**(1): 5-7.
- Hatwar GP, Gondane SU, Urkude SM and Gahukar OV. 2003. Effect of micronutrients on growth and yield of chilli, *Soils and Crops*, **13**: 123-125.
- Heitholt JJ, Sloan JJ and Mackown CT. 2002. Copper, manganese and zinc fertilization effects on growth of soybean on a calcareous soil, *Journal of Plant Nutrition*, **25**: 1727-1740.
- Hemantaranjan A, Trivedi AK and Mariram. 2000. Effect of foliar applied boron and soil applied iron and sulphur on growth and yield of soybean (*Glycine max* L. Merr.), *Indian Journal of Plant Physiology*, **5**(2): 142-144.
- Hu YS, Ma YH, Sun YL & Guo G. 1994. Effect of B application on the agronomic traits, yields and oil contents of a double-row rape (*Brassica napus* L.) cultivar, *Oil Crops (China)*, **16**: 43-46.
- Huang C, Garker SJ, Langridge P, Smith FW and Graham RD. 2000. Zinc deficiency upregulates expression of high affinity phosphate transporter genes in both phosphate sufficient and deficient barley roots, *Plant Physiology*, **124**:415 – 422.
- Ibrahim ME, Bekheta MA, A.El-Moursi and Gafar NA. 2007. Improvement of growth and seed yield quality of *Vicia faba* L. plants as affected by application of some bioregulators, *Australian Journal of Basic and Applied Science*, **1**(4): 657-666.
- Jauhari S, Srivastava R and Srivastava PC. 2005. Effect of Zinc on growth, flowering, corm attributes, post harvest life, leaf and corm nutrients status in gladiolus cv. Red Beauty. *Prog. Horticulture*, **37**(2): 423-428.
- Jackon ML.1973: Soil Chemical analysis. Pentice Hall of India Pvt.Ltd. New Delhi.
- Kaya C and Higgs D. 2002. Response of tomato (*Lycopersicon esculentum* L.) cultivars to foliar application of zinc when grown in sand culture at low zinc, *Scientia Horticulturae*, **93**:53-64.

- Kobraee S, Shamsi K and Rasekhi B. 2011. Effect of micronutrients application on yield and yield components of soybean. *Annals Biological Research*, **2**(2): 476-482.
- Kohnaward P, Jalilian J and Pirzad A. 2012. Effect of foliar application of micronutrients on yield and yield components of safflower under conventional and ecological cropping systems, *International Research Journal of Applied and Basic Sciences*, **3**(7): 1460-1469.
- Liu P, Yang Y and Yang YA. 2003. Effect of molybdenum and boron on quality of soybean, *Scientia Agricultura Sinica*, **36**(2): 184-189.
- Marschner H. 1998. Mineral Nutrition of Higher Plants. Harcourt Brace & Company. Publishers, London, New York, Tokyo.
- Mortvedt JJ, Murphy LS, Follett RH. 1999. Fertilizer Technology and Application. Meister Publishing, Willoughby, OH.
- Mosavi SR, Galavi M and Ahmadvand M. 2007. Effect of zinc and manganese foliar application on yield, quality and enrichment on potato (*Solanum tuberosum* L.), *Asian Journal of Plant Science*, **6**(8): 1256-1260.
- Mousa MA, EAE and Zedan ZS. 2010. Effect of nitrogen fertilizers and some micro nutrient on flax yield and chemical composition characters, *Journal of Plant Production, Mansura University* : 2090-3677.
- Moussavi-Nik M, Rengel Z, Hollam G J and Ascher J. 1997. Seed manganese content is more important than Mn fertilization for wheat growth under Mn deficient conditions, *Plant Nut for Sustain Food Product and Environ*, **4**:267-268.
- Mondal, D.K., Sounda, G., Panda, P.K., Ghosh, P., Maitra, S. and Roy, D.K. 1997. Effect of different irrigation levels and nitrogen doses on growth and yield of sesame (*Sesamum indicum* L.). *Indian Agriculturist*. **41**(1): 15-21.
- Narimani H, Rahimi MM, Ahmadikhah A and Vaezi B. 2010. Study on the effects of foliar spray of micronutrient on yield and yield components of durum wheat. *Archieves of Applied Science Research*, **2**(6): 168-176.
- Nath MR and Biswas J. 2002. Studies on effect of boron on vegetative and reproductive growth in tuberose (*Polianthes tuberosa* L.) cv. Single, *The Orissa Journal of Horticulture*, **30**(2): 39-42.
- Prakash V, Nandi PK. Association-dissociation behavior of sesame alpha globulin in electrolyte solutions. *Journal of Biological chemistry*. 1975; 252: 240-243.
- Rangarajan N, Ambilly M. Phytochemical screening of *Sesamum indicum* seed extract. *World journal of pharmacy and pharmaceutical sciences*. 2012; 1: 1298-1308.
- Pable D and Patil DB. 2011. Effect of sulphur and zinc on nutrient uptake and yield of soybean, *International Journal of Agricultural Sciences*, **7**(1): 129-132.

- Pandey N, Pathak GC and Sharma CP. 2006. Zinc is critically required for pollen function and fertilisation in lentil, *Journal of Trace Elements in Medicine and Biology*, **20**: 89-96.
- Potarzycki J and Grzebisz W. 2009. Effect of zinc foliar application on grain yield of maize and its yielding components, *Plant Soil Environment*, **55**(12): 519-527.
- Rajput DK, Megh Naresh, Srivastava PC, Singh SK and Gangawar MS. 2003. Effect of S, B and Zn application and their interactions on growth, yield and nutrients uptake of *Tagetes minuta* L. *Indian Perfumers*, **47**(1): 91-97.
- Ramamoorthy K and Sudarsan S. 1992. Supply of zinc and boron on yield and seed quality in groundnut. *Annals of Plant Physiology*, **6**: 33-38.
- Rabi S, Channal HT, Hebsur NS, Patil BN and Dharmatti PR. 2008. Effect of sulphur, zinc and iron nutrition on growth, yield, nutrient uptake and quality of safflower (*Carthamus tinctorius* L.), *Karnataka Journal of Agricultural Science*, **21**(3): 382-385.
- Renukadevi A and Savithri P. 2003. Sunflower (*Helianthus annuus* L.) oil yield and quality as influenced by boron application, *Madras Agricultural Journal*, **91**: 74-76.
- Sakal R, Sin RB and Singh AP. 1994. Effect of boron application on black gram and chickpea production in calcareous soil, *Fertilizer News*, **37**(5):31-32.
- Salwa AI, Mohsen E, Abass M and Behary SS. 2010. Amelioration productivity of sandy soil by using amino acids, sulphur and micronutrients for sesame production, *Journal of American Science*, **6**(11):250-257.
- Schlegel H, Godbold DL and Hutchinson A. 1987. Whole plant aspects of heavy metal induced changes in CO₂ uptake and water relations of spruce (*Picea abies*) seedlings, *Physiology Plantarum*, **69**: 265-270.
- Sepehr E, Malakouti MJ and Rasouli MH. 2002. The effect of K, Mg, S and micronutrients on the yield and quality of sunflower in Iran. 17th WCSS. 14-21 August 2002, Thailand.
- Shanker H, Bhushan C and Lallu L. 1999. Effect of levels of Zn on growth, dry matter and yield of sesame varieties, *Journal Oil Seeds Research*, **16**(1):74-77.
- Shehu HE. 2014. Manganese and Zinc nutrition of Sesame (*Sesamum indicum*) in Mubi, Northern Guinea Savannah Zone of Nigeria, *International Research Journal of Agricultural Science and Soil Science*, **4**(8): 159-166.
- Silva CAT, Cagol A, Silva TRB and Nobrega LHP. 2011. Boron application before sowing of sunflower hybrid, *Journal of Food Agriculture and Environment*, **9**: 580-583.
- Sing AK and Sing CS. 1996. Response of tomato to boron application. *Punjab Horticulture Journal*, **34** (1-2): 86-89.

- Singaravel R, Imayavaramban V, Thanunathan K and Shanmughapriya V. 2002. Effect of micronutrients on yield and nutrient uptake of sesame (*Sesamum indicum* L.) in a Vertisol soil, *Sesame and safflower Newsletter*, **17**:46-48.
- Sultana N, Iked T and Kashem MA. 2001. Effect of foliar spray of nutrient solutions on photosynthesis, dry matter accumulation and yield in sea water-stressed rice, *Environmental and Experimental Botany*, **46**: 129-140.
- Talashilkar SC and Chavan AS. 1996. Response of groundnut to calcium, sulphur and boron with and without FYM on Inceptisol, *Journal of Indian Society Soil Science*, **44**(2): 343-344.
- Thalooth AT, Tawfik MM and Magda MH. 2006. A comparative study on the effect of foliar application of zinc, potassium and magnesium on growth yield and some chemical constituents of mung-bean plants grown under water Stress Conditions, *World Journal of Agricultural Science*, **2**(1): 37-46.
- Thirupathi MK, Thanunathan K, Prakashand M and Imayavaramban V. 2001. Use of biofertilizer, phytohormone and zinc as a cost effective agro technique for increasing sesame productivity, *Sesame and Safflower Newsletter*, **16**:46-50.
- Tiwari KP, Nanideo KN, Tomar RKS and Raghu JS. 1995. Effect of macro and micronutrients in combination with organic manures on the production of sesame (*sesamum indicum* L.), *Journal of Agronomy*, **40** (1): 134-136.
- Tiwari RK, Namdeo KN, Girish JA and Jha G. 2000. Effect of nitrogen and sulphur on growth, yield and quality of sesame (*Sesamum indicum* L.) varieties, *Research on Crops*, **1**: 163-7.
- Vishwanath HS, Anilakumar KR, Harsha SN, FarhathKhanum A, Bawa S. 2011. In vitro antioxidant activity of *Sesamum indicum* seeds. *Asian Journal of Pharmaceutical and Clinical Research* ,0974-2441.
- Vyakaranahal BS, Shekharagouda M. Patil SA, Prabhakar AS and Giriraj K. 2001. Effects of planting date and chemicals (TIBA and boron) spray on seed yield, its attributes and quality of single and three way cross sunflower hybrids during different seasons, *Seed Research*, **29**: 127-135.
- Xiong ZT, Liu C and Geng B. 2006. Phytotoxic effects of copper on nitrogen metabolism and plant growth in *Brassica pekinensis* Rupr., *Ecotoxicology and Environmental Safety*, **64**: 273-280.
- Yadav RA, Tripathi AK and. Yadav AK. 2009. Effect of micronutrients in combinations with organic manures on production and net returns of sesame (*sesamum indicum*), *Annals of Agricultural Research*, **30**(1 & 2):53-58.
- Yilmaz A, Kiz H E, Torun B, Gulkin I, Karanlk S, Bagci A and Cakmak I. 1997. Effects of different zinc application methods on grain yield and zinc

concentration in wheat cultivars grown on zinc deficient calcareous soils, *Journal of Plant Nutrient*, **20**:461-471.

Zahoor R, Basra SMA, Munir H, Nadeem MA and Yousaf S. 2011. Role of boron in improving assimilates partitioning and achene yield in sunflower, *Journal of Agriculture and Social Science*, **7**: 49-55.

Zeidan MS, Hozayn M and Abd El-Salam MEE. 2006. Yield and quality of lentil as affected by micronutrient deficiencies in sandy soils, *Journal of Applied Science Research*, **2**(12): 1342-1345.

Appendix





Plate 1. Some photographs shows the experimental works.