

**MANAGEMENT OF *TOMATO YELLOW LEAF CURL VIRUS*
THROUGH INSECTICIDES, BOTANICALS AND LIGHT
REFLECTING SILVER COLORED MULCH**

BY

SOMIYA AKTER



**DEPARTMENT OF PLANT PATHOLOGY
SHER-E-BANGLA AGRICULTURAL UNIVERSITY**

DHAKA -1207

JUNE, 2017

**MANAGEMENT OF *TOMATO YELLOW LEAF CURL VIRUS*
THROUGH INSECTICIDES, BOTANICALS AND LIGHT
REFLECTING SILVER COLORED MULCH**

REGISTRATION NO. 11-04394

A Thesis

Submitted to the Faculty of Agriculture,

Sher-e-Bangla Agricultural University,

Dhaka, in partial fulfillment

of the requirements

for the degree

of

MASTER OF SCIENCE

IN

PLANT PATHOLOGY

SEMESTER: JANUARY-JUNE, 2017

Approved by:

Dr. Md. Belal Hossain

Associate Professor

Department of Plant Pathology

Sher-e-Bangla Agricultural University

Supervisor

Dr. Nazneen Sultana

Professor

Department of Plant Pathology

Sher-e-Bangla Agricultural

University

Co-supervisor

Prof. Khadija Akhter

Chairman

Examination Committee

Department of Plant Pathology



Dr. Md. Belal Hossain
Associate Professor
Department of Plant Pathology
Sher-e Bangla Agricultural University
Dhaka-1207, Bangladesh
Cell Number: +88 01711988444
E-mail: mbhossainsau@yahoo.com

CERTIFICATE

This is to certify that thesis entitled, “MANAGEMENT OF TOMATO YELLOW LEAF CURL VIRUS THROUGH INSECTICIDES, BOTANICALS AND SILVER COLORED MULCH” submitted to the Department of Plant Pathology, Faculty of Agriculture, Sher-e-Bangla Agricultural University, Dhaka, in partial fulfillment of the requirements for the degree of MASTER OF SCIENCE IN PLANT PATHOLOGY, embodies the result of a piece of bona fide research work carried out by SOMIYA AKTER bearing Registration No. 11-04394 under my direct supervision and guidance. No part of the thesis has been submitted for any other degree or diploma in any institute.

I further certify that such help or source of information, as has been availed of during the course of this investigation has duly been acknowledged.

.....
Dated: June, 2017
Dr.Md.Belal Hossain
Dhaka, Bangladesh
Supervisor

Dedicated
To
My Beloved Parents

ACKNOWLEDGEMENT

All praises to the “Almighty Allah” who enabled the author to pursue higher education in Plant Pathology and to submit the thesis for the degree of Master of Science (M.S.) in Plant Pathology.

*The author wishes to express her profound sense of appreciation and heartiest gratitude to her Supervisor, **Dr. Md. Belal Hossain**, Associate Professor, Department of Plant Pathology, Sher-e-Bangla Agricultural University, Dhaka for his help, scholastic supervision, continuous encouragement and constructive suggestion throughout the period of research and for taking immense care in preparing this manuscript.*

*The author expresses her immense gratitude to her Co-supervisor, **Dr. Nazneen Sultana**, Professor, Department of Plant Pathology, Sher-e-Bangla Agricultural University, Dhaka for his valuable advice and constructive criticism during the critic period of research work.*

The author is grateful to all the respectable teachers of the Department of Plant Pathology and the Department of Horticulture, Sher-e-Bangla Agricultural University for giving necessary suggestions. The author would like to extend her appreciation to all the staffs of the Department of Plant Pathology, Sher-e-Bangla Agricultural University, Dhaka for their co-operation and encouragement during the study period

The author is also thankful to SAURES (Sher-e-Bangla Agricultural University Research System) for providing research support. The author would like to give thanks to her friends Yeasin, NaharJoly, Mamun, Mou for their cordial help and support in research work and thesis writing. The author can never repay the debt of his beloved parents, uncle, aunty, sisters, brothers and well wishers for their inspiration, constant encouragement and sacrifice for his higher education.

The author expresses her immense gratefulness to all of them who assisted and inspired her to achieve higher education. The author also regret for her inability for not mentioning everyone by name contributing in the present piece of research work.

The Author

MANAGEMENT OF *TOMATO YELLOW LEAF CURL VIRUS* THROUGH INSECTICIDES, BOTANICALS AND LIGHT REFLECTING SILVER COLORED MULCH

ABSTRACT

A field experiment was conducted at the Plant Pathology experimental field of Sher-e-Bangla Agricultural University, Dhaka, Bangladesh during October 2016 to March 2017. The study was aimed to manage the *Tomato yellow leaf curl virus* (*TYLCV*) through insecticides, botanicals and physiological method using light reflecting silver colored mulch. In this study, in total 7 treatments including control were considered viz. T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = light reflecting silver colored mulch and T₇ = Control. High yielding tomato variety BARI Tomato-14 that is susceptible to *TYLCV* was used. The field experiment was carried out in randomized complete block design with three replications. All the treatments showed significant influence on different assayed parameters in the test crop tomato. The lowest disease incidence (%), disease severity (%) and whitefly association was found in T₆ (light reflecting silver colored mulch) that was 13.33%, 1.96% and 2.0 respectively at 60 DAT. The highest disease incidence %, Disease severity % and whitefly association was found in T₇, (control treatment) that was 46.67%, 46.22% and 18 respectively at 60 DAT. From the relationship study between disease incidence (%), and disease severity (%) with whitefly association, it was revealed that disease incidence and severity of *TYLCV* was increased with increasing of whitefly population and vice-versa. Among the treatments, growth parameters, yield and yield attributers were also found better in T₆ treatment (light reflecting silver colored mulch). From this study we can concluded that physical method is the best option instead of insecticides for controlling the insect vectors of *TYLCV*.

LIST OF CONTENTS

Chapter	Title	Page No.
	ACKNOWLEDGEMENTS	i
	ABSTRACT	ii
	LIST OF CONTENTS	iii-iv
	LIST OF TABLES	v
	LIST OF FIGURES	vi
	LIST OF APPENDICES	vii
	ABBREVIATIONS AND ACRONYMS	viii
I	INTRODUCTION	1-4
II	REVIEW OF LITERATURE	5-25
III	MATERIALS AND METHODS	26-33
	3.1 Location of the experimental field	26
	3.2 Characteristics of soil	26
	3.3 Weather condition during the experiment	26
	3.4 Plant material	27
	3.5 Collection of insecticides and Multineem (Neem oil)	27
	3.6 Preparation of Silver colored mulch and Neemax	27
	3.7 Raising of seedlings	28
	3.8 Layout and design	29
	3.9 Fertilizer and manure application	29
	3.10 Treatments of the Experiment	30
	3.11 Cultivation of Tomato	29-31
	3.12 Evaluation the efficacy of treatments in whitefly association	32
	3.13 Parameter assayed	32
	3.14 Statistical analysis	33
IV	RESULTS AND DISCUSSIONS	33-50
	4.1 Incidence (%) of <i>TYLCV</i>	33-35
	4.1.2 Number of healthy leaves/plant	34

LIST OF CONTENTS (Cont'd)

Chapter	Title	Page No.
IV	RESULTS AND DISCUSSIONS	
	4.1.3 Number of infected leaves/plant	35
	4.1.4 Disease severity %	35-38
	4.2 Whitefly association	39-40
	4.3 Relationship between disease incidence (%) and whitefly association	40-41
	4.4 Relationship between disease severity and whitefly association	41-42
	4.5.1 Number of branches/plant	42
	4.6.1 Plant height (cm)	43
	4.6.2 Number of flowers/plant	44
	4.6.3 Weight of fruits/plant	45
	4.6.4 Single fruit weight	45
	4.6.5 Average fruit diameter	45-48
	5 Fruit yield	49
	DISCUSSION	50-52
V	SUMMARY AND CONCLUSION	53-54
	REFERENCES	55-69
	APPENDICES	70-73

LIST OF TABLES

Table No.	Title	Page No.
1.	Disease incidence (%) of <i>TYLCV</i> under different treatments at 30, 45 and 60 DAT	35
2.	Number of healthy leaves/plant in different treatments at 30, 45 and 60 DAT	36
3.	Number of infected leaves/plant in different treatments at 30, 45 and 60 DAT	38
4.	Effect of different treatments on whitefly association at 30, 45 and 60 DAT in pre and post treatment	39
5.	Number of branches/plant at 30, 45 and 60 DAT under different treatments	43
6.	Effect on plant height of <i>Tomato Yellow Leaf Curl Virus</i> under different treatments	43
7.	Effect on yield and yield contributing parameters of tomato through management of tomato yellow leaf curl virus using different treatments	46

LIST OF FIGURES

Figure No.	Title	Page No.
1.	Aluminum foil on selected plot (T ₆).	28
2.	Neemax (Neem leaves extract).	28
3.	Typical symptoms of <i>TYLCV</i> showed in tomato in the experiment field	33
4.	Tomato plants in Silver colored mulch plot	35
5.	Tomato plants in control plot	36
6.	Whitefly association at 30, 45 and 60 DAT under different treatment	40
7.	Relationship between disease incidence and whitefly association	41
8.	Relationship between disease severity and whitefly association	42
9.	Effect on yield of tomato using different treatments against <i>Tomato Yellow Leaf Curl Virus</i>	47
10.	Experimental site	63
11.	Layout of the experiment field	65
12.	Plot treatment with silver mulch material	68

LIST OF APPENDICES

Appendix No.	Title	Page No.
I	Agro-Ecological Zone of Bangladesh showing the experimental location	68
II	Monthly records of air temperature, relative humidity, rainfall and sunshine hours during the period from October 2016 to March 2017	69
III	Characteristics of experimental soil analyzed at Soil Resources Development Institute (SRDI), Farmgate, Dhaka.	69
IV	Layout of the experiment field	70
V	Pictorial view of treatment plots	71

ABBREVIATIONS AND ACRONYMS

%	=	Percentage
AEZ	=	Agro-Ecological Zone
BBS	=	Bangladesh Bureau of Statistics
BCSRI	=	Bangladesh Council of Scientific Research Institute
Ca	=	Calcium
cm	=	Centimeter
CV %	=	Percent Coefficient of Variation
DAS	=	Days After Sowing
DMRT	=	Duncan's Multiple Range Test
e.g.	=	exempli gratia (L), for example
<i>et al.</i> ,	=	And others
etc.	=	Etcetera
FAO	=	Food and Agricultural Organization
g	=	Gram (s)
GM	=	Geometric mean
i.e.	=	id est (L), that is
K	=	Potassium
Kg	=	Kilogram (s)
L	=	Litre
LSD	=	Least Significant Difference
M.S.	=	Master of Science
m ²	=	Meter squares
mg	=	Miligram
ml	=	MiliLitre
NaOH	=	Sodium hydroxide
No.	=	Number
°C	=	Degree Celceous
P	=	Phosphorus
SAU	=	Sher-e-Bangla Agricultural University
USA	=	United States of America
var.	=	Variety
WHO	=	World Health Organization
Mg	=	Microgram

CHAPTER I

INTRODUCTION

Tomato (*Solanum lycopersicum* L.) is belongs to the nightshade family Solanaceae. It is one of the most popular and nutritious vegetable all over the world including Bangladesh and second most growing vegetables next to potato. The world tomato productions scenarios are about 150 million tons of tomatoes were produced in the world in 2009. The largest producer China (41,864,750 tons), accounted for about one quarter of the global production followed by USA (12,902,000 tons) and India (11,979,700 tons) (FAO, 2013). At present, in Bangladesh 6.85% area in under tomato cultivation both in summer and winter, the total production of tomato was about 2,32,000 tons from 24,700 hectares of land with an average yield of 9.39 tons per hectare (BBS, 2016). The yield of the tomato in Bangladesh is very low compare to those of some advanced tomato growing countries (Sharfuddin and Siddique, 1985).

Tomato is often described as “poor man orange” because it is a rich source of minerals, vitamins and organic acids. Nutritionally, it is a significant dietary source of vitamin A and C. Furthermore, recent studies have shown the importance of lycopene, a major component of red tomatoes, which has strong antioxidant properties that can act against human diseases such as cancer and heart disorders. It is also a good source of calcium, iron, minerals etc. (Matin *et al.*, 1996).

Tomato production in Bangladesh is affected by many factors, among them insect pest attack is the major one. Damage caused by the insect pest to commercial tomato may be directly through phloem feeding, or indirectly by the transmission of plant viruses such as *Tomato Yellow Leaf Curl Virus (TYLCV)* (Mehta *et al.*, 1994). *TYLCV* is one of the most important diseases of tomato causing heavy losses in yield and quality of fruits. It is one of the devastating diseases of the tomato crop and depending on the severity and stage

of the infection causing heavy losses in yield (Kalloo, 1988). Yield loss exceeds 90 per cent, when infection occurred within four weeks after transplanting in the field (Saikia and Muniyappa, 1989).

The whitefly (*Bemisia tabaci*) is one of the most economically important pests of tomato in many tropical and sub-tropical regions (Block, 1982). It is polyphagous pests of great significance in agriculture worldwide (Kontsedalov *et al.*, 2012) and currently its host range has crossed 600 plant species (Oliveria *et al.*, 2001). It causes damage by sucking cell sap, secreting the honey dews and transmits a number of viral diseases (Khan and Ahmed, 2005). The insect whitefly breeds throughout the year and the female lays stalked yellow spindle shaped eggs singly on the lower surface of the leaf. Nymphs and adults suck the sap usually from the under surface of the leaves and excrete honeydew. Leaves appear sickly and get coated with sooty mold (Jayaraj *et al.* 1986). The whitefly serves as the potential vector for the spread of *Tomato Yellow Leaf Curl Virus* disease causing severe damage to tomato crop. The whitefly can acquire the virus after feeding on infected plants for 15 to 30 minutes, and can transmit the virus to tomato plants after about 24 hours of incubation within the insect. A period of at least 15 minutes feeding on the new tomato host is subsequently required for transmission of the virus. The whitefly retains the virus for up to 20 days and does not transmit it to its progeny. Symptoms develop on young plants after 10 to 14 days. Hot and dry conditions favor the whitefly, and therefore, help the spread of *TYLCV*. Whitefly populations decrease after heavy rain showers. Under normal conditions whiteflies are passively wind-driven over long distances. During the night they settle on the lower leaf surfaces. Disease incidence increases rapidly and can reach 100% infection at harvest. In the field, disease incidence varies with location rather than with season. Tobacco is a symptomless host and can therefore serve as a source for re-infection of tomato crops.

Hence, whitefly infestation can cause severe and also transmitted *Tomato Yellow Leaf Curl Virus* performed transmission by it, and crop damage can be reached up to 100%, so, it is urgently needed to control whitefly for successful tomato production in our country. Among the various control practices to suppress the prevalence of whitefly insecticides are the mostly used. Several crop protection chemicals belonging to organochlorine, organophosphate and carbamate group have been used to control insect pests. However, the application of these insecticides left a film of persistent poison over the foliage and fruits (Dikshit *et al.*, 2000) and insect developed resistance to these insecticides (Denholm *et al.*, 1996). Therefore, there is a need to replace these conventional insecticides with newer molecules with a lesser dose of a few grams per hectare. Profenophos (organophosphate group), imidacloprid (neo – nicotinyl group), cypermethrin (synthetic pyrethroid group), indoxacarb (oxadiazine group), profenophos + cypermethrin (mixture of organophosphate and synthetic pyrethroid) are new molecules and reported to be effective and economical in controlling insect- pests in many vegetable crops (Shah *et al.*, 2012). The organochlorine and organophosphorus compounds have been reported to pose a potential threat to all types of ecosystem (Nayar *et al.*, 1992). Different groups of insecticides have been recommended to control this white fly (Satpathy *et al.*, 2004), and Suryawanshi *et al.*, 2000). To consider the negative effects of insecticides used in controlling the whitefly population less toxic and environmentally friendly techniques are need to be introduced.

The purpose of this study was to find out the suitable chemical that would be less toxic, botanicals or other management practice like mulching through this way it may be possible to control the insect vector, whitefly. In some previous research has also evaluated less toxic and more environmentally safe phytochemicals to control whitefly. For example- Neemax (Neem leaf Extract) and Multineem (Neem Oil) and different types of mulching to regulate the population of whitefly.

OBJECTIVES

This study was carried out to achieve the following specific objectives-

1. To evaluate the disease incidence and severity of *Tomato yellow leaf curl virus (TYLCV)*.
2. To control the insect vector, whitefly (*Bemisia tabaci*) by using selected insecticides, botanicals and Silver colored mulch for management of *TYLCV*.
3. To increase the ultimate yield of tomato.

CHAPTER II

REVIEW OF LITERATURE

Tomato (*Solanum lycopersicum* L.) is one of the most popular and widely grown vegetables in the world ranking second in importance to potato in many countries. Pest and diseases are the major limiting factors in tomato cultivation. Tomato leaf curl virus disease has recently emerged as a major threat to the cultivation and production of tomato in Bangladesh.

2.1 Historical Perspective

The first incidence of Tomato yellow leaf curl virus (*TYLCV*) in Israel drew attention of the scientists due to its high prevalence in the field and severe damage to the crops (Cohen and Nitzany, 1966). In the early 1960s *TYLCV* was reported as one of the most economically important viruses among the Gemini viruses infecting tomato in different tomato growing areas of the world (Polston and Anderson, 1997). Worldwide distribution of *TYLCV* as a severe virus of tomato was recognized by Goodman in a review paper published in (1981). Semi-persistent transmission of the virus by whitefly (*Bemisia tabaci*) and non-availability of tomato cultivars makes the situation more vulnerable in respect to the management of *TYLCV* (Polston and Anderson, 1997). In the paper published by Polston and Anderson (1997), Al-Musa (1982), Tomlinson (1987) suggested the management of *TYLCV* by growing tomato seedlings in insect proof protective conditions, manipulation of the sowing time, planting of trap crops in the tomato field, as well as judicious use of insecticide as integrated pest management components.

Tomato yellow leaf curl virus (*TYLCV*) mainly affects tomatoes but other crops paprika (*Capsicum annuum*), bean (*Phaseolus vulgaris*), tobacco, lisianthus (*Eustoma grandiflorum*), Zinnia, etc. were also sensitive. Some weeds, with or without symptoms that were widely distributed in France can serve as reservoirs

(*Solarium nigrum*, *Datura stramonium*, and *Malva sp.*) reported by Dalmon and Marchoux (2000).

In Spain two viral species of tomato yellow leaf curl virus were present, *TYLCV-Sar* and *TYLCV-Is*. *TYLCV-Sar* (43.4%) and *TYIXV-Is* (56.6%) coexisted in tomato crops and displacement of *TYLCV-Sar* for *TYLCV-Is* was observed. A search for alternative hosts that may serve as disease reservoirs was conducted by testing 210 samples of 95 weed species. The following species were found to be infected: *Conyza sumatrensis*, *Chenopodium murale*, *Datura stramonium*, *Dittrichia viscosa*, *Malva parviflora*, *Solarium nigrum*, *Convolvulus sp.*, and *Cuscuta sp.* This was the first reference of *Conyza sumatrensis*, *Chenopodium murale*, *Convolvulus sp.* and *Cuscuta sp.* as natural hosts of *TYLCV* reported by Jorda *et al.* (2001).

Tomato is affected by two types of leaf curl virus viz. *Tomato yellow leaf curl virus (TYLCV)* and *Tomato leaf curl virus (ToLCV)*. *Tomato yellow leaf curl virus* was first reported from Israel by Avidov in 1940. Martinez *et al.* (2003) reported the infection of this disease around 7 million ha of crop plants in 40 countries and so far it is not reported from Indian subcontinent (Rishi, 2004; Glick *et al.*, 2009).

2.2 Disease symptoms

In the world, *Tomato yellow leaf curl virus (TYLCV)* was first studied by Cohen and Harpez (1964) in Israel. They studied the symptoms, damaging nature and involvement of whitefly with a new disease of tomato plant in Israel. The disease was studied extensively by Cohen and Nitzany (1966) in respect to transmission and host range and named the causal virus as *Tomato yellow leaf curl virus (TYLCV)*.

Singh and Sastry (1979) reported that *TYLCV* was characterized by severe stunting of plants with downward rolling and crinkling of leaves. The newly formed leaves also exhibited chlorosis symptoms. Older leaves became leathery and brittle. The

nodes and internodes were much reduced in size. The infected plant looked pale and produced more lateral branches resulting in bushy growth.

Dhanju and Verma (1987) mentioned that *TYLCV* was a complex disease with symptoms of crinkling, yellowing and premature withering of leaves together with stunting and profuse branching of plant. They observed that the disease occurred due to combined infection more than one virus.

Pilowsky and Cohen (1990) demonstrated that in Israel *TYLCV* caused severe damage of tomatoes. The affected plants were markedly stunted and their branches and petioles tend to assume erect positions. Leaflets were rolled upward and inward showing interveinal chlorosis. Infected plants were smaller than healthy plants. Fruits sets were greatly reduced and infected young plants produced almost no marketable yield.

Abdel Salam (1991) stated that tomato leaf curl virus, causing various symptoms of leaf curl interveined yellowing and stunting. Both *Phaseolus vulgaris* cv. *Bouniful* and *Xanthium* developed lesion on the primary leaves folded by systemic infection and developed lesion on the primary leaves followed by systemic infection.

Moriones *et al.* (1993) observed symptoms of *TYLCV* as typical yellowing and curling of leaf margin and general stunting of tomato plants in eastern Spain in autumn 1992. This was the first report of *TYLCV* in Spain.

Bosco (1993) reported the epidemiology of *TYLCV* and distribution of *B. tabaci* in Sardinia and some others parts of Italy. The vector was found on nine wild and six cultivated plant species besides tomato. None of the wild plant species was naturally infected by *TYLCV*, but *Solarium* could be infected experimentally and showed clear typical symptoms of *TYLCV*.

As reported by McGlashan *et al.* (1994) from Jamaica tomato fields during the

spring 1993 and 1994, which displayed symptoms, consisted of upward curling of the leaves, severely reduced leaf size, yellowing of the leaf margin and veins, flower abscission and severe plant stunting.

Kegler (1994) reviewed disease of tomato plants infected by *TYLCV* and noted that the infected plants were stunted, developed small chlorotic leaflets and curled lamina between the veins.

Polizzi *et al.* (1994) suggested that the type of symptoms varied depending on the temperature and the time of infection. However, stunting reduced leaf and mild chlorosis having reduced number of fruits and fruit size were observed.

2.3 Diagnosis and identification of virus

Czosnek *et al.* (1988) purified the *TYLCV* particles from infected tomato and datura plants and detected typical twined particles, characteristic of members of the geminivirus group. They confirmed that viruses containing fractions of purified preparations were infective in membrane transmission test with the white fly vector. They also characterised the genome of the virus particles as a circular single-strand DNA containing about 2800 nucleotides.

Navot *et al.* (1989) developed a method for rapid detection of *TYLCV* in squashes of infected plants and insects vector. They described that DNA sequence of *TYLCV* was detected specifically and sensitively by hybridization of infected plant tissues quashed onto a nylon membrane (Squash blot) with a specific DNA probe. No treatment of the sample was necessary before quashing and hybridization. The DNA of *TYLCV* could be detected in squash blots of tomato leaves, roots, flowers and fruits. Viral sequences were also detected in single whitefly that fed on infected plants. The authors also detected tobacco mosaic virus, Potato virus Y and two others RNA viruses in infected tobacco plants using the method. The squash blot method was applied for *TYLCV* infections in the field in Israel and for diagnosing *TYLCV* in Turkey.

Czosnek *et al.* (1990) surveyed for the first time on the TYLCV distribution in different countries of the world. *TYLCV* bigeminivirus was diagnosed in tomatoes collected from Mediterranean countries, America, Western Africa and Southeast-Asia by hybridizing tomato leaflets squash onto nylon membrane with a virus specific DNA probe. Samples positive for *TYLCV* were counted. The results revealed the worldwide distribution of *TYLCV*.

Abdel-Salam (1990) mechanically inoculated Egyptian isolates of *Beet curly top virus (BCTV)* and *TYLCV* to beet and tomato, respectively, and observed that both the geminiviruses were mechanically transmitted. When tested serological using agar gel double-diffusion test with an authentic American *BCTV* antiserum, both the Egyptian isolates and isolates of *TYLCV* from Jordan reacted positively showing their strong serological relationship. The facts were also confirmed by using immunosorbent electron microscopy test.

Acquisition of *TYLCV* by individual whitefly in relation to the length of the access period, the virus concentration and the developmental stage of plant tissues were studied by Zeidan and Czosnek (1991). The results revealed that the frequency of *TYLCV* detections increased with the length of access period. DNA was detected in 15% of whiteflies tested after a period of access to infected tissues for 30 minutes, regardless of whether it had high or low virus content. However, DNA was detected in all insects tested after an eight hours period of access to all the plant. The insects fed on youngest leaves of infected plants having high virus content, acquired detectable *TYLCV*-DNA within two hours. Viruliferous whiteflies retained *TYLCV* for at least 13 days when placed on uninfected tomato plants. In the experiments they used southern-blot analysis of nucleic acid extracted from a single whitefly to detect genomic DNA of *TYLCV*.

In Bangladesh Akanda (1991) collected 23 tomato samples on the basis of symptoms from different parts of Bangladesh and noted six different types of

symptoms prevalent on tomato. The author specially identified yellow mosaic and purple vein as two different symptoms. Finally from those samples six different viruses like *Cucumber mosaic virus (CMV)*, *Tobacco mosaic virus (TMV)*, *Potato virus (PVY)*, *Broad bean wilt virus (BBWV)*, *Tomato rattle virus (ToRV)* and *Alfa alfa mosaic (AMV)* were identified on the basis of symptoms, electron microscopic study, inoculation test and serological test Akanda *et al.* (1991a and 1991b).

However, the authors commented that the two major symptoms (*Yellow mosaic and purple vein*) in respect to prevalence and crop damage could not be identified. The authors named the two viruses as *Tomato yellow leaf curl virus (TYLCV)* causing yellow mosaic symptom and *Tomato purple vein virus (TPVV)* causing purple vein symptom for the first time from Bangladesh.

Green (1998) published a manual on the rapid detection of plant viruses specially *TYLCV* in the name of “Making leaf tissue squashes on membranes for virus detection”. The author described the method as an effective, sensitive and reliable method for virus diagnosis from the plant extract but simpler than the method suggested by Navot *et al.* (1989).

2.4 Vector and transmission of virus

Cohen and Harpaz (1964) studied the involvement of whitefly with a new disease of tomato plant in Israel, which was later identified as *TYLCV*.

Cohen and Nitzany (1966) reported that *TYLCV* was caused by a whitefly-borne virus, which could not be transmitted mechanically and named the causal virus as *TYLCV* for the first time. They noted that the minimum acquisition and inoculation period was 15-30 minutes, the latent period in the vector was at least 21 hours and the virus was persistent in the vector for a period up to 20 days. They found that it was semi-persistent in nature. They also noticed that the females of whitefly were more efficient than male vector of *TYLCV*.

Singh and Sastry (1979) stated that this disease was transmitted by an insect vector, the whitefly (*Bemisia tabaci* Gen.) in the field. Even a single viruliferous whitefly was able to transmit the virus. The virus was neither seed nor sap transmitted.

Makkoub (1978) found two different *TYLCV* isolates on the basis of symptoms produced on tomato and stated that both the isolates were transmitted by whitefly. Goodman (1981) mentioned that *TYLCV* was a whitefly transmitted geminivirus and it was highly prevalent in the Mediterranean region.

Thanapase *et al.* (1983) reported that the phloem of the tomato leaf curl infected plants contained virus particles of the geminivirus type, which could be transmitted by grafting. The host range of *TYLCV* included *Datura stramonium*, *Nicotiana glutinosa* and the tomato cvs. Sida and Marglobe.

Cherif and Russo (1983) examined, tissue samples of tomato plant from Tunisia naturally infected and graft inoculated with tomato yellow' leaf curl virus disease by electron microscopy. Their observation was that the tomato yellow leaf curl w'as a viral disease associated with a non-mechanically transmissible by virus.

Al-Hitty and Sharif (1987) reported that cucumber could be the best host of whitefly due to trapping of vector. *TYLCV* infection was reduced by 48% if planted as trap crop in tomato field. Such treatment also delayed the appearance of virus symptom by 17 days.

Verma *et al.* (1989) stated that the incidence of *TYLCV* on tomato was directly related to the population density of the vector developed during January when incidence of the disease also began to increase.

Brunt *et al.* (1990) noted *TYLCV* as a whitefly transmitted geminivirus having single stranded, circular DNA in the genome present in two parts (twinned particle). They recorded nine different plant species including tomato as its host.

The geminate particles size was 20 nm in diameter and 30 nm in length and these were phloem or phloem restricted in the host. Lukyanenko (1991) pointed out that *TYLCV* transmitted by whitefly was the most serious virus disease of tomato in tropical and sub-tropical Asian countries and part of Africa

Mansour and Al-Musa (1992) reported that the whitefly was an efficient vector. A single whitefly was able to transmit the virus. The minimum acquisition and the inoculation feeding period were 60 and 30 minutes, respectively and the latent period was 11 days.

Brown and Bird (1992) noted that plant viruses transmitted by whiteflies cause over 40 diseases of vegetable and fibre crops worldwide. Depending on the crops, season, whitefly prevalence and other factors, the yield losses ranged from 22-100%.

McGrath and Harrison (1995) compared the cultures of whitefly from Ivory coast (IC), Pakistan (PK) and the USA (USA B-type) for the frequency with which they transmitted three tomato virus isolates namely Indian tomato leaf curl virus from Bangalore (*ITmLCV*) and tomato yellow leaf curl bigeminiviruses from Nigeria (*TYLCV-Nig*) and Senegal (*TYLCV-Sen*). The results demonstrated that the frequency of transmission from tomato to tomato depended both on the whitefly culture and the virus isolates.

Konate *et al.* (1995) surveyed the whitefly transmitted geminivirus disease of cassava, okra, tobacco and tomato in Burkina Faso. They used triple antibody sandwich ELISA with cross-reacting monoclonal antibodies for identification of the virus. They identified *African cassava mosaic virus (ACMV)* from cassava, *Okra leaf curl virus (OLCV)* from okra, *Tobacco leaf curl virus (TLCV)* from tobacco and *Tomato yellow leaf curl virus (TYLCV)* from tomato. *TYLCV* was noted as an economically serious disease reaching a high incidence in March following a peak population of the vector whitefly. They compared the epitope

profile of the viruses and found that *TLCV* and *TYLCV* had the same epitope profile, which suggested that these two viruses were the same viruses.

Murad *et al.* (2001) reported that whiteflies were able to transmit *TYLCV* 8 h after they were caged with infected tomato plants. After increasing acquisition access periods (AAPs) on infected tomato plants, the stylets, the head, the midgut, a hemolymph sample, and the salivary glands dissected from individual insects were subjected to polymerase chain reaction (PCR) without any treatment; the presence of *TYLCV* was assessed with virus-specific primers. *TYLCV* DNA was first detected in the head of *B. tabaci* after a 10 min. AAP. The virus was present in the midgut after 40 min. and was first detected in the hemolymph after 90 min. *TYLCV* was found in the salivary glands 5.5 hr after it was first detected in the hemolymph.

Sohrab *et al.* (2013) also found that, tomato leaf curl virus (ToLCV) inciting yellow mosaic disease in Luffa was sap transmissible and opined that sap transmission for most of the Begomovirus is difficult, except a few. ToLCV is a bipartite virus and due to the presence of the DNA B genes (involved in virus movement), such viruses tend not to be phloem limited and thus likely to be mechanically transmissible. Graft transmission of ToLCV was reported by Vasudeva and Samraj in 1948 and many workers adopted it for screening (Divakaran, 2007 and Yadav, 2011).

ToLCV is transmitted in nature by whitefly, *Bemisia tabaci* (Naik *et al.*, 2004; Rajasri *et al.*, 2011). The greenhouse whitefly, *Trialeurodes vaporariorum* and aphid, *Myzus persicae* can acquire the virus non-specifically but are not able to transmit it (Antignus *et al.*, 1994).

According to Naik *et al.* (2004) five adult whiteflies were required for cent per cent transmission of *ToLCV* from tomato to tomato, when acquisition and inoculation access periods were 24 h each. The minimum acquisition access and

inoculation access periods were 10 and 20 min. respectively. A latent period of 6 h was necessary for *B. tabaci* to become viruliferous.

Rashid *et al.* (2008) noted that, when 3, 5 and 10 viruliferous whiteflies per plant were released, the disease transmission was 20, 30 and 70 per cent, respectively. Cent per cent transmission of TYLCV could be obtained with 15 whiteflies. Rajasri *et al.* (2011) reported 20 per cent transmission of *ToLCD* with a single adult whitefly and cent per cent was obtained with 15 adults /plant.

Prasannath *et al.* (2014) found that the incidence and types of insect-transmitted virus diseases of plants vary with the environmental factors, which have direct or indirect relationships on population dynamics of vectors. Alternative indirect strategies are essential to mitigate the environmental and health hazards of pesticide usage, which is the most predominant method of vector management used at present.

2.5 Yield loss

Sastry and Singh (1973) reported that timely use of correct insecticides not only reduce the white fly population but also checks the spread of the disease to a greater extent. They observed that foliar sprays with Dimethioate (0.05%) Methylparathion (0.02%) and oxydemetomethyl (0.02%) and phorate 10G (15 Kg/ha) at the time of planting not only reduced the population of white fly from 245 to 41 but also resulted in less spread of leaf curl virus. When the plants are infected within 20 days of planting the loss may be upto 92% while infections in 35 and 50 days old crop resulted 74% and 20%, yield loss, respectively.

Al-Musa (1982) worked on *TYLCV* in Jordan and found that in the Jordan valley the incidence of the virus at the end of the season ranged from 0-13.2% in the spring grown tomatoes and 93-100% in field grown tomatoes. He noted that 63% yield loss under greenhouse conditions when tomato plants were inoculated at 15 weeks after planting.

Tomlinson (1987) described the relative importance of 27 viruses in order of economic significance affecting 24 field and vegetable crops in 28 region of the world. He found that *TYLCV* was a leading virus in Thailand, Israel, Ivory Coast, Tunisia and some other countries. *TYLCV* was also considered as one of the economically important 18 viruses affecting six protected vegetable crops in Israel and some other countries.

Martelli and Quacquarelli (1982) listed 40 different virus infecting tomato in different countries of the world, which cause 70- 100%, yield loss of the crop. They also reported that tomato mosaic virus causing tomato mosaic disease, tomato leaf curl virus causing tomato yellow leaf curl disease, and tomato yellow leaf curl virus causing tomato leaf curl disease caused 80, 90, and 100% yield loss of tomato respectively, in most of the tomato growing countries during the survey.

Lukyanenko (1991) pointed out that *TYLCV* transmitted by whitefly was the most serious disease of tomato in tropical and subtropical Asian countries and parts of Africa where yield losses due to this disease were 100%.

MacIntosh *et al.* (1992) reported the severe occurrence of *TLCV*, *TYLCV* and *Abotilon mosaic virus* (AMV) three bigeminiviruses in different European countries causing severe yield loss of crops.

Whitefly transmitted geminiviruses cause over 40 diseases of vegetables and fibre crops worldwide were reviewed by Brown and Bird (1992). During the past decade both prevalence and distribution of whitefly transmitted plantviruses have increased and the impact have been devastating. Depending on the crop season, whitefly prevalence and other factors the yield losses ranged from 22-100%. They also remarked that *TYLCV* was one of the most damaging viruses of tomato prevalent worldwide.

Pilowsky *et al.* (1993) conducted an experiment using *TYLCV* tolerant tomato cultivars TY-20 and TY-10 and susceptible cultivars Naama and Ravit and

tolerant cultivars showed only mild symptoms, whereas the susceptible cultivars became markedly stunted with much reduced fruit set and yield and upward rolled leaflets. Infected TY-20 and TY-10 plants were smaller than healthy plants. Yield reduction 50% and 54% in infected TY-20 and TY-10 plants, respectively. Infected Naama and Ravit plants were stunted and produced severe disease symptoms resulting 99% yield reduction.

Polston *et al.* (1994) reported that a new virus disease of tomato was observed in Dominican Republic in 1989 and the causal agent was identified as *TYLCV*, which caused 90% yield loss of tomato.

According to Green and Kalloo (1994) *TYLCV* was distributed all over the world, especially it was a threat of tomato production in the Mediterranean Basin, West and East Africa. *TYLCV* caused 50- 70% yield reduction usually, which may be as high as 100%. They also reported that *TYLCV* resistant and tolerant tomato cultivars were available in Israel, Egypt and France, which transferred from wild *Solanum* species like *Lycopersiconpimpinellifolium*, *L.peruvianum*, *L. hirsutum* and *L.cheesmanii*, however, they concluded that the level of resistant or tolerant and its stability need to have further investigation.

Considering the prevalence and severity of *TYLCV*, Alam *et al.* (1994) studied on its effects on cellular components of infectedleaves and revealed that the virus infection caused 44% and 50% of chlorophyll and B-carotene, respectively compared to healthy plant. They also observed 25% reduction of phosphorus in infected leaves whilenitrogen, protein and carbon content in infected leaves were increased. Organic acids like oxalic acid, citric acid and melanic acid were found to be drastically reduced in infected leaves of tomato.

Ali *et al.* (2005) reported that in Bangladesh and north east India tomato is cultivated at a commercial scale but one of the limiting factors affecting the successful cultivation of this crop is the existence of whiteflies. Damage caused by

this insect pest to commercial tomato may be directly through phloem feeding, or indirectly by the transmission of plant viruses such as Tomato Yellow Leaf Curl Virus (*TYLCV*). In spite of a variety of control measures applied against pests, crop losses have consistently shown an increasing trend. High infestation levels were found from mid-February to mid-March observed the insect vectors Whitefly (*Bemisia tabaci* Genn.) attack tomato plants during April-November (Maximum damage during Aug-Oct) causing heavy losses.

Pun *et al.* (2005) also reported that the incidence of white fly population had a significant and positive correlation with temperature and sunshine hours while a negative correlation with relative humidity and total rainfall. The disease incidence had a significant and positive correlation with white fly population.

2.6 Management of *TYLCV*

Efforts to manage tomato leaf curl disease have been made since the early phase of plant protection as this disease causes severe damage to tomato crop. Effects of cultural practices, plant protection chemicals, botanicals and bioagents in the management of disease have practical importance and scientific interest. Many workers have conducted extensive studies on these aspects.

Incidence of *TYLCV* is generally characterized by great regional and seasonal variations, which are usually attributed to respective fluctuations in the population density of the whitefly vector Nitzany (1975).

TYLCV resistance from *Lycopersicon pimpinellifolium* is monogenic with incomplete dominance inheritance Pilowsky and Cohen (1975). Al Musa (1982) studied the effect of some intercrops on *TYLCV* of tomato. In the field trial cucumber, eggplant and crop were planted in alternate rows of tomato 30 days before the tomato seedlings were transplanted. *TYLCV* was effectively delayed in cucumber interplant plots whereas; corn or eggplant was not found suitable.

Vani *et al.* (1989) evaluated yellow, transparent polyethylene and straw mulch for the management of mosaic disease in muskmelon. All type of mulches reduced the incidence of mosaic disease caused by *Cucumber green mottle mosaic virus* (CGMMV) and *Watermelon mosaic virus-1* (WMV-1). The reduction was greater in yellow color mulch. Mulching also increased plant growth and yield.

Csizinsky *et al.* (1995) conducted field experiment on the effect of six deferent plastic mulch like blue, yellow, orange, red aluminum, red, white and black on fruit yields and insect vectors of tomato. Aluminum and orange mulch reduced the whitefly numbers, delayed virus infection and increased the yield. Virus symptom development was not delayed and yield did not increase in yellowmulch inside of low number of whiteflies. They concluded that under high insect stress, the insect repellent, soil microclimate-modifying and biologically beneficial effects of the mulch be considered when a mulch color will be selected for tomato production.

Ahmed *et al.* (1996) reported that intercropping tomato with coriander (*Coriandrum saliva*), as whitefly repellent can be an effective disease control strategy against TYLCV.

The effect of mulching with polyethylene sheets of deferent colours (black, transparent, white, and aluminum) on *Bemisia tabaci* and spread of TYLCV. Polyethylene mulching reduced whitefly and TYLCV infection. But mulching with aluminum colour was found to be most effective in reducing number of whiteflies and delay the infection of TYLCV (Davino *et al.*, 1996).

Azam *et al.* (1997) investigated that insecticides (carbofuran, endosulfan, Dimethoate, buprofezin and triazophos) and cultural methods (covering the plants with polyester for 30, 45 or 60 days) are most effective for the control of whitefly and TYLCV in tomatoes. Plants covered in polyester had the lowest populations of whitefly the lowest incidences of TYLCV and the highest yields. Of the insecticides, endosulfan had the most affect.

Cohen *et al.* (1998) reported that UV-blocking nets greatly reduced the population of key insect pests in greenhouses and correspondingly reduced the incidence of virus disease carried by various insects. The use of such films can lead to a major breakthrough in reducing the use of chemical insecticides in conventional agriculture.

Effect of netting in the seedbed was assessed to control *TYLCV* in tomato. Disease incidence and yield did not vary significantly in treated and control plots. Low density of vector during seedling stage might be the reason for such results Kung (1999).

Wongklom (1999) evaluated effectiveness of nylon net (40 mesh) as a physical barrier to control white fly and *TYLCV*. Results indicated that nylon net barrier is effective in controlling whitefly and *TYLCV* incidence.

Xienqui (2000) evaluated the effect of interplanting tomato with the vegetable soybean, corn, sweetpotato, cucumber, okra, on whitefly population and incidence of *TYLCV* in the field. All the crop combination partially reduced *TYLCV* infection. Among the intercrops cucumber and vegetable soybean were much preferred by whiteflies as compared to others.

Pilowsky and Cohen (2000) reported that 25 wild *Lycopersicon* accessions were screened in the greenhouse for resistance to the whitefly-borne *TYLCV*. High levels of resistance were detected in 7 of 9 accessions of *L. peruvianum* and in all 5 accessions of *L. chilense* tested. In contrast, plants of 7 accessions of *L. hirsutum* and 3 of 4 accessions of *L. pimpinellifolium* were highly susceptible. Plants of accession CIAS 27 (*L. pimpinellifolium*) showed moderate resistance to *TYLCV*.

Simone and Momol (2001) reported that to identify early symptoms of *TYLCV* and rogue infected and infected-looking plants from field and place in plastic bags immediately at the beginning of the season, especially during first 3-4 weeks.

Spread of any whiteflies to healthy plants should be prevented.

Ahmed *et al.* (2001) used imidacloprid insecticide, in two applications at four rates (47.6, 71.4, 95.2, and 119g) for indirectly controlling TYLCV through control of its vector whitefly in conjunction with integrated pest management (IPM) practices, in the field crops of tomato cv. Peto 86 California. This spray regimen was compared with standard applications of cypermethrins at 10 to 15 days intervals (with or without IPM) throughout the growing season. In three field trails in Sudan (in the winter season of 1997 and in the summer and winter seasons of 1998), a combination of IPM practices and two applications of Confidor at the two highest rates immediately after sowing and 6 weeks later protected tomato plants against the disease until 12 weeks after sowing. All rates of Confidor reduced disease incidence compared with standard chemical control applied in an integrated strategy and quantitative efficacy increased with increase of insecticide rate.

Kalb (2004) suggested growing seedlings in an insect proof net house (50 mesh or fine), spraying infected plants with imidacloprid before rouging, interplanting tomato with bait plants like cucumber, application of systemic insecticides as soil drenches during seedling stage. Rotation of insecticides is necessary otherwise resistance may develop in the vector. Chemical control is ineffective when disease incidence is high.

Tahir *et al.* (2004) used five different planting dates (May to July) at 15 days interval to manage cotton leaf curl virus (*CLCuV*) in the field. Maximum *CLCuV* incidence was recorded in June 1 planting. Results suggested that plantings should be done before June 1 to minimize the disease loss. .

Momol and Perneny (2006) used imidacloprid (Admire®) in the transplant water. Rates recommended are Admire®, 16 oz/A. Do not use Pravado® if plants were treated with imidacloprid or similar insecticide at transplanting. Insect growth

regulator insecticides can be applied when scouts find nymphal densities to exceed 5 to 10 per leaflet by standard sampling procedures. Repellants (e.g. crop oil, UV-reflective mulch) can be used to interfere with secondary virus spread.

Bajpai (2005) obtained better management of chilli leaf curl virus with combined application of commercial botanical viricides, Pre-vental B.V/ Action 100 (0.2 per cent) with imidacloprid (1ml/3ltr) by reducing the incidence to 6 and 5 per cent respectively against 89 per cent in control. Reddy (2006) observed suppression of tomato leaf curl disease with *Clerodendron inermis* and *Gliricidia* leaf extract at the early stage of the crop (45 DAT) with 13.37 and 14.50 per cent incidence as compared to 48.33 per cent in control.

Arunakumara *et al.* (2010) observed reduction in tomato early blight disease with 0.1 and 0.2 percent perfect. Meena (2012) reported the efficacy of perfect in preventing the growth and sporulation of *Alternaria porii*, causing purple blotch of onion under in vitro conditions and also noticed reduction in disease incidence of 28.29 per cent against 72.09 per cent in control.

Karthikeyan *et al.* (2009) obtained 90 per cent reduction of leaf crinkle disease in black gram with the leaf extracts of *Mirabilis jalapa* and *Bougainvillea spectabilis*. Extracts of *Thuja orientalis*, *Tamarix brachystachy*, and *Lawsonia inermis* had exhibited inhibitory effects on *TYLCV* multiplication in tomato treated plants with protection periods of 10 to 12 days (Al-ani *et al.*, 2011).

The application of 3.5 % neemazol, 2% neem oil and 5% NSKE were found superior in reducing the whitefly population in okra by Naik *et al.* (2012).

Samiyappan (2003) reported the efficacy of *Pseudomonas fluorescens* in suppressing the viral infection and inducing systemic resistance in tomato plants against whiteflies and thrips.

Mishra *et al.* (2012) screened fifty rhizobacterial isolates against tomato leaf curl virus (ToLCV) disease under glasshouse condition and found that application of rhizobacteria based bioformulations to seed, soil and foliage significantly reduced the disease severity from 85.72 to 28.58 per cent with two isolates of *Pseudomonas* sp. Mishra *et al.* (2014) reported that, application of chitin based *Pseudomonas* reduced the disease severity of 90.33 to 80.33 per cent and also observed that, addition of chitosan has enhanced the bioefficacy of *Pseudomonas* against *ToLCV*.

Treatment of tobacco mosaic virus (*TMV*) contaminated tools with 20 per cent solution of nonfat dry milk (*NFDM*) and 0.1per cent Tween-20 completely eliminated *TMV* transmission in *Petunia* (Lewandowski, *et al.*, 2010).

Abdelbacki *et al.* (2010) observed suppression of Tomato yellow leaf curl virus infection with the application of native or modified whey proteins fractions at 15 days of treatment.

The chemicals like chlorinated hydrocarbons, organophosphates, neonicotinoids, pyridine-azomethines, and pyrethroids were reported to be effective against whitefly. But, whiteflies have developed resistance to many of these chemicals and efficacies had decreased over time (Ahmed *et al.*, 2001).

In addition to these insecticides, oils, insecticidal soaps, and insect growth regulators have also been used for controlling whiteflies. The most effective and widely used class of insecticides to reduce whitefly populations is the neonicotinoids of which thiomethoxam, imidacloprid, and dinotefuran have been widely used to reduce incidence of tomato leaf curl virus in many tropical countries (Polston and Lapidot, 2007).

Iersel *et al.* (2000) reported that, the application of imidacloprid to *Poinsettia* by subirrigation as a practical and efficient method to control silverleaf whiteflies. Zacharia (2006) observed reduction in the bittergourd distortion mosaic disease

with imidacloprid 0.025 per cent. The application of imidacloprid 0.05 per cent at 15 days interval was found effective for the control of whitefly population to reduce the incidence of ToLCD (Reddy, 2006).

Ramu *et al.* (2011) carried out an experiment to manage *B. tabaci*, the vector of yellow vein mosaic of mesta and found that, acetamiprid (@ 0.2g/l) and thiamethoxam (@ 0.2g/l) were found most effective which recorded lowest whitefly population, disease incidence (3.15%) and highest fibre yield(18.00q/ha) and followed by imidacloprid (@ 0.2ml/l) and triazophos@ 2ml/l.

Chandrashekharaiyah *et al.* (2013) found that, imidacloprid applied green gram plots recorded less whitefly population, yellow vein mosaic incidence and highest yield.

Patel *et al.* (2013) carried out a field experiment to know the effect of different insecticides, biopesticides and its combination on incidence of tomato yellow leaf curl virus (TYLCV) disease and population of whitefly on tomato cv. Pusa Ruby. The disease incidence of *TYLCV* was recorded periodically from 15 to 90 days after transplanting (DAT) with an interval of 15 days by visual observation. Among the all treatments; thiacloprid treated plots showed lowest average whitefly population and proved to be effective treatment with lowest mean disease incidence 30.24 per cent and highest marketable fruit yield 18.38 q/ha⁻¹ while the more average whitefly population was observed in Cowdung + cow urine @ 20ml/l treated plots and control plots showed highest disease incidence and more number of whiteflies per plant.

Prasannath *et al.* (2014) conducted a study to determine the variation of virus disease incidence and the population of insect vectors in field grown tomato (*Solanum lycopersicum* L.) cv. Thilina in different environmental conditions using two crop management systems namely, existing management system with pesticide applications and an integrated management package (IMP) with less

reliance on pesticides. Results revealed that there was no significant ($p=0.05$) difference between the two types of management systems in terms of virus disease incidence indicating the equal efficiency of the tested IMP treatment and existing pest control method on virus disease management. Abundance of insect vectors and beneficial insect populations were significantly ($p<0.05$) influenced by the interaction effect of the management system and location.

Subba *et al.* (2017) conducted a research to study the population dynamics of whitefly (*Bemisia tabaci* Genn.) infesting tomato (*Lycopersicon esculentus* L.) and their sustainable management using biopesticides. The insecticide acetamiprid was found most lethal against whitefly providing 76.59% suppression, closely followed by extracts of neem + *Spilanthes* providing 62.39% suppression. Neem and *Spilanthes* individually did not produce good results but when used as a mixture they recorded better results. Highest yield (30.15 t/ha) were recorded from acetamiprid treated plots followed by neem + *Spilanthes* (27.55 t/ha). Azadirachtin and Plant extracts are biopesticides having less or no hazardous effects on human health and environment. Thus they can be incorporated in IPM programmes and organic farming in vegetable cultivation.

Jha and Kumar (2017) conducted a field trial on a tomato crop variety Avinash 2 to evaluate the efficacy of different insecticidal treatments against whitefly, *Bemisia tabaci*. Three sprays at ten days interval of ten treatments with three replications were applied in the field. The treatment were profenophos @ 500 g a.i. ha⁻¹, imidacloprid @ 20 g a.i. ha⁻¹, cypermethrin @ 25 g a.i. ha⁻¹, indoxacarb @ 50 g a.i. ha⁻¹, profenophos 40% + cypermethrin 4% @ 440 g a.i. ha⁻¹, neem seed kernel extract 5%, neem oil 2%, tobacco decoction 5%, yam bean seed extract 5% and control (water spray). The data (population of whitefly) were recorded one day before first spraying and first, fifth and seventh day of each spray. The data revealed that after each spray all the insecticidal treatments were significantly superior over control in reducing whitefly population and efficacy

was maximum in imidacloprid followed by profenophos 40% + cypermethrin 4% and it was minimum in tobacco decoction while efficacy of other insecticides were in between these insecticides.

CHAPTER III MATERIALS AND METHODS

The present study regarding management of *Tomato yellow leaf curl virus* (TYLCV) through insecticides, botanicals and Silver colored mulch has been conducted during October 2016 to March 2017 at the experimental fields of Sher-e-Bangla Agricultural University, Sher-e-Bangla Nagar, Dhaka. Required materials and methodology are described below under the following headings and subheadings.

3.1 Location of the experimental field

The experiment was conducted at the central farm of Sher-e-Bangla Agricultural University, Dhaka. The site is 22°46'N and 90°22' E Latitude and at Altitude of 9 m from the sea level. The location of the experiment field have been in Appendix I.

3.2 Characteristics of soil

The soil of the experimental site is a medium high land belonging to the Modhupur Tract under the Agro Ecological Zone (AEZ) 28. The soil texture was silty loam with a pH 6.7. Soil samples of the experimental plot was collected from a depth of 0 to 30 cm before conducting the experiment and analyzed in the Soil Resources Development Institute (SRDI), Farmgate, Dhaka. Details of the mechanical analysis of soil sample have been in Appendix II.

3.3 Weather conditions during the experiment

The weather condition of the experimental site was under the sub-tropical monsoon climate, which is characterized by heavy rainfall during Kharif season (April to September) and in the Rabi season (October to March) low rainfall associated with moderately low temperature, low humidity and short day. There was no rain fall during the month of December, January and February, little rain in

March. Rabi is the more favorable for vegetable production. Details of the meteorological data in respect of temperature, rainfall and relative humidity during the study period were collected from Bangladesh Meteorological Department, Agargoan, Dhaka- 1207, Dhaka and have been presented in Appendix III.

3.4 Plant material

The tomato variety “BARI Tomato-14” was used in this study. It was an open pollinated high yield indeterminate type variety developed by the Vegetable Division of Horticulture Research Center, Bangladesh Agricultural Research Institute (BARI), Gazipur.

3.5 Collection of insecticides and Multineem (Neem oil)

Three insecticides namely Imidacloprid, ACmix and Subicron and Multineem (neem oil) were collected from local market.

3.6 Preparation of Silver colored mulch and Neemax

For preparation of Silver colored mulch, at first we have collected clear polythene paper and aluminum foil paper for the local market and then aluminum foil was pasted on polythene paper to prepare the silver metallic mulch. Neem leaves were collected from Sher-e-Bangla Agricultural University campus for preparation of Neemax (Neem leaves extract). For preparation of neem leaves extracts, collected leaves were weighted in an electric balance and then washed in the water. After washing the big leaves were cut into small pieces. For getting extract, weighted plant parts were blended in a mortar & pestle and then distilled water was added into the mortar. The pulverized mass was squeezed through 3 folds of fine cotton cloth. For getting 1:2 (w/v) ratio 200 ml of distilled water was added with 100 g plant parts.



Figure 1. Aluminum foil on selected plot (T₆).



Figure 2. Neemax (Neem leaf extract).

3.7 Raising of seedlings

Tomato seedlings were raised in seedbeds situated on a relatively high. The size of each seed bed was 3 m × 1 m. The area was well prepared with spade and made into loose, friable and dried mass to obtain fine tilth. All weeds and stubbles were removed and the soil was mixed with well decomposed cow dung. Sevin 85 SP was applied around each seedbed as precautionary measure against ants and cutworms. Ten grams of seeds were sown in each seedbed. After sowing, the seeds were covered with light soil to a depth of about 0.6 cm. Complete germination of the seeds took place within 4-6 days of sowing. Necessary shading by bamboo mat was provided over the seedbed to protect the young seedlings from the

scorching sunshine or heavy rain. Dithane M-45 was sprayed on the seedbeds at the rate of 2g/l to protect the seedlings from damping-off and other diseases. Weeding, mulching, and irrigation were done from time to time as and when needed. No chemical fertilizer was used in the seedbed.

3.8 Layout and design

The experiment comprised 7 treatments of single factor and laid out in Randomized Complete Block Design (RCBD) with three replications. The whole field was divided into three blocks and each block consisted of 7 plots. Altogether there were 21 unit plots. Each plot was 6 m² (3 m × 2 m) in size. The distance between plot to plot was 1.0 m and distance between plant to plant was 60 cm and row to row was 40 cm (Appendix IV).

3.9 Fertilizer and manure application

The following doses of manure and fertilizers were used.

<u>Manure/fertilizer</u>	<u>Dose/ha</u>
Cow-dung	10 ton
Urea	400 kg
TSP	250 kg
MP	200 kg

3.10 Treatments of the Experiment

In total seven (7) treatments were considered in this experiment. These were as follows:

T₁ = Imidacloprid

T₂ = ACmix

T₃ = Subicron

T₄ = Neemax (Neem leaf Extract)

T₅ = Multineem (Neem Oil)

T₆ = Light reflecting silver colored mulch

T₇ = Control

3.11 Cultivation of tomato

The seedlings were always kept under close observation. Necessary intercultural operations were done throughout the cropping season to obtain proper growth and development of the tomato plants.

3.11.1 Land preparation

The selected land for the experiment was first opened in October, 2016 by power tiller and expose to the sun for a week. After one week the land was ploughed and cross-ploughed several times with a power tiller and laddering was done to obtain good tilth. Weeds and stubble's were removed and the large clods were broken into smaller pieces to obtain a desirable tilth of soil for sowing of seeds. After removal of the weeds, stubbles and dead roots, the land was leveled and the experimental plot was separated in to the unit plots and were prepared as 10 cm raised beds.

3.11.2 Transplanting of seedlings

Healthy and uniform sized 30 days old seedlings were uprooted separately from the seedbeds. The seedbeds were watered before uprooting the seedlings so as to minimize the root injury. The seedlings were transplanted in the pits of the experimental plots in the afternoon on 10 November, 2016 maintaining a spacing of 40 cm and 60 cm between the rows and plants, respectively. Light irrigation was given immediately after transplanting by using a watering cane. In order to gap filling and to check the border effect, some extra seedlings were also transplanted around the border area of the experimental field.

3.11.3. Intercultural operations

The following intercultural operations were done for better growth and development of the plants during the period of the experiment.

3.11.3.1 Gap filling

Gap filling was done in place of dead or wilted seedlings in the field using healthy seedlings of the same stock previously planted in the border area.

3.11.3.2 Weeding and mulching

Weeding and mulching were accomplished as and whenever necessary to keep the crop free from weeds, for better soil aeration and to break the soil crust. It also helps in conservation of soil moisture. Four subsequent weeding were done manually at 15, 30, 45 and 55 DAS to keep the plots free from weeds. The selected plots under the treatment of T6 were Silver colored mulch.

3.11.3.3 Staking

When the plants were well established, staking was given to each plant by Dhaincha (*Sesbania* sp.) and Bamboo sticks to keep them erect.

3.11.3.4 Drainage

Stagnant water effectively drained out at the time of heavy rains.

3.1.3.5 Irrigation

Irrigations were given throughout the growing season as and when necessary.

3.11.3.6 Spraying Spraying Insecticide, Botanicals for 3 times at 25, 40 and 55 DAT in the plot

3.12 Identification and estimation of disease incidence (%) and disease severity (%)

Identification of the virus disease was done mainly through visual observation of typical symptoms of *TYLCV* infection like upward curling, cupping, with or without marginal chlorosis, smaller leaflets and stunting of the plant (Green and Kalloo 1994 and Sinistera *et al.*, 2000). The incidence of *TYLCV* was calculated by counting the infected plants 30, 45 and 60 DAT on the basis of the appearance of symptoms. The plants were inspected every day morning to note the appearance of the symptoms starting from the following day of transplantation.

The following formulas were used to calculate the percentage of disease incidence and severity

$$\text{Disease incidence (\%)} = \frac{X_i}{X} \times 100$$

X = Total number of plants in a unit

X_i = Number of infected plants in a unit plot

$$\text{Disease severity} = \frac{A_i}{A} \times 100$$

A_i = Number of infected leaves

A = Total number of leaves in selected infected plant

3.12. Evaluation the efficacy of treatments in whitefly association

The sampling on the incidence of whitefly association and the occurrence of *TYLCV* diseases by direct visual method (Hirano *et al.*, 1993). The sampling of the study for whitefly association was taken at vegetative, early flowering and early fruiting at 15 days interval. The plants were carefully checked visually for the presence of whitefly. Sometimes plants were shaken gently to observe their presence and count their number accurately. While the population of whitefly was

very low the number was recorded per 5 plants. Sampling on whitefly incidence was taken at both pre and post application of treatments. Two post treatment counts were taken at each vegetative, early flowering and early fruiting stages.

3.13 Evaluation the efficacy of treatments in whitefly association:

The following parameters were assayed:

1. Number of infected plants/plot
2. Number of healthy leaves/plant
3. Number of infected leaves/plant
4. Whitefly association
5. Number of branches/plant
6. Plant height (cm)
7. Number of flowers/plant
8. Number of fruits/plant
9. Single fruit weight (g)
10. Weight of fruits/plant (kg)
11. Average fruit diameter (cm)
12. Fruit yield (t/ha)

3.14 Statistical analysis

Data were analyzed by using computer based software MSTAT-C. The means of growth and yield data were compared by DMRT Bar diagram and graphs were also used to interpret the data as and when necessary.

CHAPTER IV

RESULTS AND DISCUSSIN

The present experiment was conducted to study the management of *Tomato Yellow Leaf Curl Virus (TYLCV)* through insecticides, botanicals and Silver colored mulch. The data on disease incidence (%), Disease severity (%), whitefly association and yield & yield attributing characters are studied. The results have been presented and discussed under the following headings and subheadings.

4.1 The Morphological Features which are identical,in relation to disease incidence(%) and disease severity (%) of *Tomato yellow leaf curl virus (TYLCV)* in tomato

The typical symptoms produced by *TYLCV* in tomato in the experiment field which was used for identification of the virus are shown in figure 3. On the basis of symptomology appear in tested plants, it was observed that the disease incidence (%) and disease severity (%) of *TYLCV* under different treatments was varied significantly.



Figure 3. Typical symptoms of *TYLCV* showed in tomato in the experiment field.

4.1.1 Effect of different treatments on disease incidence (%) at 30, 45 and 60DAT

The disease incidence (%) of *TYLCV* under different treatments was varied significantly. It was found that the disease incidence (%) of *TYLCV* was ranged 11.11 to 33.33%, 13.33 to 40.00 (%) and 13.33 46.67 (%) at 30, 45 and 60 DAT respectively.

At 30 DAT, the lowest disease incidence (11.11%) was found in T₆ (Silver colored mulch) and the highest disease incidence (33.33%) was recorded in T₇ (control treatment). Among the chemical treatments, the lowest disease incidence (15.56%) was found in T₁ (Imidacloprid) followed by T₂ (ACmix) and T₃ (Subicron) and there disease incidence was 17.78% and 17.78% respectively. In case of botanical treatments Multineem, T₅ (Neem oil) showed moderate disease incidence (22.22%) at 30 DAT which was nearest with other botanical treatment, Neemax, T₄ (24.44%).

At 45 DAT, the lowest disease incidence (13.33%) was also found in T₆ (Silver colored mulch) and the highest disease incidence (40.00%) was recorded in T₇ (control treatment). Among the chemicals treatments, the lowest disease incidence (22.22%) was found in T₁ (Imidacloprid) and T₂ (ACmix) treatments which is statistically same with each other followed by T₃ (Subicron) and it's disease incidence was 24.44%. In case of botanical treatments, Multineem, T₅ (Neem oil) showed moderate disease incidence (26.67%) which is statistically different with Neemax, T₄ (31.11%).

At 60 DAT, the lowest disease incidence (13.33%) was again found in T₆ (Silver colored mulch) and the highest disease incidence (46.67%) was found in T₇ (control treatment). Among the chemical treatments, the lowest disease incidence (24.44%) was found in T₁ (Imidacloprid) which is statistically different with T₂ (ACmix) and T₃ (Subicron) and there disease incidence was 26.67% and 28.89% respectively. In case of botanical treatments, Multineem, T₅ (Neem oil) showed

moderate disease incidence (28.89%) which is statistically different with Neemax, T₄ (33.33%). These results are presented in Table 1. From the all investigation, it was noticed that the disease incidence (%) was very low in Silver colored mulching plots as clearly shown in Figure 3.

Table 1. Disease incidence (%) of *TYLCV* under different treatments at 30, 45 and 60 DAT

Treatments	Disease incidence (%) of <i>TYLCV</i>		
	30 DAT	45 DAT	60 DAT
T ₁	15.56 e	22.22 e	24.44 e
T ₂	17.78 d	22.22 e	26.67 d
T ₃	17.78 d	24.44 d	28.89 c
T ₄	24.44 b	31.11 b	33.33 b
T ₅	22.22 c	26.67 c	28.89 c
T ₆	11.11 f	13.33 f	13.33 f
T ₇	33.33 a	40.00 a	46.67 a
LSD _{0.05}	1.014	1.109	1.127
CV (%)	5.855	7.367	8.214

T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax (Neem leaf extract)

T₅ = Multineem (Neem oil) T₆ = Silver colored mulch, T₇ = Control



Figure 4. Tomato plants in Silver colored mulch plot.



Figure 5: Tomato plants in control plot

4.1.2 Number of healthy leaves/plant

Number of healthy leaves/plant affected by *TYLCV* differed significantly under different treatments. Results revealed that the treatment T₆ (Silver colored mulch) produced the highest number of healthy leaves/plant (34.07, 98.31 and 95.50 at 30, 45 and 60 DAT respectively) followed by T₁ (Imidacloprid) and T₂ (ACmix) at 45

and 60 DAT where the lowest number of healthy leaves/plant (27.20, 51.75 and 45.15 at 30, 45 and 60 DAT respectively) was achieved from Control treatment (T₇). Among the treated plants, the lowest number of healthy leaves/plant was achieved from T₅ (Multineem) followed by T₄ (Neemax).

Table 2. Number of healthy leaves/plant in different treatments at 30, 45 and 60 DAT

Treatments	Number of healthy leaves/plant		
	30 DAT	45 DAT	60 DAT
T ₁	31.33 c	90.85 b	87.45 b
T ₂	32.27 b	77.66 c	73.85 c
T ₃	30.93 c	71.63 d	67.03 d
T ₄	30.20 d	62.30 f	57.51 f
T ₅	30.73 c	68.87 e	64.67 e
T ₆	34.07 a	98.31 a	95.50 a
T ₇	27.20 e	51.75 g	45.15 g
LSD _{0.05}	0.607	1.142	2.014
CV (%)	6.348	10.556	9.417

T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax (Neem Seed Kernel Extract extract) T₅ = Multineem (Neem oil) T₆ = Silver colored mulch, T₇ = Control

4.1.3 Number of infected leaves/plant

Number of infected leaves/plant affected by *TYLCV* differed significantly under different treatments. Results revealed that the treatment T₆ (Silver metallic plastic mulch) showed the lowest number of infected leaves/plant (0.67, 2.07 and 1.65 at 30, 45 and 60 DAT respectively) followed by T₁ (Imidacloprid) and T₂ (ACmix) where the highest number of infected leaves/plant (7.02, 14.20 and 20.87 at 30, 45 and 60 DAT respectively) was achieved from Control treatment (T₇). Among the

treated plants, the highest number of infected leaves/plant was achieved from T₄ (Neemax) followed T₅ (Multineem).

Table 3. Number of infected leaves/plant in different treatments at 30, 45 and 60 DAT

Treatments	Number of infected leaves/plant		
	30 DAT	45 DAT	60 DAT
T ₁	1.47 d	6.47 e	13.19 d
T ₂	2.43 bc	8.57 cd	15.24 c
T ₃	2.07 d	9.07 c	15.76 c
T ₄	3.17 b	10.40 b	17.33 b
T ₅	2.83 b	9.13 c	15.83 c
T ₆	0.67 e	2.07 f	1.65 e
T ₇	7.02 a	14.20 a	20.87 a
LSD _{0.05}	0.401	0.514	1.046
CV (%)	5.246	7.374	8.221

T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = Silver colored mulch, T₇ = Control

4.1.4 Effect of different treatments on disease severity (%) at 30, 45 and 60 DAT

It was found that the disease severity (%) of *TYLCV* was ranged 1.96 to 25.80%, 2.10 to 27.43(%) and 1.72 to 46.22(%) at 30, 45 and 60 DAT respectively. At 30 DAT the lowest disease incidence was found (1.96%) in T₆ (Silver colored mulch) and the highest disease severity was (25.80%) was recorded in T₇ (Control treatment). Among the chemical treatments the lowest disease severity (4.69%) was found in T₁ (Imidacloprid) which is statistically similar with T₂ (ACmix) and T₃ (Subicron) and disease severity was 7.53% and 6.69% respectively. In case of botanical treatments Multineem, T₅ (Multineem) showed moderate disease incidence (9.20%) which is statistically similar with other botanical treatment, Neemax, T₄ (10.49%).

At 45 DAT the lowest disease Severity (2.10%) was found in T₆ (Silver colored mulch) and the highest disease severity (27.43%) was recorded in T₇ (Control treatment). Among the chemicals treatments the lowest disease severity (7.12%) was found in T₁ (Imidacloprid) which is statistically similar with T₂(ACmix) and T₃ (Subicron) and disease incidence was 11.26% and 12.66% respectively. In case of botanical treatment Multineem,T₅ (Neem oil) showed moderate disease severity (13.25%) which is statistically similar with Neemax,T₄ (16.69%).

At 60 DAT the lowest disease severity (1.72%) was found in T₆ (Silver colored mulch) and the highest disease severity (46.22%) was found in T₇ (Control treatment). Among the chemical treatments the lowest disease severity (15.08%) was found in T₁ (Imidacloprid) which is statistically similar with T₂ (ACmix) and T₃ (Subicron) and disease incidence was 20.63% and 23.51% respectively. In case of botanical treatment Multineem,T₅ (Neem oil) showed moderate disease incidence (24.47%) which is statistically similar with Neemax,T₄ (30.13%).

Table 4. Effect of different treatments on disease severity (%) of TYLCV in tomato at 30,45 and 60 DAT

Treatments	Disease Severity (%)		
	30 DAT	45 DAT	60 DAT
T ₁	4.69 e	7.12 e	15.08 e
T ₂	7.53 d	11.26 cd	20.63 d
T ₃	6.69 d	12.66 c	23.51 c
T ₄	10.49 b	16.69 b	30.13 b
T ₅	9.20 bc	13.25 c	24.47 c
T ₆	1.96 f	2.10 f	1.72 f
T ₇	25.80 a	27.43 a	46.22 a
LSD _{0.05}	1.076	1.213	2.057
CV (%)	6.274	8.152	8.875

T₁ = Imidacloprid, T₂ = Acme, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = Silver colored mulch, T₇ = Control

4.2 Effect of different treatments on whitefly association at 30, 45 and 60 DAT in pre and post treatment

In terms of whitefly association, significant variation was observed among the performance of treatments against whitefly (Figure 6). It was observed that the lowest whitefly association (2.0, 4.0 and 2.0 at 30, 45 and 60 DAT respectively) was found in T₆ (Silver colored mulch) followed by T₁ (Imidacloprid), T₂ (ACmix) and T₃ (Subicron) treatments. The highest whitefly association (19, 23 and 18 at 30, 45 and 60 DAT respectively) was recorded in Control treatment (T₇) during crop duration. Among the treated plants the moderate whitefly association (14, 14 and 12 at 30, 45 and 60 DAT respectively) was observed in T₄ (Neemax) which was statistically identical with T₅ (Multineem) at different observation. The results of the present study demonstrated that T₆ (Silver colored mulch), T₁ (Imidacloprid) and T₂ (ACmix) treatment were as the best to control whitefly association.

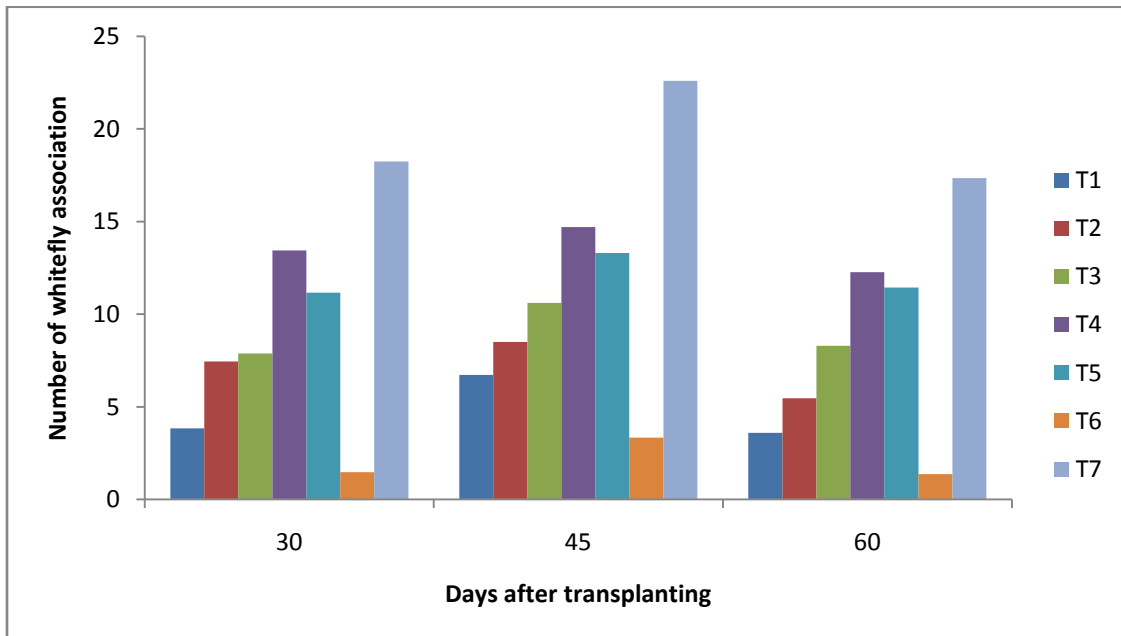


Figure 6. Whitefly association at 30, 45 and 60 DAT under different treatment

T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = Silver colored mulch, T₇ = Control

4.3. Relationship between disease incidence (%) and whitefly association

From the figure 7, it is revealed that % disease incidence was increased with the increased of whitefly association. The highest disease incidence (46.67 %) was recorded in T₇ (control) and whitefly association (18) were recorded in T₇. The lowest number of whitefly (2.0) were recorded in the T₆ (mulch treatment) and disease incidence was 13.33%.

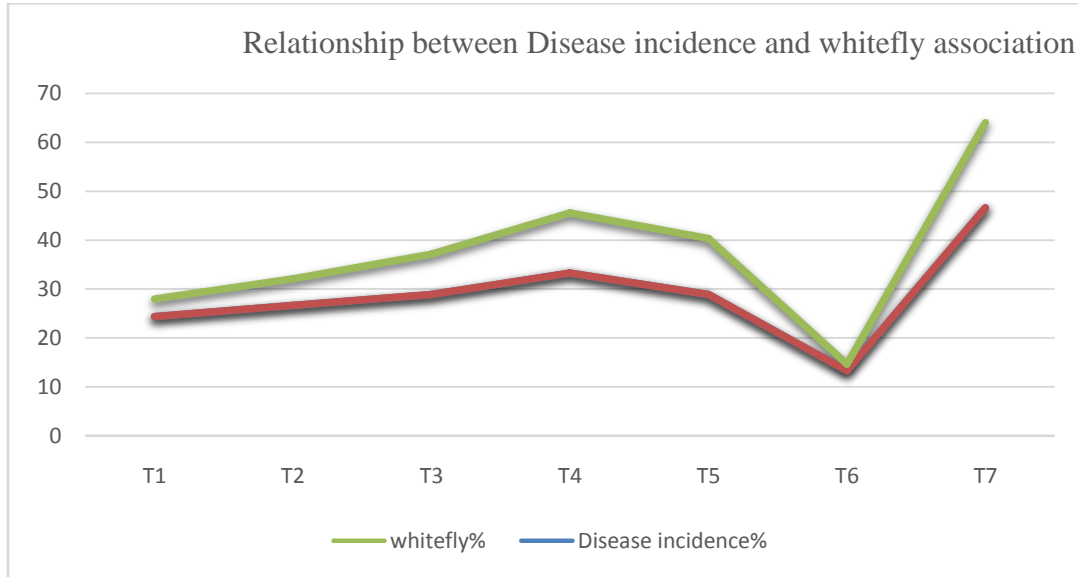


Figure 7: Relationship between disease incidence and whitefly association

T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = Silver colored mulch, T₇ = Control

4.4. Relationship between disease severity and whitefly association:

The highest disease severity and whitefly association were recorded in T₇

From the figure 8, it is revealed that % disease severity was increased with the increased of whitefly association. The highest disease severity (46.22 %) was recorded in T₇ (control) and whitefly association (18) were recorded in T₇. The lowest number of whitefly (2.0) were recorded in the T₆ (mulch treatment) and disease severity was 2.87%.

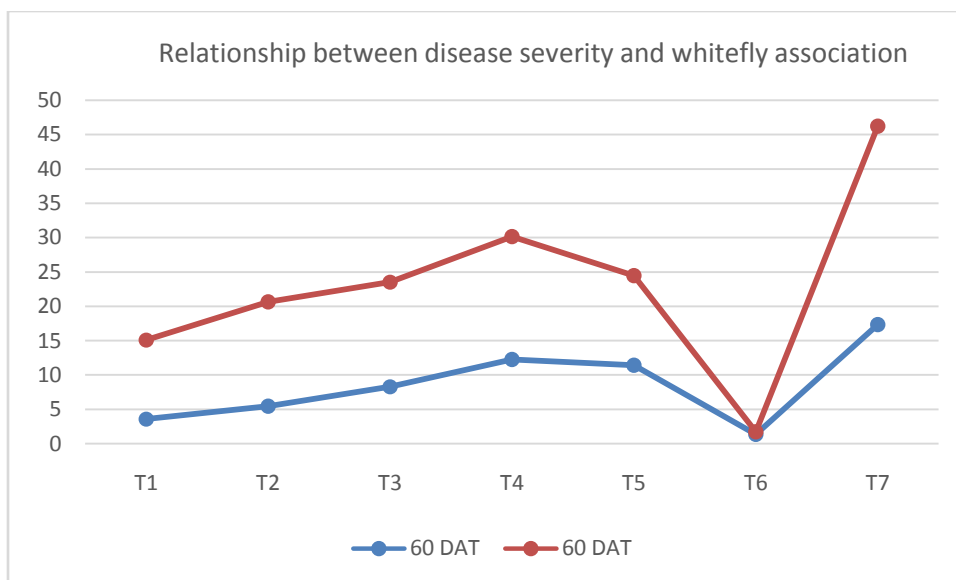


Figure 8: Relationship between disease incidence and whitefly association

T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = Silver colored mulch, T₇ = Control

4.5 The Morphological Features which are identical, in relation to growth promoting characters in tomato against *Tomato Yellow Leaf Curl Virus*

4.5.1 Number of branches/plant

Significant variation was found for number of branches/plant at different days after transplanting affected by *TYLCV* considering the performance of different treatments applied to the test crop (Table 5). It was found that the treatment T₆ (Silver mulch) showed the highest number of branches/plant (6.20, 7.21 and 8.07 at 30, 45 and 60 DAT respectively) followed by T₁ (Imidacloprid) and T₂ (ACmix) where the lowest number of branches/plant (7.00, 7.33 and 5.22 at 30, 45 and 60 DAT respectively) was achieved from Control treatment (T₇). Among the treated plants, the lowest number of branches/plant was achieved from T₄ (Neemax) followed T₅ (Multineem).

Table 5. Number of branches/plant at 30, 45 and 60 DAT under different treatments

Treatments	Number of branches/plant		
	30 DAT	45 DAT	60 DAT
T ₁	5.60 b	6.08 b	6.91 b
T ₂	5.47 b	5.93 b	6.73 b
T ₃	5.27 c	5.38 d	6.19 d
T ₄	4.13 d	5.28 d	6.14 d
T ₅	5.20 c	5.67 c	6.50 bc
T ₆	6.20 a	7.21 a	8.07 a
T ₇	4.00 d	4.33 e	5.22 e
LSD _{0.05}	0.152	0.216	0.264
CV (%)	4.934	6.115	6.539

T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = Silver colored mulch, T₇ = Control

4.5.2 Plant height (cm)

The results on the effect of *TYLCV* infection on plant height of tomato under different treatments are summarized in (Table 6). Significant variation was found on plant height among the treatments. Irrespective of treatments healthy plants produced higher plant height in comparison to *TYLCV* infected plants. The T₆ (Silver colored mulch) treatment produced the highest plant height 99.21 cm at followed by T₁ (Imidacloprid) ,94.25 cm and T₂ (ACmix), 91.39 cm The lowest plant height 79.99 cm was achieved from Control treatment (T₇). Among the treated plants, the lowest plant height was achieved from T₅ (Multineem), 85.75 cm followed by T₄ (Neemax), 83.83 cm.

Table 6. Effect on plant height of *Tomato Yellow Leaf Curl Virus* under different treatments

Treatments	Plant height (cm)
T ₁	94.25 b
T ₂	91.39 c
T ₃	87.37 d
T ₄	83.83 e
T ₅	85.75 f
T ₆	99.21 a
T ₇	79.99 g
LSD _{0.05}	1.361
CV (%)	10.642

T₁=Imidacloprid, T₂= ACmix, T₃= Subicron, T₄=Neemax, T₅= Neem oil, T₆= Silver mulch, T₇= Control

4.6 The Morphological Features which are identical, in relation to yield contributing parameters in tomato against *Tomato Yellow Leaf Curl Virus*

4.6.1 Number of flowers/plant

Number of flowers/plant was significantly affected by *TYLCV* under different treatments (Table 7). Among the different treatments, T₆ (Silver mulch) performed best for number of flowers/plant 34.60 which was significantly different from all other treatments. It was also found that the treatment Control treatment (T₇) showed lowest number of flowers/plant 21.36 Among the treated plants, the lowest number of flowers/plant was achieved from T₄ (Neemax) ,23.22 which was statistically identical with T₅ (Multineem), 24.73.

4.6.2 Number of fruits/plant

Number of fruits/plant was significantly influenced by *TYLCV* performed against different treatments applied to the crop to control whitefly (Table 7). Results indicated that the treatment, T₆ (Silver colored mulch) produced highest number of fruits/plant (31.84) which was significantly different from all other treatments. The Control treatment (T₇) showed lowest number of fruits/plant (16.08) which was also significantly different from all other treatments. Among the treated plants, the lowest number of fruits/plant was found from T₄ (Neemax), 20.93.

4.6.3 Weight of fruits/plant

Weight of fruits/plant affected by *TYLCV* was significantly against different treatments applied to the crop (Table 7). Results revealed that the treatment, the highest weight of fruits/plant (3.08 kg) was found from the treatment, T₆ (Silver mulch) which was significantly different from all other treatments. The lowest weight of fruits/plant (1.24 kg) was produced from the Control treatment (T₇) which was also significantly different from all other treatments. Among the treated plants, the lowest weight of fruits/plant was found from T₄ (Neemax)

4.6.4 Single fruit weight

The *TYLCV* transmitted by whitefly had significant influence on single fruit weight of tomato against different treatments applied (Table 7). The treatment, T₆ (Silver mulch) produced highest single fruit weight (95.68 g) followed by T₁ (Imidacloprid) which was significantly different from all other treatments. The lowest single fruit weight (76.75 g) was produced from the Control treatment (T₇) which was also significantly different from all other treatments. Among the treated plants, the lowest single fruit weight (90.25 g) was found from T₄ (Neemax) which was statistically identical with T₃ (Subicron).

4.6.5 Average fruit diameter

Whitefly transmitted virus, *TYLCV* significantly influenced the average fruit diameter of tomato against different treatments (Table 7). The treatment, T₆ (Silver mulch) exhibited the highest average fruit diameter (14.57 cm) followed by T₁ (Imidacloprid) which was significantly different from all other treatments. The lowest average fruit diameter (8.04) was found from the Control treatment (T₇). Among the treated plants, the lowest single fruit weight (10.15 cm) was found from T₄ (Neemax) which was statistically identical with T₅ (Multineem).

Table 7. Effect on yield and yield contributing parameters of tomato through management of tomato yellow leaf curl virus using different treatments

	Yield and yield contributing parameters				
	Number of flowers/plant	Number of fruits/plant	Weight of fruits/plant (kg)	Single fruit weight (g)	Average fruit diameter (cm)
T ₁	32.12b	29.39 b	2.74 b	93.17 b	13.67 b
T ₂	30.63b	28.10 c	2.57 c	91.57 c	12.91 c
T ₃	27.24c	25.80 d	2.34 d	90.52 d	11.47 d
T ₄	23.22d	20.93 f	1.89 f	90.25 d	10.15 e
T ₅	24.73d	22.26 e	2.03 e	91.43 c	10.49 e
T ₆	34.60a	31.84 a	3.08 a	95.68 a	14.57 a
T ₇	21.36e	16.08 g	1.24 g	76.75 e	8.04 f
LSD _{0.05}	1.617	0.614	0.116	0.327	0.415
CV (%)	8.636	6.421	4.713	8.334	6.528

T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = Silver colored mulch, T₇ = Control

5. Fruit yield

Fruit yield/ha performed best from the treatment, T₆ (Silver colored mulch) against *TYLCV* and found significant variation among all the treatments (Fig. 4 and Appendix X). Results exposed that the treatment, T₆ (Silver colored mulch) gave highest fruit yield (53.35 t/ha) where the lowest fruit yield (21.41 t/ha) was produced from Control treatment (T₇). The second highest and third highest fruit yield (47.49 and 44.51 t/ha respectively) was achieved from T₁ (Imidacloprid) and T₂ (ACmix) respectively. Among the treated plants, the lowest fruit yield (10.15 t/ha) was found from T₄ (Neemax) which was statistically identical with T₅ (Multineem).

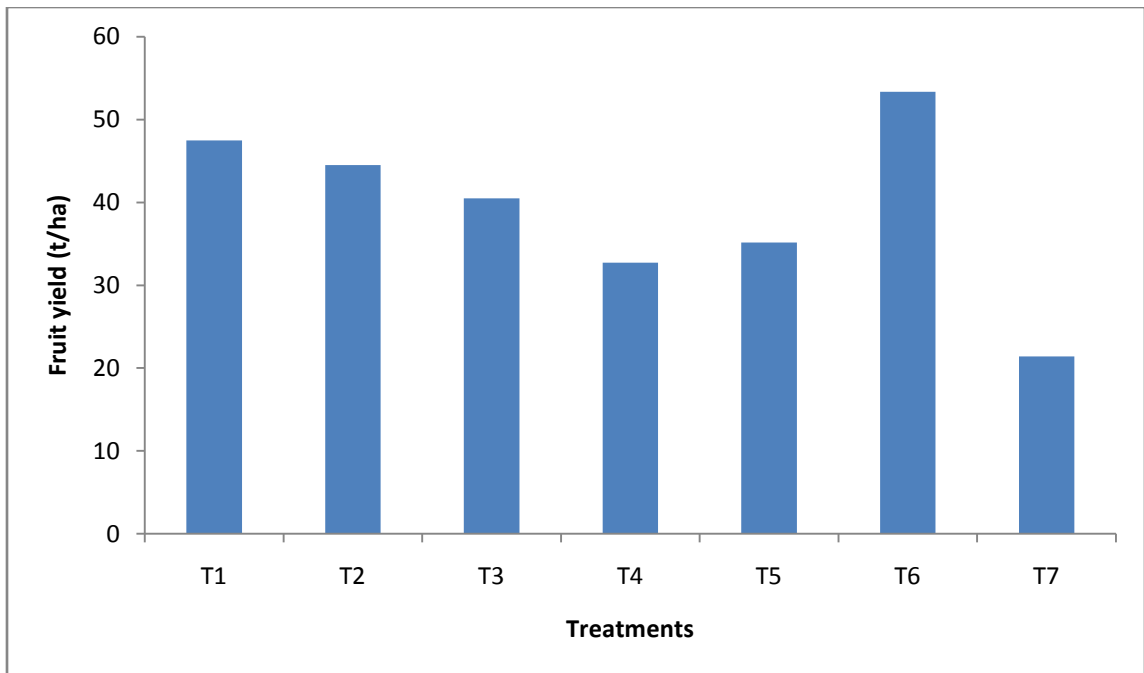


Fig. 8. Effect on yield of tomato using different treatments against *Tomato Yellow Leaf Curl Virus*

T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = Silver colored mulch, T₇ = Control

DISCUSSION

The present study was undertaken to study the management of *tomato Yellow Leaf Curl Virus* through insecticides, botanicals and Silver colored mulch. Considerable control of *Tomato Yellow Leaf Curl Virus* was observed through different management practices. Different treatment were used *viz.* T₁ = Imidacloprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = Silver colored mulch, T₇ = Control for the management of *Tomato Yellow Leaf Curl Virus*. Under the present study, T₆ (Silver colored mulch) gave the best performance against *Tomato Yellow Leaf Curl Virus*.

5.1 Disease incidence and severity (%):In this study, it was noticed that the disease incidence (%) due to *Tomato Yellow Leaf Curl Virus* was found in almost all the plots at 80 DAS. The highest percentage disease incidence was recorded in the T₇ (control treatment) followed by T₄, T₅, T₃, T₂, T₁ and T₆ respectively. Among the treatments, the lowest disease incidence (%) was found in T₆ (Silver colored mulch). The results regarding the percent disease incidence of *TYLCV* in tomato that observed in this study is almost similar as observed by Sastry and Singh (1973), Al-Musa (1982), Green and Kalloo (1994) and Ali *et al.* (2005). Azam *et al.* (1997) investigated that insecticides (carbofuran, endosulfan, Dimethoate, buprofezin and triazophos) and cultural methods are most effective for the control of whitefly and *TYLCV* in tomatoes. Ahmed *et al.* (2001) used Imidacloprid insecticide, for indirectly controlling *TYLCV* through control of its vector whitefly.

Vani *et al.* (1989) evaluated that different type of mulches reduced the incidence of viral diseases and also found that mulching increased plant growth and yield. Mulching with polyethylene sheets of different colours (black, transparent, white, and aluminum) had significant effect on *Bemisia tabaci* and spread of *TYLCV*. Polyethylene mulching reduced whitefly and *TYLCV* infection. But mulching with

aluminum colour was found to be most effective in reducing number of whiteflies and delay the infection of *TYLCV* (Davino *et al.*, 1996).Azam *et al.* (1997) and also investigated that insecticides (carbofuran, endosulfan, Dimethoate, buprofezin and triazophos) and cultural methods (covering the plants with polyester for 30, 45 or 60 days) are most effective for the control of whitefly and *TYLCV* in tomatoes.

5.2. Morphological features

5.2.1. Number of, flowers and fruits per plant

The yield of individual treatment depends on the number of flowers and fruits per plant. The highest number of flower and fruits per plant were recorded in T₆(Silver colored mulch) (followed by T₁, T₂ and T₃. But considering the economic condition/cost-benefit ratio T₅ (Multineem)was the best in both block, because there was no significant different among T₄ (Neemax), regarding these mentioned parameters. The lowest number of leaves, flowers and fruits per plant were recorded in the T₇ (control treatment). The same results were found in the previous study that was conducted by Sayed *et al.*, (2018). The finding of the previous work was Imidacloprid gave the better results regarding the flowers and fruits/plant than Sobicron.

5.3. Yield and yield contributing characters

The highest yield per plant and plot was recorded in T₆ (Silver colored mulch) followed by the treatment T₁, T₂,T₃,T₅ ,T₄ and T₇ (control treatment). But considering the economic condition/cost-benefit ratio T₅was the best in the study, because there was no significant different among T₄, T₅, regarding yield per plant and plot. Where the lowest yield per plant/plot was founded in T₇ (control treatment). The same results were found in the previous study that was conducted by Sayed *et.al*, (2018). There is no more previous report over yield of okra against *YVMV* in our country.

5.4. Growth promoting parameters

5.4.1. Number of branches per plant

The highest number of branches were recorded in T₆ (Silver colored mulch) followed by T₁, T₂ and T₃. But considering the economic condition/cost-benefit ratio T₅ (Multineem) was the best in both block, because there was no significant different among T₄ (Neemax) regarding these mentioned parameters. The lowest number of leaves, flowers and fruits per plant were recorded in the T₇ (control treatment). The same results were found in the previous study that was conducted by Sayed *et al.*, (2018). The finding of the previous work was Imidacloprid better than Sobicron.

5.4.2. Plant height

The highest height of plant was recorded in T₆ (Silver colored mulch) followed by T₁, T₂ and T₃. But considering the economic condition/cost-benefit ratio T₅ (Multineem) was the best in both block, because there was no significant different among T₄,(Neemax) regarding these mentioned parameters. The lowest number of leaves, flowers and fruits per plant were recorded in the T₇ (control treatment). The same results were found in the previous study that was conducted by Sayed *et al.*, (2018). The finding of the previous work was Imidacloprid better than Sobicron.

CHAPTER V

SUMMARY AND CONCLUSION

The experiment was conducted to manage the *Tomato Yellow Leaf Curl Virus* through selected insecticides, botanicals and Silver colored mulch. The study was conducted in the field allowed for the Department of Plant Pathology of Sher-e-Bangla Agricultural University, Dhaka during winter season. High yielding tomato variety BARITomato-14 was used. The field experiment was carried out in randomized complete block design with three replications. In total 7 treatments including control were considered viz. T₁ = Imidachloroprid, T₂ = ACmix, T₃ = Subicron, T₄ = Neemax, T₅ = Multineem, T₆ = Silver metallic plastic mulch and T₇ = Control.

In case of % disease incidence of *TYLCV*, the lowest disease incidence was recorded 11.11%, 13.33% and 13.33% at 30, 45 and 60 DAT respectively was recorded in T₆. The highest disease incidence 33.33, 40.00 and 46.67% at 30, 45 and 60 DAT respectively was recorded in T₇ (control). In case of disease severity, the lowest disease severity (1.96%, 2.01% and 1.72% at 30, 45 and 60 DAT respectively) was also recorded in T₆ (Silver mulch) and the highest disease severity (25.80, 27.43, 46.22%) was also found in T₇ treatment. All the selected treatments used in the study for controlling the insect vector, whitefly of *TYLCV* were showed significant influence on different assayed parameters in the test crop tomato. Among the set treatments, T₆ (Silver mulch) was gave the best performance in all regards of assayed parameter, the best performance was the reduced whitefly association in remarkable level. In case of insect vectors, whitefly association, the whitefly association was the lowest in T₆ treatment (Silver mulch) there were 2.0, 4.0 and 2.0 at 30, 45 and 60 DAT respectively. The highest whitefly association was recorded in T₇ (control treatment) that was 19, 23 and 18 at 30, 45 and 60 DAT respectively.

Among the set treatments, T₆ (Silver mulch) gave the best results regarding growth promoting characters, yield and yield attributes. The highest plant height (74.93, 94.05 and 99.21 cm at 30, 45 and 60 DAT respectively), number of healthy leaves/plant (34.07, 98.31 and 95.50 at 30, 45 and 60 DAT respectively), number of branches/plant (6.20, 7.21 and 8.07 at 30, 45 and 60 DAT respectively), number of flowers/plant (0.37, 5.31 and 34.60 at 30, 45 and 60 DAT respectively), number of fruits/plant (31.84), weight of fruits/plant (3.08 kg), single fruit weight (95.68 g), average fruit diameter (14.57 cm) and fruit yield (53.35 t/ha) were obtained from the treatment, T₆ (Silver mulch). Again, the lowest plant height (60.87, 75.76 and 79.99 cm at 30, 45 and 60 DAT respectively), number of healthy leaves/plant (27.20, 51.75 and 45.15 at 30, 45 and 60 DAT respectively), number of branches/plant (7.00, 7.33 and 5.22 at 30, 45 and 60 DAT respectively), number of flowers/plant (0.13, 3.44 and 21.36 at 30, 45 and 60 DAT respectively), number of fruits/plant (16.08), highest weight of fruits/plant (3.08 kg), weight of fruits/plant (1.24 kg), single fruit weight (76.75 g), average fruit diameter (8.04) and fruit yield (21.41 t/ha) were obtained from control treatment (T₇).

Among the treated plants, the results on different parameters, achieved lower performance from botanical treatment T₄ (Neemax) and T₅ (Multi Neem) in most of the cases where the treatment T₁ (Imidacloprid) showed comparatively better performance next to treatment T₆ (Silver mulch). From the above findings on different parameters studied, it can be concluded that the treatment T₆ (Silver colored mulch) was best against whitefly association in test crop tomato against *TYLCV* for tomato cultivation compared to other considering treatments including control treatment.

REFERENCES

- Abdelbacki, A. M., Soad, H. T., Sitohy, M. Z., Dawood, A. I., Abd-El, H. M. M., Rezk, A. A. 2010. Inhibition of Tomato Yellow Leaf Curl Virus (TYLCV) using whey proteins. *Virologica J.* 7:26.
- Abdel-Salam. 1990. Mechanical transmission of two Egyptian isolates of beet curly top and tomato yellow leaf curl viruses. *Bulletin of Faculty of Agriculture University of Cairo* 41: 825-842.
- Abdel-Salam. 1991. Tomato yellow leaf curl virus in Egypt-characterization, partial purification and antiserum production. *Bulletin of the Faculty of Agriculture, University of Cairo*, 42(2): 507-520.
- Ahmed, N. E., Kanan, H. O., Sugimoto, Y., Ma, Y.Q., and Inanaga, S. 2001. Effect of imidacloprid on incidence of Tomato yellow leaf curl virus. *Plant Dis.* 85: 84–87.
- Ahmed, N.E., FI.Q. Kanan.,Y.Sugimoto.,Y.Q.Ma and S. Inanaga. 2001.Effect of imidacloprid on incidence of Tomato yellow leaf curl virus *Plant Dis.*85:84-87.
- Ahmed, N.E., Mohamed, M.B., and Rahman, A.A. 1996. Effect of *Conundrum sativum* as a repellent to whitefly on tomato yellow leaf curl virus and yield in farmers fields. Final report pp-121. In integrated pest management FAO/ARC/ (GCP/ SUD/ 025/ NET).
- Akanda, A. M., K,Tsuno and S, Wakimoto. 1991b. Serodiagnosis of viruses infecting some crops of Bangladesh. *J. Fac. Agr. Kyshu Univ.* 35: 121-129.

- Akanda, A.M. 1991. Studies on the virus and mycoplasma disease of vegetable crops in Bangladesh. A thesis for the partial fulfilment of Doctor of Agriculture. Submitted to the Plant Pathology Laboratory. Department of Agronomy, Faculty of Agriculture, Kyushu University, Fukuoka, Japan 181 pp.
- Akanda, A.M., K,Tsuno and S,Wakimoto. 1991a. Serological detection of four plant viruses in cucurbitaceous crops from Bangladesh. Ann. Phytopath. Soc. Japan 57: 499-505.
- Alam, N., A. Khair and A.M. Akanda. 1994. Biochemical alteration due to infection of yellow leaf curl virus in tomato. Bangladesh J. Life Sci. 6:15-22.
- Al-ani, R. A., Adhnab, M. A., Hamad S. A. H., and Diwan S. N. H. 2011. Tomato yellow leaf curl virus, identification, virus vector relationship, strain characterisation and a suggestion for its control with plant extracts in Iraq. African J. Agric. Res. 6 (22): 5149-5155.
- Ali, S., Khan, M.A., Habib, A. and Rasheed, S. (2005). Correlation of environmental conditions with okra yellow vein mosaic virus and *Bemisia tabaci* population density. Journal of Agricultural Biolpgy. 7:142-144.
- Al-Ilitty, A. and II.L. Sharif. 1987. Studies of host plant performance of *Bemisia tabaci* (Genn) on some crops and effect of using host on the trap, spread of tomato yellow leaf curl virus to tomato in the plastic house. Araf J. Plant Prot.: 19-23.
- Al-Musa, A. 1982. Incidence, economic importance and control of tomato yellow leaf curl in Jordan. Plant Dis. 66:561-563.

- Antignus, Y., Perslman, M., Ben-Joseph, .R. and Cohen, S. 1994. Association of nucleic acid and coat protein of tomato yellow leaf curl virus (TYLCV) with the whitefly vector *Bemisia tabaci*. In: 1st International Symposium on Geminiviruses. September 14-17, 1994. Almeria, Spain. 66 p.
- Arunakumara, K. T., Kulkarni, M. S., Thammaiah, N. and Hegde, Y. 2010. Fungicidal management of early blight of tomato. *Indian Phytopath.* 63 (1): 96-97.
- Avidov (Klein) Z. 1940. *Bemisia tabaci* in Israel. *Hassadeh* 20: 84–86.
- Azam, K.M., S.A. Razvi, Zoub- Ali and A.A. Al Rauesi. 1997. Management of whitefly (*Bernina tabai* Gennadius) and tomato leaf curl virus in tomato crops. *Indian J. Plant Protect.* 25:1, 36-41.
- Bajapai, N. K. 2005. Use of viricide for management of chilli leaf curl virus, *Spice India* March 2005: 15-16.
- BBS (Bangladesh Bureau of Statics). (2016). *Statistical Pocket Book of Bangladesh*. Statistics Division, Ministry of Planning, Govt. of the People's Republic of Bangladesh, Dhaka. p.215.
- Block, K.R. (1982). Geminivirus diseases. *Plant Dis.* 1982; 66:266-270.
- Bosco, D. 1993. Epidemiological studies on tomato yellow leaf curl bigemini virus (TYLCV) in Italy. *Informatore-Fitopatologico* 43: 33-36.
- Brown, K. and J. Bird. 1992. Whitefly-transmitted geminiviruses and associated disorders in the Americas and the Caribbean Basin, *Plant Dis.* 76:220-225.
- Brunt, A., K. Crabtree and A. Gibbs. (Edt.) 1990. Tomato yellow leaf and Geminivirus. In *viruses of tropical plants*. CAB International, Wallingford, Oxon, OX-10 eight DE, UK. P. 578-580.

- Chandrashekharaiyah, G. Basana Gowda, Kumar, A. N. D. T. and Chakravarthy, A. K. 2013. Bio-efficacy of *Verticillium lecanii* against whitefly on greengram. *Insect Environ.* 19 (2): 83-84.
- Cherif, C. and M. Russo. 1983. Cytological evidence of the association of a Gemini virus with the tomato yellow leaf curl in Tunisia. *Phytopathologische Zeitschrift, INRAT, Ariana, Tunisia*, 108 (318): 221 - 225.
- Cohen, S. and F.E. Nitzany. 1966. Transmission and host range of the tomato yellow leaf curl virus, *Phytopath.* 56:1127-1131.
- Cohen, S., and I. Harpeiz. 1964. Periodic, rather than continual acquisition of a new tomato virus by its vector the tobacco whitefly (*Bemisia tabaci* Bennadius). *Entomol. Exp. Appl.* 7:155-166.
- Cohen, S., Y. Antignus., M. Lapidot., N. Mor and Y. Masika. 1998. The use of uv-absorbing plastic sheets to protect crops against insect and spread of virus disease. 14th international Congress of plastic in Agriculture. Tel Aviv, Isreal. 29-33.
- Csizinsky, A.A., Schuster, D.J., and Kring, J.B. 1995. Color mulches influence yield and insect pest populations in tomatoes. *J. Amer. Soc. Hort. Sci.* 120 (5): 778-784.
- Czosnek, H., R. Ber., Y. Antignus., S. Cohen., N. Navot and D. Zamir. 1988. Isolation of tomato yellow leaf curl virus, a geminivirus. *Phytopath.* 78: 508-512.
- Czosnek, H., Y. Zakay., N. Navot., M. Zeidan., N. Kedar., H. Rabinowitch and D. Zamir. 1990. Screening *Lycopersicon* accessions for resistance to tomato

- yellow leaf curl virus. Presence of viral DNA and symptom development. *Plant Dis.* 75:279-281.
- Dalmon, A. and G. Marchoux. 2000. Tomato yellow leaf curl virus host plants. *Phytoma.* 527: 14-17.
- Davino, M., Urso, F. D., Arciddia, R., Carbone, M., G. , G., and Durso, F. 1994. Investigation on the epidemiology of *Tomato yellow leaf curl virus* (TYLCV) in sicily. *Petria.* 4: 151-160.
- Denholm I, Gorman K, Jarvis W, Cahill M. (1996). Resolution of baseline response to buprofezin in *Bemisia tabaci* (Homoptera: Aleyrodidae). *Bulletin of Entomological Research.* 1996; 86(2):117-122.
- Dhanju, K.S. and J.P. Verma. 1987. Natural occurrence of mixed infection of tomato leaf curl virus and tobacco mosaic virus in tomato. *Indian journal of Virology.* Dept, of PI. Path. Naryana Agric. Univ., India. 2(1): 108-110.
- Gallitelli, D., E. Luisini., G.P
- Dikshit AK, Lal OP, Shrivastava YN. (2000). Persistence of pyrethroid and nicotinyl insecticides on okra fruits. *Pesticides Research Journal.* 2000; 12(2):227-231.
- Divakaran, A. 2007. Molecular characterisation of tomato with special reference to ToLCV resistance. M.Sc.(Ag) thesis, Kerala Agricultural University, Vellanikkara, Thrissur, 71p.
- FAO (Food and Agriculture Organization). (2016). *FAO Production Yearbook.* Basic Data Branch, Statistics Division, FAO, Rome, Italy. p. 289.
- Glick, E., Levy, Y., and Gafni, Y. 2009. The viral etiology of tomato yellow leaf curl disease – a review. *Plant Protect. Sci.* 45: 81–97.

- Goodman, T.M. 1981. Geminiviruses, *J. Gen. Virol.* 54:9-21.
- Green, S. K. 1998. Making leaf tissue squash on membranes for virus detection. Asian Vegetables Research and development Center. Shanhua, Taiwan 15pp.
- Green, S.K. and C. Kalloo. 1994. Leaf curl and yellowing disease of tomato. *In:* Technical Bulletin No. 21. Leaf curl and yellowing viruses of pepper and tomato. In an overview Asian Vegetable Research and Development Centre. 21-51 pp.
- Green, S.K. and Kalloo, G. (1994). Leaf curl and yellowing viruses of pepper and tomato: an overview. Technical Bulletin No. 21. Asian Vegetable Research and Development Center, Taiwan. 51p.
- Iersel, M. W. V., Ronald D. O. and Hall, D. B. 2000. Imidacloprid applications by subirrigation for control of silverleaf whitefly (Homoptera: Aleyrodidae) on Poinsettia, *J. Econ. Entomol.* 93(3): 813-819.
- Jayaraj, S., Rangarajan, G.A.V., Murugesan, S., Santharam, G., Vijayaraghavan, S. and Thangaraj, D. (1986). Outbreak of whitefly *Bemisia tabaci* (G.) on cotton in Tamil Nadu and its management. All Indian Group Discussion on Whitefly in Cotton, Guntur. P. 65-77.
- Jha, S.K. and Kumar, M. (2017). Relative efficacy of different insecticides against whitefly, *Bemisia tabaci* on tomato under field condition *Journal of Entomology and Zoology Studies*; 5(5): 728-732.
- Jorda, C., Font, I., Martinez, P., Juarez, M., Ortega, A., Lacasa, A. 2001. Current status and new natural hosts of *Tomato yellow leaf curl virus* (TYLCV) in Spain. *Plant Dis.* 85 (4): 445.

- Jorda, C., I. Font, P. Martinez, M. Juarez, A. Ortega and A. Lacasa. 2001. Current status and new natural hosts of Tomato yellow leaf curl virus (TYLCV) in Spain. *Plant Dis.* 85 (4): 445.
- Kalb, T. 2004. Fact sheet on *Tomato yellow leaf curl virus* (TYLCV). AVRDC publication 04-608. <http://www.avrdc.org>.
- Karthikeyan, G., Doraisamy, S., Rabindran, R. and Ganapathy, T. 2009. Evaluation of antiviral principles for the induction of systemic resistance in blackgram (*Vigna mungo*) against Urdbean Leaf Crinkle Virus. *Arch. Phytopathol. Pl. Prot.* 42(12): 1172–1186.
- Kegler, H. 1994. Incidence, properties and control of tomato yellow leaf curl virus. *Plant protect.* 29:119-132.
- Khan JA. and Ahmed J. (2005). Diagnosis, monitoring and transmission characteristics of cotton leaf curl virus. *Current Sci.* 2005; 88:1803-1809.
- Konate, G., N. Barrow., D. Fargette., M.M. Swanson., and B. D. Flarrison. 1995. Occurrence of whitefly transmitted geminivirus in crops in Burkina Faso and their serological detection and differentiation. *Ann. Appl. Biol.* 126: 121-129.
- Kontsedalov S, Abu-Moch F, Lebedev G, Czosnek H, Horowitz AR, Ghanim M. (2012). Bemisia tabaci Biotype Dynamics and Resistance to Insecticides in Israel During the Years 2008-2010. *J. Integr. Agric.* 2012; 11(2):312-320.
- Kung, S. 1999. Effect of net protected nursery on *Tomato yellow leaf curl virus* incidence and severity. Rc-AVRDC training reports 1999. 250p. ([http://www.arc-avrdc.org/pdffiles/kung \(17-N\) pdf](http://www.arc-avrdc.org/pdffiles/kung%20(17-N).pdf))-

- Lewandowski, D. J., Hayes, A. J., and Adkins, S. 2010. Surprising results from a search for effective disinfectants for Tobacco mosaic virus–contaminated tools. *Plant Dis.* 94:542-550.
- Lukyanenko, A.N. 1991. Disease Resistance in Tomato. *In:* Kalloo, G. (Ed.) 1991. Genetic Improvement of Tomato. Springer Verlag, Berlin Fleidelberg, Germany, p.99-119.
- Macintosh, S., D.J. Robinson and B.D. Harrison. 1992. Detection of three white fly transmitted geminiviruses occurring in Europe by tests with heterologous monoclonal antibodies. *Ann. Appl. Biol.* 121:297-303.
- Makkoub, K.M. 1978. A study on tomato viruses in the Jordan valley with special emphasis on tomato yellow leaf curl. *Plant Dis. Rept., University of Jordan, Amman*, 62(3): 259-262.
- Mansour, A. and A. Al-musa. 1992. Tomato yellow leaf curl virus : host range and virus vector relationships. *Plant pathology. Department of Plant Protection, Faculty of Agriculture, University of Jordan, Amman, Jordan.* 41:122-125.
- Martelli, G.P. and A. Quacquarelli. 1982. The present status of tomato and pepper viruses. *Acta Horticulturae* 127:39-64.
- Martinez Y., Quinones M., Fonseca D., Miranda I. 2003. National surveys of begomovirus in tomato crop in Cuba. *Revista de Proteccion Vegetal*, 18:168–175.

- Matin, M.A., Haq, M.A., Karim, M.R. and Baksha, E. (1996). Farm Level Yield Analysis of Tomato Cultivation in Selected Areas of Bangladesh: An Economic Profile. *Bangladesh J. Agril. Res.* 21(1): 50-57.
- McGlashan, D., J.Z. Polston and D. Bois. 1994. Tomato yellow leafcurl virus geminivirus in Jamaica. *Plant Dis.* 78:1219.
- McGrath, P.F. and B.D. Harrison. 1995. Transmission of tomato leaf curl Gemini viruses by *Bemisia tabaci*: effect of virus isolate and vector biotype. *Ann. Appl. Biol.* 126: 307-316.
- Meena, R. P. 2012. Bio-efficacy of plant extracts for management of purple blotch disease of onion (*Allium cepa*). *Indian Phytopath.* 65 (3) : 253-257.
- Mehta P, Wyman JA, Nakhla MK, Maxwell DP. (1994). Transmission of tomato yellow leaf curl Gemini virus by *Bemisia tabaci* (Homoptera: Aleyrodidae). *Journal of Economic Entomology.* 1994; 87:1291-1297.
- Mishra NC, Mishra SN. (2002). Impact of biopesticides on insect pest and defenders on okra. *Indian Journal of Plant Protection.* 2002; 30:99-101.
- Mishra, S., Jagadeesh , K .S., Krishnaraj, P. U., Byadagi, A. S. and Vastrad, A. S. 2012. Biological management of Tomato leaf curl virus (ToLCV) by inducing systemic resistance through Rhizobacterial strains. *Bioinfolet* 9 (3): 417 – 427.
- Mishra, S., Jagadeesh, K. S., Krishnaraj, P. U. and Prem, S. 2014. Biocontrol of tomato leaf curl virus (ToLCV) in tomato with chitosan supplemented formulations of *Pseudomonas* sp. under field conditions. *Australian J. Crop Sci.* 8(3):347- 355.

- Momol, T.M., and Pemezny, K.L. 2006. Specific Common Diseases and Florida Plant Disease Management Guide, Department of Plant Pathology. University of Florida. Florida Cooperative Extension Service, I FAS. PDMG. 3: 53. <http://cdis.ifas.ufl.edu>.
- Moriones, E.J. Arno, A. Accotto and L. Cavallaarin. 1993. Firstreport of tomato yellow leaf curl in Spain. *Plant Dis.* 77(9): 953.
- Murad,G., M.Shai and H.Czosnek.2001 .Rate of Tomato yellow leaf curl virus translocation in the circulative transmission pathway of its vector, the whitefly *Bemisia tabaci*.*Phytopath.*91 (2): 188-196.
- Naik, R. G., Muniyappa, V. and Colvin, J. 2004. Studies on the relationship of tomato leaf curl geminivirus on tomato with its vector *Bemisia tabaci* Genn. *Agric. Sci. Digest*, 24 (1): 17 – 20.
- Naik, R. H., Devakumar, N. , Rao, G. E., Vijaya, N., Khan, H. S. I. and Subha, S. 2012. Performance of botanical and fungal formulation for pest managementin organic okra production system. *J. Biopest.* , 5:12-16.
- Navot, N.,R. Ber.,11. Czosnek. 1989. Rapid detection of tomato yellow leaf curl virus in squashes of plants and insect vectors. *Phytopathology* 79: 562-586.
- Nayar KK, Ananthkrishnan TN, David BV. (1992). *General and Applied Entomology*, 1992, 430-431.
- Nitzany, F.E. 1975. *Tomato yellow leaf curl virus*. *Phytopathologia Meditcrranca* 14: 127- 129.

- Oliveria MRV, Henneberry TJ, Anderson P, Naranja SE, Elsworth PC. (2001). History, current status and collaborative research prospects for *B. tabaci*. *Crop Prot.* 2001; 20(9):709-723.
- Pilowsky, M and S. Cohen. 1990. Tolerance of tomato yellow leaf curl virus derived from *Lycopersicon peruvianum*. *Plant Disease*. Department of Plant Genetic and Breeding, Agricultural Research Organization, Volcani Centre, Bet Dagan, Israel, 74(3): 248-250.
- Pilowsky, M., S. Cohen., R. Ben-Joseph., S. Nahon and L. Stamova. 1993. Effect of tomato yellow leaf curl virus on tolerant and susceptible cultivates. *Proceeding of the XIIth Eucarpia meeting on tomato genetics and breeding, Plovdiv, Bulgaria, 27-31 July 1993.*
- Pilowsky, M. and S. Cohen. 2000. Screening additional wild tomatoes for resistance to the whitefly-borne tomato yellow leaf curl virus. *EUCARPIA TOMATO '2000. XIV meeting of the EUCARPIA Tomato Working Group, Warsaw, Poland, Acta-Physiologiae-Plantarum.* 22 (3): 351-353
- Polizzi, C., C. Asero., K.E. Cockshull., Y. Tuzel and A. Gul. 1994. Epidemiology and incidence of tomato yellow leaf curl virus (TYLCV) in greenhouse protected by screen in Italy. *Acta Hort.*, 366:345-552.
- Polston, J. E. and Lapidot, M. 2007. Management of Tomato Yellow Leaf Curl Virus: US and Israel Perspectives; In: *Tomato Yellow Leaf Curl Virus Disease*, H. Czosnek (ed.), Springer publications, pp 251–262.
- Polston, J.E. and P.K. Anderson. 1997. The Emergence of whitefly, transmitted Geminiviruses in Tomato in Western Hemisphere. *Plant Dis.* 12:1358-1369.

- Polston, J.E. and P.K. Anderson. 1997. The Emergence of whitefly, transmitted Geminiviruses in Tomato in Western Hemisphere. *Plant Dis.* 12:1358-1369.
- Polston, J.E., D.Bois., C.A.,Serra., and S.concepcion. 1994. First report of a tomato yellow leaf curl like geminivirus in the Western Hemisphere *Plant Dis.* 78:831.
- Prasannath, K., Dharmadasa, K.N.P., De Costa, D.M. and Hemachandra, K.S. (2014). Variations of Incidence, Types of Virus Diseases and Insect Vector Populations of Tomato (*Solanum lycopersicum* L.), Grown in Different Agro-ecological Regions of Sri Lanka under Two Crop Management Systems. *Tropical Agricultural Research.* Vol. 25 (3): 376– 395.
- Pun, K.B., Sabitha, D. and Balasubramaniam, G.M. (2005). Prediction of white fly population and okra yellow vein mosaic virus disease incidence in okra. *Indian Journal of Virology.* 16:19-23.
- Rajasri, M., Vijayalakshmi, K., Malathi, V., Sujatha, M. and Rao, R.D.V.J. P. 2011. Transmission and Molecular Characterization of Tomato leaf curl virus in Andhra Pradesh, South India. *Proc. IIIrd IS on Tomato Diseases*, Eds.: A. Crescenzi and A. Fanigliulo *Acta Hort.* 914, ISHS.
- Ramu, P. S., Rao, N. V., Narayana, N. H. S. and Reddy, V. C. 2011. Evaluation of insecticides to control whitefly, *Bemesia tabaci* vector of yellow vein mosaic disease on mesta. *Indian J. Pl. Prote.* 39. (1):1-3.

- Rashid, M. H., Hossain, I., Alam, M. S., Zaman, M. M. and Hannan, A .2008. Study on virus-vector relationship in TYLCV of tomato. *Int. J. Sustain. Crop Prod.* 3(1): 1-6.
- Reddy, B. A. 2006. Molecular characterisation, epidemiology and management of tomato leaf curl virus (ToLCV) in Northern Karnataka. Ph.D. thesis, University of Agricultural Sciences, Dharward,117p.
- Rishi, N. 2004. Current status of begomoviruses in the Indian subcontinent. *Indian Phytopath.* 57 (4) : 396-407.
- Saikia, A.K. and Muniyappa, V. (1989). Epidemiology and control of tomato leaf curl virus in southern India. *Tropical Agric.*, 66(4): 350-354.
- Samiyappen, R. 2003. Molecular mechanism involved in the PGPR mediated suppression of insect pest and plant pathogens attacking major agricultural and horticultural crop in India. Proceedings of 6th International workshop on Plant Growth Promoting Rhizobacteria, Calicut, India. Indian Institute of Spices Research, Calicut,India.
- Sastry, K.S.M. and S.J. Singh. 1973. Assessment of losses in tomato leaf curl virus. *Indian J. MycoL* 3:50-54.
- Satpathy S, Rai S, De N, Singh AP. (2004). Effect of insecticides on leaf net carbon assimilation rate and pest incidence in okra. *Indian Journal of Plant Protection.* 2004; 32:22-25.
- Sayed, A. (1988). Effect of the date of planting and insecticidal spray on the control of yellow vein mosaic of Lady's finger. M. Sc. Thesis, Department of Plant Pathology, B. A. U. Mymensingh 164.
- Sharfudin, A.F.M. and Siddique, M.A. (1985). *Sabji Bigyan.* 1st Edition, Bangladesh Agricultural University, Mymensingh. p. 4.

- Simone, G. W., and Momol, T. M. 2001. Vegetable Disease Recognition and Management - Florida Greenhouse Vegetable Production I landbook, Vol 3. One of a series of the Horticultural Sciences Department, University of Florida. Florida Cooperative Extension Service, IFAS MS.797. <http://edis.ifas.ufl.edu>.
- Singh, S.J. and K.S.M. Sastry. 1979. Control of the spread of tomato leaf curl virus by controlling the whitefly population. *Indian J. Hort.*, 31(2): 50-54.
- Sinistera, X., Patte, C.P., Siewnath, S. and Polston, J.E. (2000). Identification of Tomato yellow leaf curl virus-Is in the Bahamas. *Plant Disease* 84(5): 592.
- Sohrab, S. S., Karim, S., Varma, A., Abuzenadah, A. M., Chaudhary, A. G., Damanhour, G. A. and Mandal, B. 2013. Characterization of Tomato Leaf Curl New Delhi Virus infecting cucurbits: Evidence for sap transmission in a host specific manner. *African J. Biotech.* Vol. 12(32): 5000-5009.
- Subba, B., Pal, S., Mandal, T. and Kr. Ghosh, S. (2017). Population dynamics of whitefly (*Bemisia tabaci* Genn.) infesting tomato (*Lycopersicon esculentus* L.) and their sustainable management using biopesticides. *Journal of Entomology and Zoology Studies.* 5(3): 879-883.
- Tahir, M., Tariq, M., Mahmood, H.T., and Hussain, S. 2004. Effect of sowing dates on incidence of cotton leaf curl virus on different cultivars of cotton. *Plant Pathol. J.* 3(2): 61-64.
- Thanapase, W., P. Poorpol and S. Attathom. 1983. Causal agent and some important characters of tomato leaf curl disease, *Kasetsart J. Natural Sci.* 17(1): 80-89.
- Tomlinson, J.A. 1987. Epidemiology and control of virus disease of vegetables. *Ann. Appl. Biol.* 110:661-681.

- Vani, S., Varma, A., Mora, T.A., and Srivastava, K.P. 1989. Use of mulches for the management of mosaic diseases in muskmelon. *Indian Phytopathology* 42 (2): 227-235.
- Vasudeva, R. S. and Samraj, J. 1948. A leaf curl disease of tomato. *Phytopathology* 38: 364–369.
- Verma, A.K., D. Basu., P.S. Nath., S. Das., S.S. Ghatak and S. Muthapadhyak. 1989. Relationship between the population of whitefly, *Bemisia tabaci* and the incidence of tomato leaf curl virus disease. *Indian Journal of Mycological research*. Plant Virus Research Centre, Department of Plant Pathology, Bidhan Chandra Krishi Viswavidyalaya, Kalyani, West Bengal.
- Wongklom, W. 1999. Effect of physical barrier on the control of Tomato yellow leaf curl virus and its vector (whitefly). *Arc-AVRDC training Reports*. 250p. ([http://www.Arc-AVRDC.Org/pdf_files/wongklom\(17-n\).pdf](http://www.Arc-AVRDC.Org/pdf_files/wongklom(17-n).pdf)).
- Xienqui, Y. 2000. Effect of intercropping on *Tomato yellow leaf curl virus* incidence. *Arc- avrdc training Reports* 205p. (http://www.Arc-AVRDC.Org/pdf_files/023-yin-18th.pdf).
- Yadav, K. 2011. Incorporation of ToLCV resistance in bacterial wilt resistant varieties. Ph.D. (Hort) thesis, Kerala Agricultural University, Thrissur, 213p.
- Zacharia, R. M. 2006. Characterisation and management of bitter melon distortion mosaic virus. Ph.D. Thesis, Kerala Agricultural University, Thrissur. 72 p.
- Zeidan, M. and II.Czosnek.1991. Acquisition of yellow leaf curl virus by the White fly *Bemisia tabaci*. *J. Gen. Virol.* 72: 2607-2614.

APPENDICES

Appendix I. Agro-Ecological Zone of Bangladesh showing the experimental location

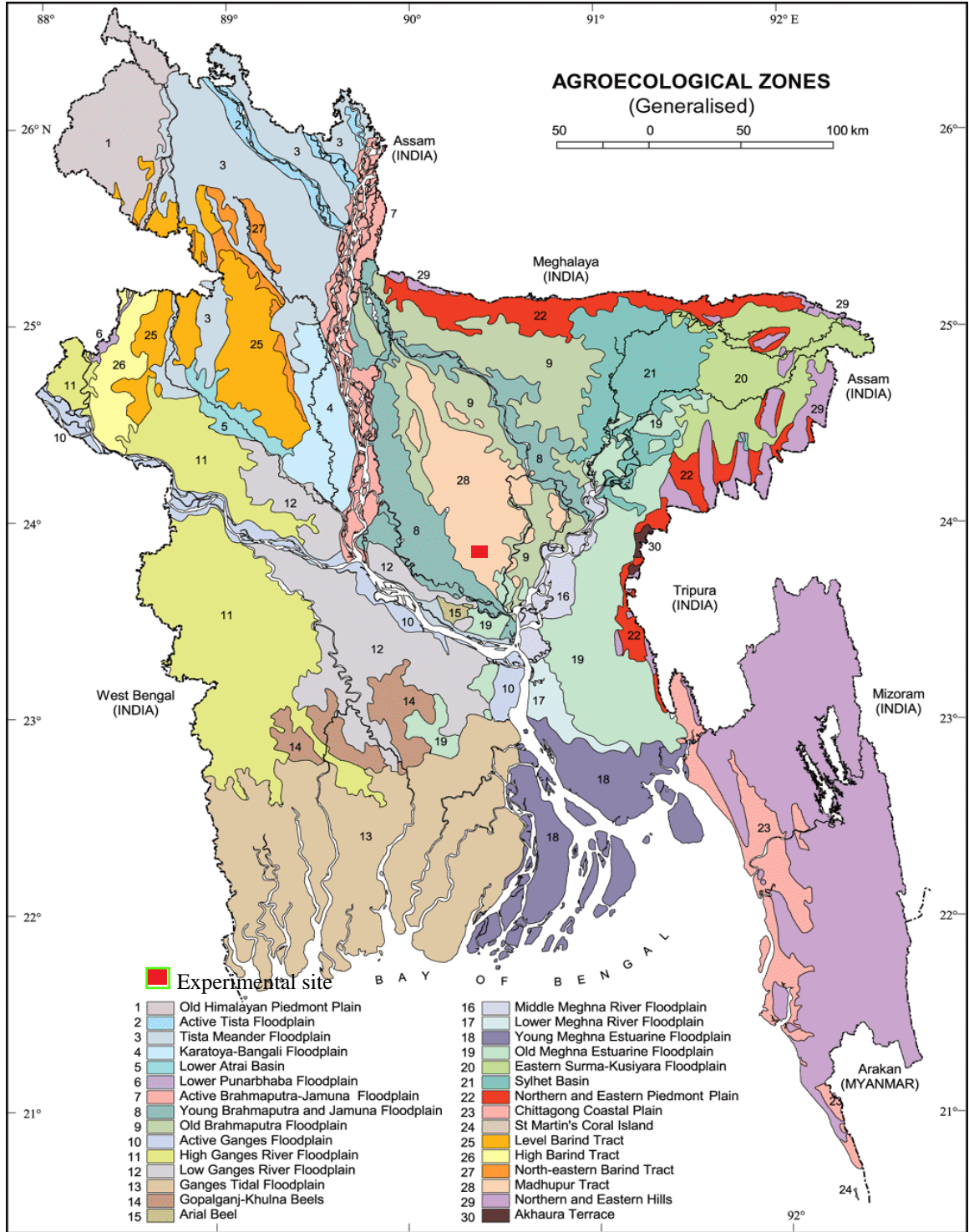


Fig. 10. Experimental site

Appendix II. Monthly records of air temperature, relative humidity, rainfall and sunshine hours during the period from October 2016 to March 2017

Month	RH (%)	Air temperature (C)			Rainfall (mm)
		<i>Max.</i>	<i>Min.</i>	<i>Mean</i>	
October, 2016	68.48	30.42	16.24	23.33	52.60
November, 2016	56.75	28.60	8.52	18.56	14.40
December, 2016	54.80	25.50	6.70	16.10	0.0
January, 2017	46.20	23.80	11.70	17.75	0.0
February, 2017	37.90	22.75	14.26	18.51	0.0
March, 2017	52.44	35.20	21.00	28.10	20.4

Source: Bangladesh Meteorological Department (Climate division), Agargaon, Dhaka-1212.

Appendix III. Characteristics of experimental soil analyzed at Soil Resources Development Institute (SRDI), Farmgate, Dhaka.

A. Morphological characteristics of the experimental field

Morphological features	Characteristics
Location	Agronomy Farm, SAU, Dhaka
<i>AEZ</i>	Modhupur Tract (28)
General Soil Type	Shallow red brown terrace soil
Land type	High land
Soil series	Tejgaon
Topography	Fairly leveled
Flood level	Above flood level
Drainage	Well drained
Cropping pattern	Not Applicable

Source: Soil Resource Development Institute (SRDI)

B. Physical and chemical properties of the initial soil

Characteristics	Value
Partical size analysis % Sand	27
%Silt	43
% Clay	30
Textural class	Silty Clay Loam (ISSS)
pH	6.7
Organic carbon (%)	0.45
Organic matter (%)	0.78
Total N (%)	0.03
Available P (ppm)	20
Exchangeable K (me/100 g soil)	0.1
Available S (ppm)	45

Source: Soil Resource Development Institute (SRDI)

Appendix IV. Layout of the experiment field

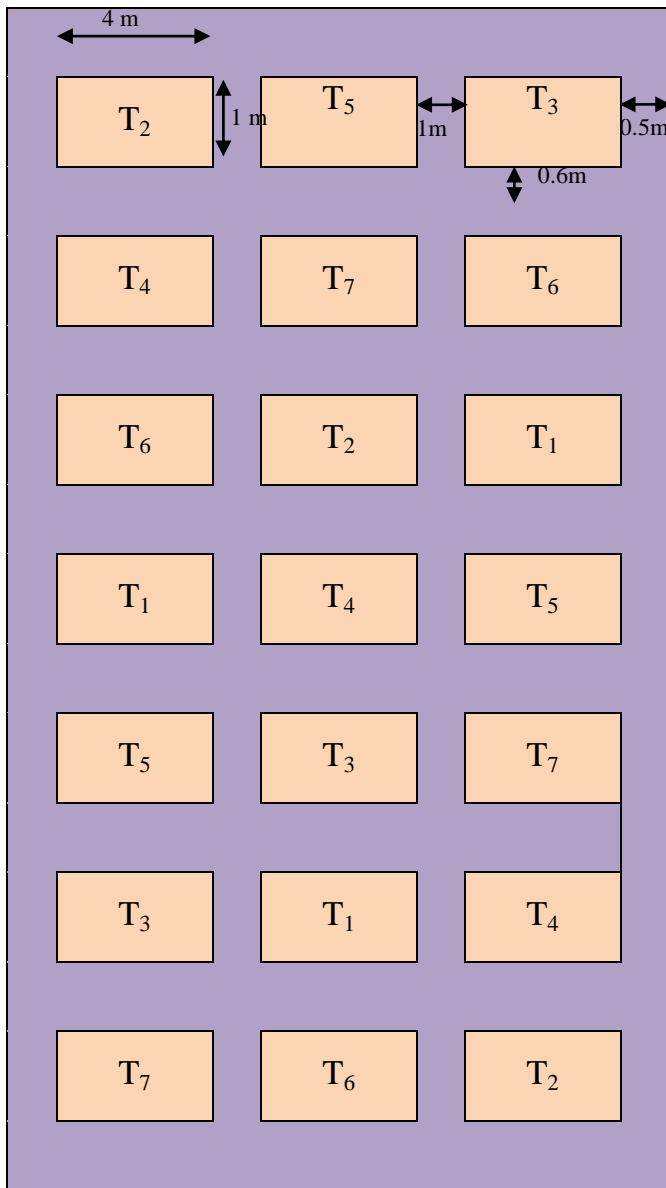


Fig. 11. Layout of the experiment field

Appendix v. Pictorial view of treatment plots



Fig 12. Plot treatment with silver mulch material