

**IDENTIFICATION OF INSECT PESTS IN SWEET POTATO AND
THEIR MANAGEMENT**

MD. ANWAR-UL-HAQUE LIMON



**DEPARTMENT OF ENTOMOLOGY
SHER-E-BANGLA AGRICULTURAL UNIVERSITY
DHAKA-1207**

JUNE, 2017

**IDENTIFICATION OF INSECT PESTS IN SWEET POTATO AND
THEIR MANAGEMENT**

BY

MD. ANWAR-UL-HAQUE LIMON

REG. NO. : 10-03802

A Thesis

*Submitted to the Faculty of Agriculture
Sher-e-Bangla Agricultural University, Dhaka
in partial fulfillment of the requirements
for the degree
of*

MASTER OF SCIENCE (MS)

IN

ENTOMOLOGY

SEMESTER: JANUARY-JUNE, 2017

Approved by:

Professor Dr. Mohammed Ali

Department of Entomology
Sher-e-Bangla Agricultural University
University

Supervisor

Professor Dr. Abdul Latif

Department of Entomology
Sher-e-Bangla Agricultural

Co-supervisor

Dr. Mst. Nur Mohal Akhter Banu

Associate Professor

Chairman

Department of Entomology and
Examination Committee



DEPARTMENT OF ENTOMOLOGY

Sher-e-Bangla Agricultural University
Sher-e-Bangla Nagar, Dhaka-1207

Memo No: SAU/ENTOM/.....

Date:

.....

CERTIFICATE

This is to certify that the thesis entitled '**Identification of Insect Pests in Sweet Potato and their Management**' submitted to the Department of Entomology, Sher-e-Bangla Agricultural University, Dhaka, in partial fulfillment of the requirements for the degree of **MASTER OF SCIENCE in ENTOMOLOGY**, embodies the results of a piece of bonafide research work carried out by **Md. Anwar-Ul-Haque Limon**, Registration No. **10-03802** under my supervision and guidance. No part of the thesis has been submitted for any other degree or diploma.

I further certify that any help or source of information, received during the course of this investigation has been duly acknowledged.

Dated: June, 2017
Dhaka, Bangladesh

Professor Dr. Mohammed Ali
Department of Entomology
Sher-e-Bangla Agricultural University
Dhaka-1207
Supervisor

DEDICATED

TO

*MY BELOVED PARENTS and
GRANDPARENTS*

ACKNOWLEDGEMENTS

All praises and compliments are due to the Supreme Regulator and Ruler of the Universe, 'Almighty Allah' for the blessing upon the successful accomplishment of education, to complete the research work and thesis leading of Master of science (MS) in Entomology.

The author likes to express his heartfelt respect, gratitude and profound indebtedness to his reverend Supervisor Professor Dr. Mohammed Ali, Department of Entomology, Sher-e-Bangla Agricultural University (SAU), Dhaka, Bangladesh for his scholastic guidance, support, encouragement, valuable suggestions and constructive criticisms throughout the study period and gratuitous labor in conducting and successfully completing the research work and writing the manuscript.

The author also extends his appreciation profound regards and cordial thanks to his Co-supervisor Professor Dr. Md Abdul Latif, Department of Entomology, SAU, Dhaka, Bangladesh for his scholastic guidance, support, encouragement, valuable suggestions and constructive criticisms throughout the study period and gratuitous labor in conducting and successfully completing the research work and writing the manuscript.

Cordial thanks to Dr. Mst. Nur Mohal Akhter Banu, Associate Professor and Chairman, Department of Entomology, Sher-e-Bangla Agricultural University, Dhaka for her continuous cooperation during the research work. The author also expresses heartfelt thanks to all the teachers of the Department of Entomology, SAU, for their valuable suggestions, instructions, cordial help and encouragement during the study period.

The author expresses his sincere appreciation to her brother, sisters, relatives, well wishers and friends for their inspiration, help and encouragement throughout the study period.

The Author

IDENTIFICATION OF INSECT PESTS IN SWEET POTATO AND THEIR MANAGEMENT

BY

MD. ANWAR-UL-HAQUE LIMON

ABSTRACT

The present study regarding identification of insect pests in sweet potato and their management has been conducted during rabi season (November 2016 to March 2017) in the experimental field of Sher-e-Bangla Agricultural University, Dhaka. The variety of BARI Sweet Potato 8 was used as test crop. Six insecticidal treatments with one untreated control were tested in this study. They were T₁: Emamectin benzoate @ 2 ml/L of water; T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing; T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application); T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval (3 times application); T₅: Voliam flexi 300SC @ 1 ml/L at 15 days interval (3 times application); T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application) and T₇: Untreated control. The experiment was laid out in Randomized Completely Block Design (RCBD) with three replications. Under the present study 9 different species of insect pests were recorded and they were belonged to 9 family under 3 orders. Among the insect pests it was observed that sweet potato weevil was more destructive for storage roots of sweet potato. Data revealed that for sweet potato weevil, the lowest number of sweet potato weevil was T₂ (1.67), while the highest number from T₇ (6.73) treatment. At 15 days early harvest, in consideration of infestation level of storage roots in number and weight basis, the lowest infestation was recorded from T₂ (3.69% and 5.68%), whereas the highest infestation (12.25% and 12.98%) was observed from T₇ treatment. At optimum days harvest, for infestation level of storage roots in number and weight basis, the lowest infestation was recorded from T₂ (4.49% and 6.49%), whereas the highest infestation (13.56% and 14.16%) was observed from T₇ treatment. After 15 days from optimum harvest, in case of infestation level of storage roots in number and weight basis, the lowest infestation was recorded from T₂ (5.31% and 6.93%), whereas the highest infestation (16.37% and 17.05%) was observed from T₇ treatment. The highest yield was recorded from T₂ (41.86 t/ha), whereas the lowest from T₇ (30.27 t/ha) treatment. It was revealed that Carbofuran 5G @ 15 kg/ha at 60 days after sowing was the best for the controlling insect pests of sweet potato among different control measures under the study and also attaining highest yield of storage roots.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE NO.
	ACKNOWLEDGEMENTS	i
	ABSTRACT	ii
	TABLE OF CONTENTS	iii
	LIST OF TABLES	v
	LIST OF PLATES	vi
	LIST OF APPENDICES	vii
	SOME COMMONLY USED ABBREVIATIONS	viii
I	INTRODUCTION	01
II	REVIEW OF LITERATURE	04
	2.1 Description, Diversity, Biology and Distribution of <i>Cylas formicarius</i>	05
	2.2 Economic Importance of Sweet Potato Weevils	08
	2.3 Management of Sweet Potato Weevils	09
	2.4 Constraints to Sweet Potato Weevil Management	22
III	MATERIALS AND METHODS	23
	3.1 Location	23
	3.2 Climate	23
	3.3 Soil	23
	3.4 Land preparation	24
	3.5 Manure and Fertilizer	24
	3.6 Vine Collection	25
	3.7 Design of Experiment	27
	3.8 Planting	25
	3.9 Treatments of the Experiment	26

CHAPTER	TITLE	PAGE NO.
	3.10 Intercultural Operations	26
	3.11 Harvesting	28
	3.12 Data Collection	28
	3.13 Calculation	28
	3.14 Pest attacking in the field	29
	3.15 Beneficial insect in the field	33
IV	RESULTS AND DISCUSSION	34
	4.1 Incidence of different insect pest in sweet potato	34
	4.1.1 Species of insect pests	34
	4.1.2 Insect population	34
	4.2 Leaf and branches infestation by different insect pests in different stages	39
	4.2.1 At early growth stages	39
	4.2.2 At mid growth stages	41
	4.2.3 At late growth stages	43
	4.3 Storage root infestation by sweet potato weevil in different stages	46
	4.3.1 At 15 days early harvest	46
	4.3.2 At optimum days harvest	48
	4.3.3 After 15 days from optimum harvests	51
	4.4 Yield contributing characters and yield	53
V	SUMMARY AND CONCLUSION	55
	REFERENCES	59
	APPENDICES	71

LIST OF TABLES

TABLE	TITLE	PAGE NO.
1.	List of the insect pests found in the experimental sweet potato field during study period	35
2.	Number of identified insect pests in sweet potato during the growing period for different management practices	37
3.	Incidence of insect pests on leaf and branches of sweet potato at early growth stage for different management practices	40
4.	Incidence of insect pests on leaf and branches of sweet potato at mid growth stage for different management practices	42
5.	Incidence of insect pests on leaf and branches of sweet potato at late growth stage for different management practices	44
6.	Incidence of sweet potato weevil on storage roots of sweet potato at 15 days early harvest in number and weight basis for different management practices	47
7.	Incidence of sweet potato weevil on storage roots of sweet potato at optimum days harvest in number and weight basis for different management practices	49
8.	Incidence of sweet potato weevil on storage roots of sweet potato after 15 days from optimum harvest in number and weight basis for different management practices	52
9.	Yield and yield contributing characters of sweet potato for different management practices	54

LIST OF PLATES

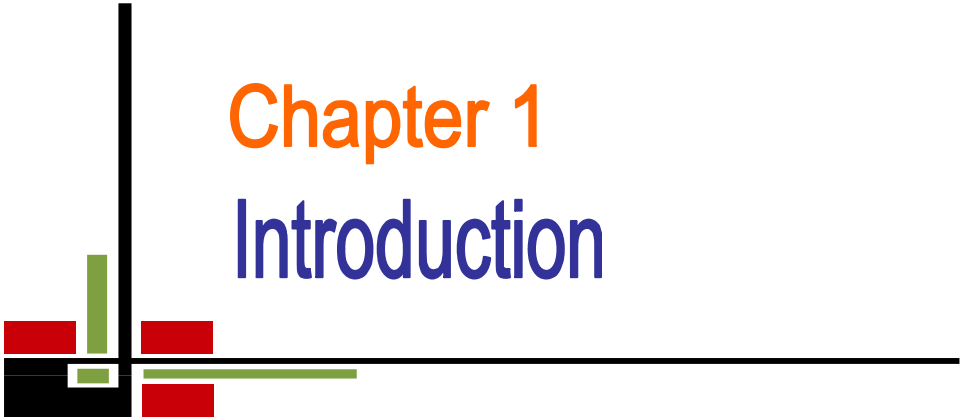
PLATES	TITLE	PAGE NO.
1.	Photograph showing Sweet potato bug	29
2.	Photograph showing Mealy bug	29
3.	Photograph showing green vegetable bug	30
4.	Photograph showing leaf hopper	30
5.	Photograph showing minor leaf feeders	30
6.	Photograph showing sweet potato beetle	31
7.	Photograph showing Tortoiseshell Beetle	31
8.	Photograph showing whitefly	32
9.	Photograph showing sweet potato weevil	32
10.	Photograph showing Ladybird beetle	33
11.	Photograph showing spider	33

LIST OF APPENDICES

APPENDIX	TITLE	PAGE NO.
I.	Monthly record of air temperature, relative humidity and rainfall of the experimental site during the period from November, 2016 to March 2017	71
II.	Analysis of variance of the data on number of identified insect pests in sweet potato during the growing period for different management practices	72
III.	Analysis of variance of the data on incidence of insect pests on leaf and branches of sweet potato at early growth stage for different management practices	72
IV.	Analysis of variance of the data on incidence of insect pests on leaf and branches of sweet potato at mid growth stage for different management practices	73
V.	Analysis of variance of the data on incidence of insect pests on leaf and branches of sweet potato at late growth stage for different management practices	73
VI.	Analysis of variance of the data on incidence of sweet potato weevil on storage roots of sweet potato at 15 days early harvest in number and weight basis for different management practices	74
VII.	Analysis of variance of the data on incidence of sweet potato weevil on storage roots of sweet potato at optimum days harvest in number and weight basis for different management practices	74
VIII.	Analysis of variance of the data on incidence of sweet potato weevil on storage roots of sweet potato after 15 days from optimum harvest in number and weight basis for different management practices	75
IX.	Analysis of variance of the data on yield and yield contributing characters of sweet potato for different management practices	75

SOME COMMONLY USED ABBREVIATIONS

FULL WORD	ABBREVIATION
Agro-Ecological Zone	AEZ
Bangladesh Bureau of Statistics	BBS
Co-efficient of variation	cv
Days After Sowing	DAS
and others	<i>et al.</i>
Etcetera	etc
Food and Agriculture Organization	FAO
Journal	J.
Least Significance Difference	LSD
Muriate of Potash	MoP
Sher-e-Bangla Agricultural University	SAU
Soil Resources Development Institute	SRDI
Triple Superphosphate	TSP



Chapter 1

Introduction

CHAPTER I

INTRODUCTION

Sweet potato [*Ipomea batatas*. (L.) Lam.] is very important crop in tropical and sub-tropical countries of the world including Africa, India, Bangladesh, China, Japan, the Pacific Island, Tropical America and the Southern part of the United States of America. Approximately 92% of the worlds sweet potato produced in Asia and the Pacific Island and 89% of this grown in China (Horton 1988b). Sweet potato is an introduced crop in Bangladesh during the second half of the 19th century (Rashid *et al.*, 1982). The crop belonging to the family Convolvulaceae in an important root crop in Bangladesh and commonly known as “Misti Alu” (Rabbi, 1995). Sweet potato is a major root crop which is grown for both export and local consumption. It is also the third most important root crop grown in eastern Africa after cassava and potato (FAO 2011).

Sweet potato is both a staple and a food security crop in eastern and southern Africa, and is mainly grown by smallholder women farmers (Mutuura *et al.*, 1992; Bashaasha *et al.*, 1995; Andrade *et al.*, 2009). Sweet potato is also grown for its vines as planting material; leaves are often eaten as a vegetable while shoots and roots are used as animal feed in many countries. In Uganda and western Kenya, the sale of fresh sweet potato roots, vines and processed foods in both local and urban markets is becoming increasingly popular regarding contributing to household cash income (Abidin 2004; Kaguongo *et al.*, 2012). Orange-fleshed sweet potato is also a rich source of beta-carotene, a precursor of bio-available vitamin A, and has potential of combating Vitamin A deficiency among rural resource constrained farmers in many developing countries (Jalal *et al.*, 1998; Jaarsveld *et al.*, 2005; Low *et al.*, 2007; Mwangi *et al.*, 2003a; Burri, 2011).

The sweet potato is a multipurpose crop. In developed countries of the world, the sweet potato is primarily used for food, prepared mostly from its flesh but large quantities are canned and some are dehydrated. In Bangladesh, it is consumed in boiled condition. Sweet potato flour is good alternative to wheat flour. Cakes, biscuits, breads and many kinds of foods can be prepared with flour of sweet potato. In industries, it is used in manufacturing alcohol, syrup and starch. Sweet potato is a high calorie producing crop. Yield of sweet potato per hectare is

higher than any other cereals. Storage roots of sweet potato are rich in starch. Leaves and stem tips of its vein are frequently used as green vegetables, salad and cattle feed (AVRDC, 1976).

Sweet potato is one of the important root crops in Bangladesh as well as in the world. Sweet potato ranks as the world's seventh most important food crop after wheat, rice, maize, potato, barley, and cassava. According to the Food and Agriculture Organization (FAO, 2012), sweet potato was under cultivation in 82,40,969 hectares of land in the world. Considering top 20 sweet potato producing countries in 2012, world's total production was 101,839,463 tons and a majority of which came from China, with a production of 77,375,000 tons (FAO, 2012). Bangladesh produces different varieties of sweet potato but some of the varieties are produced in more quantity based on consumer demand and easy cultivation technique as well as vine (planting materials) availability. It is cultivated more or less in all the districts of the country. However, the suitable areas of sweet potato cultivation are "char land" located on the both sides of rivers.

In Bangladesh sweet potato is the 4th most important source of carbohydrate after rice, wheat and potato. Sweet potato plays a significant role in increasing food security and income for the poor farmers of Bangladesh (Ahmed *et al.* 2015). The area and production under sweet potato was 24,567 hectare and 297,539 tons, respectively in Bangladesh during the year 2011 (FAOSTAT, 2011). The average per hectare production of sweet potato in Bangladesh is 9.8 tons (FAOSTAT, 2011). Most of the sweet potato producers in Bangladesh are smallholder. Smallholder farmers struggle because of their limited access to inputs (e.g. credit, technology, information) while working on low-productivity land located far distances from output markets via an inadequate, high-cost road system (Lunna and Wilson, 2015). The production of sweet potato has decreased from 435,000 MT in 1995-96 to 253,000 MT in 2011-12 (BBS, 2012).

Sweet potato is grown over a wide range of environmental and edaphic conditions. It requires low inputs and less management practices (AVRDC, 1977). A raw sweet potato contains water (77%), carbohydrate (20.1%), protein (1.6%), fiber (3%), and almost no fat. Many studies have suggested that increasing consumption of sweet potatoes decreases the risk of obesity, heart disease, diabetes and overall mortality while promoting a healthy complexion, increased energy and overall lower weight. Yield of local sweet potato in Bangladesh is 10 t/ha. But high yielding variety may produce 35-45 t/ha. At least 18 species of insect feed on sweet potato roots. Among those causing the greatest damage are the sweet potato weevil (*Cylas formicarius* Fab.), white fringed beetle, wireworm larvae and flea beetles. Sweet potato weevil is the most serious pest of sweet potato in Bangladesh. It's infestation may be found in the field, in storage sheds and in propagation beds. Losses due to insect feeding, especially sweet potato weevils may often reach 60-100% because most sweet potatoes are produced in low input agricultural system (Chalfant *et al.*, 1990). So effective management practices should be done to minimize the production loss by the insect pests and to minimize the pesticide hazards by using bio-rational insecticides.

Objectives:

1. To identify insect pests attacking in sweet potato.
2. To assess the damage severity of major insect pest.
3. To develop a suitable management packages for controlling the insect pest.



Chapter 2

Review of literature

CHAPTER II

REVIEW OF LITERATURE

Infestation of the sweet potato crop by different insect species started immediately after the crop establishment and continued for five months in each year. More than 50 insects belonging to several orders and at different stages of development infested the crop. About eight insect species caused major damage on the crop leaves, vines and tubers. The most destructive and important economic pest species were sweet potato weevil (*Cylas puncticollis* Boh.) and the clearwing moth (*Synanthedon dasycyales*). Some 21 insect species were of minor importance, as their damaging effects were not noticeable on the plant and they had low effects on the yield. Other than pests, seven insect species were found to be beneficial as predators or parasitoids of the insect pests, implying that any management practice employed for control of the major pests should consider conservation of the natural enemies.

Most of the major and minor insect pests reported by this study have also been noted by earlier studies such as Kibata (1973) with the exception of *Systates* spp.

In Zimbabwe, thirteen pests are reported to infest sweet potato tubers, stems, crowns and leaves. However, among these, only *Cylas formicarius elegantulus* and termites were not observed in this present study. Most of the insect pests identified on sweet potato in this study have been reported by Ames *et al.*, (1996).

In his book, Hill (1975), indicates that *Bermisia tabaci*, *Synanthedon dasysceles*, *Agrius convolvuli*, *Acraea acerata*, *Cylas puncticollis*, *Alcidodes dentipes* and *Aspidormorpha* spp. are the major pest of sweet potato in Africa.

Bohlen, (1973), reported that although many insect species were recorded, only a few were important pests of sweet potato. The key coleopteran pests included *Cylas puncticollis*, *Blosyrus obliquatus*, *Systates polinosus*, and Cassid beetles.

Coleopterans have also been recorded as major pests of sweet potatoes in Tanzania.

According to Panizzi and Slansky, (1985), *Prezotrachelus gestackerii* (Coleoptera: Curculionidae) was the only observed pest that had not been recorded in many places. It was the sixth most important pest observed during the second season. It's however a very minor pest and its effects do not cause any economical loss to the sweet potato. However, it has been recorded as a major pest in legumes.

Most of the minor pests observed are cosmopolitan, polyphagous and are pests of other crops. These include *Nezara viridula*, *Myzus persicae*, and *Acanthomia tomentosicollis* (Hill, 1975) and they infest many crops grown in the study site. Some insects present at the time of sampling were transients (tourists), which had no direct effect on the crop. These included banana weevil (*Cosmopolites sordidus* (Germ.)), Mango weevil (*Sternochetus mangiferae* (F)) and *Dysdercus nigrofasciatus*, which is a major pest of okra and cotton. The study revealed that *Synathedon dascyceles* was a serious pest of sweet potato in the region, and there is need to study the biology and management of this moth. Though the study did not assess yield loss, continued infestation of the crop by these insects evidently showed that there were losses which could be incurred overtime.

2.1 Description, Diversity, Biology and Distribution of *Cylas formicarius*

2.1.1 Description of *cylas* spp.

Sweet potato weevils is in the genus *Cylas* (Coleoptera: Apionidae) (Chalfant *et al.*, 1990; Smit, 1997a). Contains three species namely *Cylas brunneus* (Olivier), *Cylas puncticollis* (Boheman) and *Cylas formicarius* (Fabricius) (Wolfe, 1991). Adult weevils are elongated smooth and shine with ant like snouted beak but species can be differentiated by size and colour (Smit, 1997a). *C. formicarius* are small with a bluish black abdomen and a red thorax, *C. puncticollis* are black and large *C. brunneus* are small either black or brown (Wolfe, 1991).

2.1.2 Diversity of sweet potato weevils

Parker *et al.* (1990), stated that both *C. puncticollis* and *C. brunneus* are the most common species in East Africa. On the other hand, *Cylas formicarius* is the main pest species in Asia, the United States and Oceania, but in Africa it has been found in Natal, South Africa and on the coast in Kenya.

2.1.3 Biology of sweet potato weevils

Ames *et al.* (1996), found that all three species have a similar life history. The adult female lays eggs singly in cavities excavated in vine or in roots, preferring the latter. The egg cavity is sealed with a protective, gray fecal plug. The developing larvae tunnel in the vine or root. Pupation takes place within the larvae tunnels. A few days after eclosion, the adult emerge from the vine or roots because the female weevil cannot dig, she finds soil cracks. Alternate host of sweet potato weevils are *Ipomoea* spp. weeds.

2.1.3.1 Eggs

From Otto *et al.* (2006), Eggs are deposited in small cavities created by the female with her mouthparts in the sweet potato root or stem. The female deposits a single egg at a time, and seals the egg within the oviposition cavity with a plug of fecal material, making it difficult to observe the egg. Eggs will hatch after three to seven days depending on the environmental conditions.

According to Schimutterer (1969), the egg is oval, and yellowish-white.

2.1.3.2 Larvae

According to Allard (1990), The larvae have head, thorax and abdomen. The head is pale brown with darker brown mandibles, one pair of ocelli (stemmata), each containing two contiguous pigment spots.

From Otto *et al.* (2006), The thorax is divided into prothorax and mesothorax whereby mesothoracic spiracle located on a lobe very close to the prothorax. The abdomen is whitish, legless, slightly curved, approximately 10 millimetres length maximum width two millimetres cuticle speculate. Larvae have three

instars, first larval instar, second larval instar, and third larval instar. Total larvae period varies from 10 – 25 days.

2.1.3.3 Pupae

Otto *et al.* (2006), found that the pupa of sweet potato weevil is white and approximately six millimetres in length; pronotal width one metre, cuticle glabrous. The head and rostrum are provided with setiferous tubercles, one pair between the eyes at base and two pairs on the rostrum. The posterior pair being close to the eyes and the anterior behind the middle. Pupation occurs inside the vine or root and takes eight days for adults to emerge.

2.1.3.4 Adults

The pest is black, with a faint, metallic blue luster, and not with a distinctly shiny, copper-like sheen, body length is eight millimetres. The length of male antennal club is equal to or greater than combined length of all preceding segments. Eyes close together in dorsal view; distance between eyes is about one sixth of minimum width of rostrum.

According to Ames *et al.* (1996), At an optimal temperature of 27-30⁰ C, *C. formicarius* completes development (from egg to adult) in about 33 days. Adult longevity is two to three months and females lay between 100 and 250 eggs in this period.

According to Okonyo (2013), The male and female adult sweet potato weevils can be told apart by the shape of their antennae. The antennae of the males are straight while those of the female are round or club-shaped. Under favourable conditions, sweet potato weevils can produce 13 generations a year and can live for three to four months.

2.1.4 Distribution

Parker *et al.* (1990), found that the sweet potato weevil is one of the major pests of sweet potato worldwide. Three species have been identified in Africa. Their distribution in Africa is being surveyed and it appears that all the three species

have a similar life history, making all of them difficult targets for conventional pest control measures.

According to Nderitu *et al.* (2009), among the three, *C. formicarius* is an important pest in India, South East Asia, Oceania, the United States and the Caribbean.

2.2 Economic Importance of Sweet Potato Weevils

Sweet potato weevil is one of the most important biotic factors limiting sweet potato production in Africa (Chalfant *et al.*, 1990; Pfeiffer, 1982; Smit and Matengo, 1995). Sweet potato is the sixth most important food crop in the world (Vietmeyer, 1986) and both adults and larvae cause serious damage to leaves and stems. Adults attack the leaves of sweet potatoes, but the larvae are more injurious boring into the stems and causing serious mortality to seedlings (Daiber, 1994).

Wolfe (1990), described that adults of all three *Cylas* species feed on the epidermis of vines and leaves, scraping oval patches off petioles, young vines and leaves. Yield loss is seldom serious. Adults also feed on the external surfaces of roots causing round feeding punctures which can be distinguished from oviposition sites by their greater depth and the absence of a faecal plug. The developing larvae tunnel in the vines and roots causing significant damage. Frass is deposited in the tunnels. In response to the damage, the root produces terpenes which render the infested root part inedible (Sato *et al.*, 1981).

Allard *et al.* (1991), reported on serious larval infestations disrupting sweet potato nurseries in Ethiopia, on established plants, the larvae feed in the tubers and stems producing larvae tunnels and later, pupal chambers stem damage is believed to be the main reason for yield loss, although damage to the vascular system caused by feeding, larvae tunnel secondary rots reduce the size and number of roots. In Eastern Africa both *C. puncticollis* and *C. brunneus* can be found infesting the same root. When a root is exclusively infested by *C. puncticollis*, the core of the root might still be untouched; *C. brunneus* larvae seem to tunnel further inside the root. First severe infestations render the crop

unpalatable and therefore inedible to humans. Pest damage usually continues during storage, therefore infested tubers cannot be stored for a long time. In conjunction with other coleoptera pests, sweet potato weevil can completely destroy sweet potato plantations (Geisthardt and van Harten, 1992). Weevil feeding on storage roots induces terpenoid production that makes even slightly damaged root unpalatable (Uritani *et al.*, 1975; Sato *et al.*, 1981). Thus low weevil densities may cause devastating crop losses of up to 60 -100% (Chalfant *et al.*, 1990).

2.3 Management of Sweet Potato Weevils

2.3.1 Cultural control

Stathers *et al.* (2005), reported different successful cultural practices used in experiments conducted in East Africa, Taiwan, Philippines, Vietnam, America, India, Cuba and Indonesia. Cultural practices aimed at preventing infestation proved to be effective way of reducing weevil damage.

a) Field sanitation

Smit and Matengo (1995), studied on the farmer's cultural practices in Kenya and suggested that crop protection workers should concentrate their research and extension efforts on crop sanitation and the avoidance of adjacent planting of successive crops.

According to Stathers *et al.* (2005) removal and destruction (through burning or feeding to livestock) of infested vine and root remains. If vines are left in the field to maintain soil fertility, care should be taken to ensure they are dead or dry and not able to sprout and then provide food for weevils. If peace meal harvesting of the crop is practiced, care should be taken to remove and destroy any infested roots that are found.

Komi (2000), reported that removal of alternate host plants like *Calystegia soldanella*, *C. hederacea* and *Ipomoea indica* reduced the sweet potato weevil infestation in Japan. Removal of volunteer sweet potato plant and wild morning

glories as these may be alternative hosts (Sato *et al.*, 1981).

Geisthardt and van Harten (1992), stated that crop rotation with other crops for two to three seasons appears to be the most effective method of preventing infestations of weevils.

In a large ecosystem area or community burying can help to reduce sweet potato weevil infestation. Infested storage roots must be buried >15 cm underground (Stathers *et al.*, 2005).

b) Hilling up

Allard *et al.* (1991), reported that collection of soil around the base of plants to prevent or fill soil cracks. This practice not only protects the plants from weevil attack but can also result in increased crop yield.

According to Palaniswami and Mohandas (1994), Re-ridging the crop at tuber formation stage prevents the weevil from laying the eggs and the entry of the grubs into the tuber. The efficacy of re-ridging in sweet potato crop, as a cultural practice for reducing weevil incidence was investigated over two seasons at Vellayani, Kerala, India. Five re-ridgings between 50 and 90 days after planting, at 10 days interval, significantly reduce the weevil damage to the tubers.

Deep planting into the hill, and planting into the furrow and earthing after six weeks offers protection against weevils (Macfarlane, 1987).

c) Mulching

Geisthardt and van Harten (1992), described that application of dry leaves on the soil to keep it moist, prevent cracks and provide a more favourable place for natural enemies. Care should be taken to make sure the weevils cannot feed or develop on the mulching material.

According to AVRDC (1987), mulching with rice straw or black plastic reduced the infestation by SPW in the root zone. The availability of mulch is a problem especially in dry-lands coupled with higher temperatures. Termites are a problem during summer as they completely devour the dried grasses and other shrubs in dry lands.

d) Early harvesting

Ebregt *et al.* (2005), stated that harvesting two weeks earlier reduce the loss due to weevil from greater than 30 percent to less than five percent.

From Powell *et al.* (2001), early harvesting of the crop is practiced to ensure that infested roots are removed and destroyed. Vines left in the field should not be allowed to sprout and then provide food for weevils.

According to Stathers *et al.* (2005), timely harvesting to remove the largest storage root most at risk from weevil attack and subsequent hilling up of the soil around the remaining root to prevent weevils from accessing the root through cracks in the soil.

e) Flooding

Otto *et al.* (2006), found that flooding of the field for more than 48 hours kill the weevil larvae present in roots that have been left in the field. Flooding of fields between two consecutive sweet potato crops may reduce the immediate source of weevil from the field. Sweet potato weevils can be controlled by flooding of fields before planting (Stathers *et al.*, 2003)

Talekar (1987), reported that, flooding of infested field for at least 48 hours after completing harvest drown weevils and induces rotting of the left over plant materials and thereby reduces weevils' densities from one planting to the next. This is an option in areas where rotation is not possible

f) Intercropping

According to Stathers *et al.* (2005), In Taiwan, 103 different crops were tested as intercrops for sweet potato weevil control, the best result were obtained with

coriander (*Coriandrum sativum* L.).

According to Rajasekhara Rao (2005) and Rajasekhara Rao *et al.* (2006), Mixed cropping systems with sweet potato and other crops (ginger, okra, maize, colocasia and yam) are practiced by farmers in North Eastern Region of India. Low incidence of *C. formicarius* was noticed in these systems and the interaction of intercrops and several insect pests of tuber crops, including sweet potato weevil, in these multiple cropping systems.

g) Use of clean planting material

Allard *et al.* (1991), described the following techniques that have been used in the management of sweet potato weevils. Planting only in fields that have had no weevil infestations within the last 12 months and preferably more than 1 km away from any infested land; planting resistant or tolerant cultivars; selecting deep-rooting cultivars, with long necks between the roots and the stems (which are less susceptible because the adult weevil cannot burrow downwards more than one centimeter); planting early-maturing cultivars which can escape serious damage; earthing up of plants (hilling), particularly those cultivars with the tendency to push out of the ground; removal of all plant debris and volunteer plants after harvest; re-ridging approximately 30 days after planting as this places the roots deeper and out of reach of the weevils; planting non-infested material; and use of intercropping.

Weevils tend to lay eggs in the older woodier parts of the sweet potato vine, so if the tender tips are used for planting they are less likely to be infested by weevils (Nair, 2006).

2.3.2 Biological control

2.3.2.1 Parasitoids

Maeto and Uesato (2007), reported a new species of braconid, *Bracon yasudai* from the south-west islands of Japan. It is a solitary idiobiont ectoparasitoid of the larvae of the West Indian sweet potato weevil, *Euscepes postfasciatus*, and the sweet potato weevil, *C. formicarius*, both feeding on *Ipomoea batatas* (L.).

Palaniswami and Rajamma (1986), reported the braconids *Rhaconotus* spp. and Bracon spp. and an unidentified hymenopterous parasitoid on the larvae of sweet potato weevil.

Jansson and Lecrone (1991), reported *Euderus purpureas*, an eulophid parasitizing on *C. formicarius* in Southern Florida. Nevertheless, the success of all these parasitoids at field conditions is doubtful since they are recorded in a very few numbers.

2.3.2.2 Predators

Lagnaoui *et al.* (2000), stated that the use of ants against weevils is another component of the control strategy adopted for the sweet potato weevil in Cuba. Two species of predatory ants, *Pheidole megacephala* and *Tetramorium guineense* (*Pheidole guineensis*), are common inhabitants of banana plantations. Rolled banana leaves were used as “temporary nests” to transport the ants from their natural reservoir to sweet potato fields, where they prey upon weevils and other insects. Setting up ant colonies in the field 30 days after planting with 60-110 nests has reduced weevil infestation from three to five percent.

Allard *et al.* (1991), described that the subterranean habitat of *C. puncticollis*, whilst making it less accessible to predators and parasitoids may enhance the impact of fungal pathogens which require a protected cool, humid environment for survival and reproduction; conditions generally found under dense foliage of sweet potato. The eggs are also well protected as they are laid within vines, or in tubers and egg cavity are sealed with a fecal plug that preserves moisture, disguises location and protects the eggs from predatory mites. Potential candidates for use as biological insecticides include *B. bassiana* and *M. anisopliae*. Isolates of the former have been collected from laboratory-reared adults originally collected in Kenya.

2.3.2.3 Entomopathogenic nematodes

Entomopathogenic nematodes (EPN) have beneficial interaction with

sweetpotato within the roots and are promising for the control of sweet potato weevils (Jansson and Lecrone, 1997).

Among different species, Heterorhabditis was found to be most effective, infective and pathogenic than Steinernematids (Mannion and Jansson, 1992).

According to Jansson *et al.* (1991), Heterorhabditid nematodes were more pathogenic to pupae, than were Steinernematid nematodes. Weevil adults were the least susceptible to nematode infection. The number of applications of bacteriophora did not significantly reduce numbers of *C. formicarius* but consistently reduced damage to sweet potato tubers. Also bacteriophora, is more infective than Steinernematids carpocapsae ('All' strain). Subsequent field tests showed that one application of bacteriophora has found effective at protecting sweet potato tubers from weevil damage. This nematode persisted for over 130 and 250 days after application in two separate experiments, respectively. Nematode application rate had no effect on densities of *C. formicarius* or damage caused suggesting that a single application early in the growing season is adequate.

Weevil damage to plants treated with insecticides is intermediate to that on nematode-treated and untreated plants weevils (Jansson and Lecrone, 1997).

2.3.2.4 Entomopathogenic fungi

According to Su (1991a), most effective entomopathogenic fungi infecting sweet potato weevil has been identified as *Beauveria bassiana* (Bals.) Vuill, which can be applied as a foliar spray or in combination with pheromone trap, for its successful infection and dispersal. Spraying of *B. bassiana* solution (isolated from *C. formicarius*) at a concentration of 1.6×10^4 conidia ml⁻¹ at planting and rootstock formation, and broadcasting soybeans containing *B. bassiana* into the rows at planting controlled *C. formicarius* effectively.

Su (1991b), indicated that application of *B. bassiana* isolated from honey bees at a concentration of 1×10^6 conidia ml⁻¹ at planting and rootstock formation also gave the best results. The success of infection of *B. bassiana* on sweet potato weevil is dependent on the type of soil.

Yasuda *et al.* (1992), experimented on thirteen different soils were pernicious to sweet potato weevil out of eighty soils collected from central and southern

Taiwan because when they were added to the row at planting, mortality caused to the sweet potato weevil by *B. bassiana* was >80%. Environmental factors also govern the successful infection of sweet potato weevil by entomopathogenic fungi. Low relative humidity (<43%) and low temperatures below (<15°C) were not conducive for *B. bassiana* infection of adults of sweet potato weevil.

Yasuda (1995), developed an auto-infection system consisting of a modified sex pheromone trap and a bottle with exit holes containing conidia of *B. bassiana* ($9.3 \times 10^{10} \text{ g}^{-1}$ medium) successfully tested to control *C. formicarius* in Japan. Male weevils were attracted to the system by the action of pheromone and exited the bottle were found infected with *B. bassiana*.

From Yasuda *et al.* (2004), Formulations of *B. bassiana* conidia in a 10% corn oil mixture showed more superior infectivity in both sexes of *C. formicarius* than the formulation of conidia.

Low cost and effective technology of production of *B. bassiana* at a cottage industry level to control sweet potato weevil was successfully established and adopted by many farmers in Cuba (Lagnaoui *et al.*, 2000).

Use of the fungus is particularly attractive because it relieves farmers from the high cost of chemical pesticides. Incubation of entomopathogenic fungi in different growth media results in production of several metabolites with insecticidal activity. Squamulosone (aromadendr-1(10)-en-9-one) was isolated in large quantity from the plant *Hyptis verticillata* and incubated with the fungus *Curvularia lunata* in two different growth media (potato dextrose broth and beef extract medium) (Collins *et al.*, 2001).

C. lunata is well known for its efficient 11 α -hydroxylation of steroids (Holland and Reimland, 1985; Chen and Wey, 1990) and also has been used to transform a number of terpenes (Azerad, 2000). *B. bassiana*, the fungus which is responsible for the muscardine disease in insects, was also known to affect the bioconversion of many substrates including alkaloids, steroids and terpenes. This strain, formerly known as *B. sulfurescens* or *Sporotrichum sulfurescens* has been

used to effect the reduction of various carbonyl compounds (Davies *et al.*, 1989). *B. bassiana* is also known to selectively hydroxylate non-activated carbon atoms (Lamare *et al.*, 1991; Holland, 1992). Incubation of cadina-4, 10 (15)-dien-3-one with *B. bassiana* results in the production of nine novel sesquiterpenes which are effective against *C. formicarius elegantulus* (Buchanan *et al.*, 2000). Labo-Lima (1990) conducted bioassays to evaluate the pathogenicity of the fungal pathogens *M. anisopliae* and *Beauveria bassiana* against *C. puncticollis*. Mortality rates obtained were encouraging for further research on the control of *C. puncticollis* with these fungi.

2.3.3 Chemical Control

Allard *et al.* (1991), stated that sweet potato weevil is a difficult target for conventional pest control measures as the larvae feed in the storage roots in the ground, or inside the woody base of the stems. This means that with the possible exception of systemic insecticides, which are costly and pose the risk of residual contamination of the tubers, there is no effective chemical control of the larvae, nor of the other stages found within the plant tissue.

In Ethiopia, insecticidal screening trials tested the use of foliar sprays applied three months after planting, followed by four applications at fortnightly intervals, and also root dipping prior to planting. Deltamethrin and pirimiphos methyl gave good control of sweet potato pests (Allard, 1990).

Recommendations for the use of 19 insecticides for the control of sweet potato weevils are provided by Kasasian (1978).

Rajamma and Pillai (1991) studied on several insecticides were tested for the management of sweet potato weevil by using them after planting, either by foliar spray or basal granular applications and found showed that 0.05% fenthion or endosulfan spray at monthly intervals with or without running or soil application of carbofuran or phorate granules each 1 kg a.i. ha⁻¹ 45 days after planting days

after planting are all effective in reducing infestation by *C. formicarius* resulting in greater marketable tuber yields.

Some of the insecticides are also used for vine dipping for successful control of sweet potato weevil. Fenvalerate, permethrin and deltamethrin each 0.003% are the most effective insecticides to *C. formicarius* (Rajamma, 1990).

Teli and Salunkhe (1994), tried dipping cuttings in insecticide solution before planting and spraying the crop one month after planting, and further three times, at three week intervals, subsequently with cypermethrin or fenvalerate each 375 g a.i. ha⁻¹, was most effective in reducing damage caused by the insect. Similar works on the use of several other insecticides to achieve better control of sweet potato weevil are available (Mason *et al.*, 1991; Sinha, 1994).

Misra *et al.* (2001), reported that combination of vine dipping of 0.05% monocrotophos and three foliar sprays with 0.05% endosulfan proved very effective than that of basal application of phorate granules followed by vine dipping of 0.05% monocrotophos. Soil drenching at 50 and 80; 60 and 90; and 50, 65 and 80 days after planting were equally effective in suppressing the incidence and intensity of weevil damage (Palaniswami and Mohandas, 1996). Among the single soil drenching, application on the 65th day was assessed to be the most effective against the weevil.

Palaniswami *et al.* (2002), reported that endosulfan, fenthion and fenitrothion each at 0.05% applied as soil drench at 50 and 80 days after planting were effective against *formicarius* and their residues in tubers at harvest were lower than the detectable levels. Chlorpyrifos and fensulfothion granules are more toxic to sweet potato weevil adults than the other insecticides, while their persistence was about 10 months (Hwang and Hung, 1991).

2.3.4 Host Plant resistance

Rajasekhara Rao (2002, 2005), stated that plant resistance provides a pivotal role in the management of insect pests. The physical attributes of the tuber namely,

the shape, length, neck length, colour of the skin, flesh colour and thickness plays an important role in preference by *C. formicarius* apart from the inherent nutritional quality of the sweet potato plant and tuber. Round tubers are preferred by more than elongate and spindle-shaped ones.

Teli and Salunkhe (1996), reported that round and oval tubers of sweet potato were more infested in the field by *C. formicarius* than long stalked, spindle and elongate ones. Pink and red coloured tubers are considered less susceptible than white and brown coloured ones. Cultivars with thin foliage and lobed leaves with purple coloration at emergence were found less susceptible.

Stathers *et al.* (2003), reported that plant resistance in sweet potato to environmental stress is underway in many parts of the world to fulfil the requirements of farmers situated in arid and semi-arid tropics. Only a limited success has been achieved, because of the inconsistent expression of the resistance (Rajasekhara Rao, 2002). Drought stress may increase the activity of oviposition stimulant present in the genotypes because weevils deposited more eggs on drought-stressed plants (Mao *et al.*, 2004).

Harrison *et al.* (2003), described that some of the plant metabolites are produced and influenced by environment, which would have a bearing on resistance or tolerance. The analyses showed that the levels of resin glycosides and caffeic acid vary between sweet potato genotypes and within genotypes among years or areas of production and have shown insecticidal activities (Jackson and Peterson, 2000). This may indicate a relationship between the quantity of these two compounds and the antibiosis of sweet potato.

According to Singh *et al.* (1993), host plant nutritional parameters are also believed to affect incidence of sweet potato weevil. The relationship between potash and silica in sweet potato stems is negatively correlated with *C. formicarius* infestation. Nitrogen and potassium influence the storage root-surface chemistry of sweet potato genotypes which have a bearing on resistance to sweet potato weevil (Marti *et al.*, 1993).

Rajasekhara Rao (2002), reported that selection of sweet potato genotypes with decreased volatile attractants (kairomones) and/or increased deterrents may significantly facilitate developing resistance to sweet potato weevil. Plant resistance can also be induced through alterations in nutritional regimes of the crop. A triterpenol acetate was identified in the root surface on a sweet potato weevil susceptible genotype 'Centennial', but not in more resistant genotype (Nottingham *et al.*, 1989).

Wilson *et al.* (1988), demonstrated that a methylene chloride surface extract of the periderm of tubers of the susceptible 'Centennial' stimulated oviposition of *C. formicarius elegantulus*. Nottingham *et al.*, (1987) also showed that ovipositional stimulant resided in the tuber periderm, not in the core of the tuber.

Allard *et al.* (1991), researched on breeding and evaluating sweet potato germplasm for resistance. The development of insect- resistance is seen as a viable component of integrated pest management programmes. Mechanisms of resistance to sweet potato weevil in sweet potato include antibiosis, antixenosis (non – preference) and escape (for example, long and thin storage roots set deep in the soil and scattered within growing hills). Resistance characters identified as under polygenic inheritance include fleshy root density, dry matter and starch content, root depth, vine thickness and tuber chemistry.

Anota and Odebiyi (1984), found no evidence that nitrogen, starch, dry matter, or moisture content played a role in tuber resistance of five resistant sweet potato cultivars tested in Nigeria, but carotene content was identified as a major factor. No oviposition preference for tubers or vines was apparent. There was a lower survival rate in all life stages, smaller body weights and a longer developmental period of *C. puncticollis* raised on resistant cultivars. Two orange fleshed and 18 white fleshed cultivars evaluated in the field for resistance to *C. puncticollis*. Cultivars (TIS 3053 and TIS 3030) exhibited the least root damage, whereas Cultivars (TIS 2532, TIS 3017 and TIS 3030) showed the least shoot damage (all white – fleshed) (Hahn and Leuschner, 1981). Cheng (1981) identified 14 sweet potato lines resistance to *C. puncticollis*, but their yields were lower than those

of the majority of susceptible lines (Cheng, 1981). Screening of 700 cultivars of sweet potato in Nigeria revealed low resistance levels to *C. puncticollis* (Hahn and Leuschner, 1981).

Shakoor *et al.* (1984), identified seven resistant sweet potato lines, including KSP 20, KSP1K, SP 19 and KSP 20 following screening of 93 accessions of diverse origin. In trials conducted in the Philippines by Petro *et al.* (1986), local cultivars Karingkit and Kadulaw gave the highest yields, whereas the improved cultivars tested exhibited undesirable agronomic characteristic and susceptibility to *C. puncticollis*.

2.3.5 Pheromonal control

According to Downham *et al.* (2001), the behaviour in both the laboratory and the field indicate that there is probably a sex pheromone for *C. puncticollis*, as is the case for *C. formicarius*. In rearing rooms, gregarious clumping behaviour has been noted, as well as the obvious attraction of males towards the females. In the field, males are predominantly found under the foliage, presumably seeking females. Simple laboratory tests have shown that males are attracted to females. The feasibility of using virgin, unmated females as baits for males in simple traps was investigated in Kenya (Allard, 1990).

Heath *et al.* (1986), stated that precise decision tool to assess the time of sweet potato weevil incidence on sweet potato, suggest the use of sex pheromones (boehmeryl acetate). Soon after the identification of female sex pheromone of *C. formicarius*, the course of weevil management in different parts of sweet potato growing countries around the world changed dramatically. The success of sweet potato weevil control in these countries is unequivocally assigned to the sex pheromone. The sex pheromone, alone contributed to significant reductions in weevil populations and tuber damage, which resulted in greater marketable storage root yield.

According to Yasuda *et al.* (1992), the sex pheromone (Z)-3-dodecen-1-ol (E)-2-

butenoate, proved to be a successful mating disruptant for the control of *C. formicarius* (Mason and Jansson, 1991; Yasuda, 1995; Yasuda *et al.*, 1992). It is also used to trap both sexes of sweet potato weevil. With 100 mg of sex pheromone, the attraction was 60.88%. Entrapment of weevils differs significantly during different parts of the day.

Yasuda *et al.* (1992), reported that pheromone traps installed close to the ground trapped more males than those set above the ground. Most of the males approached the traps by walking. Different types of traps used for monitoring and mass trapping. When the sex pheromone discovered and synthesized, various designs and types of traps used for the weevil management viz. Light, sticky, water, plastic funnel live trap etc. Talekar and Lee (1989) reported that female sex pheromone identified in *C. formicarius elegantulus* was also active in *C. formicarius*.

According to Li (1998), sweet potato weevil sex pheromone traps each 30 hectares with a distance of 15 m between traps reduced the tuber damage by 10.1% and controlled the sweet potato weevil up to 53.1-58.2% in China.

Pillai *et al.* (1996), reported that trap containing synthetic sex pheromone (100 traps per hectare) is highly effective for mass trapping the male sweet potato weevil which significantly reduced population build-up, consequently resulted in greater marketable tuber yield. Pheromone traps each 40 per hectare reduced sweet potato tuber damage by 65% by sweet potato weevil and use of chlorpyrifos each two kilogrammes active ingredient per hectare in addition to the pheromone reduced the damage by an extra 10% over the pheromone treatment (Hwang and Hung, 1991).

Yasuda *et al.* (2004), described that integration of sex pheromone traps with insecticides or entomopathogenic fungi are practiced successfully. Development of a new pheromone formulation to increase exposure time to insecticide for control of the sweet potato weevil. The attracted males located and tried to mate with the ball. They were thereby efficiently exposed to the insecticide for a longer time. The concentration of the new formulation was extremely low

compared to the conventional formulations, and therefore, lowers the cost of application. The formulation was a combination of sex pheromone, butenoate, and an insecticide impregnated into blue ball made of the diatomaceous soil. The male weevils were attracted to the visual stimulation in addition to the sex pheromone (Smit *et al.*, 2001).

2.4 Constraints to Sweet Potato Weevil Management

Stathers *et al.* (2005), indicated that pesticides kill natural enemies that under natural circumstances quite effectively control weevil populations, and can also present health, risks for human and animals. In some countries planting materials are dipped into on synthetic pesticide before planting, which can delay pest infestation for several months, however, most pesticides are expensive and highly toxic therefore dipping is only likely to be economical for large scale commercial root production or vine multiplication nurseries. Other varieties escape weevil damage because their storage roots mature quickly and can be harvested early. Breeders have spent many years trying to develop varieties that are resistant to the weevil. So far they have not been successful (Stevenson *et al.*, 2009). However, varieties that form root relatively deep in the soil are less attacked because the weevils cannot easily reach the roots to lay eggs (Kays *et al.*, 1993). Chemical control is not effective because the weevils are protected for, at least part of their life cycle by their development within roots or stems, where they are not easily reached by pesticides (Mason *et al.*, 1991)

Kabi *et al.* (2001), recorded that the larvae feed on storage root in the ground or inside the woody base of the stems. This means with the possible exception of systemic insecticides, which are costly and pose risk of residual contamination of the roots there is no effective chemical control of all stages of the pest found within the plant tissues. This subterranean habitat also makes the insect less accessible to predator and parasitoids but increases the impact of pathogens and nematodes, natural enemies which requires a protected, cool and humid environment for survival and reproduction (Stathers *et al.*, 2005).

CHAPTER III

MATERIALS AND METHODS

The present study regarding documentation of insect pests in sweet potato and their management has been conducted during rabi season (November 2016 to March 2017) in the experimental field of Sher-e-Bangla Agricultural University, Dhaka. Required materials and methodology are described below under the following sub-headings.

3.1. Location

The experiment was conducted in the experimental farm of Sher-e-Bangla Agricultural University, Dhaka, Bangladesh. The location of the experimental site was $23^{\circ}74'$ N latitude and $90^{\circ}35'$ E longitude and an elevation of 8.2 m from sea level (Anon., 1989).

3.2. Climate

The climate of the study site was under the subtropical climate, characterized by three distinct seasons, the Rabi from November to February and the Kharif- I, pre-monsoon period or hot season from March to April and the Kharif- II monsoon period from May to October (Edris *et al.*, 1979). The monthly average temperature, relative humidity and rainfall during the crop growing period were collected from weather yard, Bangladesh Meteorological Department and presented in Appendix I. During the experimental period the maximum temperature (27.1°C) and highest rainfall (30 mm) was recorded in the month of February 2017, whereas the minimum temperature (12.4°C) and no rainfall was recorded in the month of January 2017.

3.3. Soil

Soil of the study site was silty clay loam in texture belonging to series. The area represents the Agro Ecological Zone of Madhupur tract (AEZ-28) with pH 5.8-6.5, CEC-25.28 (Haider *et al.*, 1991). The selected plot was medium high land and the soil series was Tejgaon (FAO, 1988).

3.4. Land Preparation

The land is ploughed or dug to a depth of about 20 cm and harrowed to pulverize the soil. Mound method, ridge and furrow method, bed method and flat method are practiced in sweet potato cultivation in different localities. It is preferable to plant sweet potato on mounds in areas experiencing problems of drainage. In sloppy lands, ridge and furrow system is recommended for the control of soil erosion. The higher yield recorded when planted on mounds is probably be due to better soil aeration permitted by mounds and less tendency for soil compaction. In Bangladesh, higher yields were reported with trench planting followed by ridge and flat method of planting in alluvial soils under irrigated conditions (Bhuiyan *et al.*, 2006). The experimental plot was opened in the first week of November 2016 with a power tiller, and was exposed to the sun for a week, after which the land was ploughed three times followed by laddering to obtain desirable tilth. The corners of the land were spaded and later clods were broken into smaller pieces. After ploughing and laddering, all the stabbles and uprooted weeds were removed and then the land was ready. The field layout and design of the experiments were followed immediately after land preparation.

3.5. Manure and Fertilizer

The fertilizers N, P, K in the form of Urea, Triple Super Phosphate (TSP), Muriate of Potash (MP) respectively and as an organic manure, Cowdung were applied. The entire amount of organic manure, TSP, MP and half amount of Urea were applied as basal during the final land preparation. Rest amount of Urea was applied 60 days after planting of vine in the side of row. The dose and method of application of fertilizers are shown in Table (BARI).

Fertilizer name	Amount/ha
Cowdung (ton)	8-10
Urea(kg)	160-250
TSP(kg)	150-170
MP(kg)	150-250

3.6. Vine Collection

Vines of sweet potato that were used in this experiment were collected from Bangladesh Agricultural Research Institute (BARI), Joydebpur, Gazipur-1701. The variety name was BARI Sweet Potato -8.

3.7. Design of Experiment

The experiment was laid out in Randomized Completely Block Design (RCBD) with three replications. The experimental field was divided into three blocks maintaining 1m block to block distance and each block was subdivided into 7 plots for 7 treatments each maintaining 3 m x 2 m plot size. Thus the total number of plots was 21. The plot to plot distance was 0.5 m was kept to facilitate different intercultural operations.

3.8. Planting

Time of planting has been identified as one of the most important factors affecting growth, yield and quality of roots (Nedunchezhiyan and Byju, 2005). In Malaysia, higher root yield was noticed during drier growing season (January to July) compared to wet season (August to December) (Zaharah and Tan, 2006). In India, sweet potato is grown throughout the country, utilizing monsoon rain during *kharif*, (June-August), and with supplemental irrigation during *rabi* (October-December). The major area under sweet potato is planted during *kharif* season, though *rabi* planted sweet potato enjoys warm sunny days and cool nights with moderate rainfall conducive for higher root yield. Investigations carried out at different regions of India on the optimum time of planting revealed that rainfed crops should be planted immediately after onset of monsoon (Nedunchezhiyan and Reddy 2006) whereas dry season crops should be planted in October-November (Nath *et al.*, 2006) for higher storage root yield. In India, it is observed that the tuber yield of sweet potato is comparatively more during *rabi* than *kharif* season (Nawale and Salvi 1983; Nedunchezhiyan and Byju, 2005). In Taiwan, high yields of sweet potato were obtained when the mean daily temperature was maintained around 22°C for the 1st 60 DAP (Sajjapongse, 1989). In Korea, as per the demand of sweet

potatoes, transplanting time is advanced from May-June to March-April and the harvesting time is advanced from September-October to June-August with the modification of cultural requirements (Jeong, 2000). In the experiment field sweet potato vines were planted in the 21st November, 2016.

3.9. Treatments of the Experiment

Seven treatment combinations were tested under the present study. These are as follows

T₁: Emamectin benzoate @ 2 ml/L before tuber formation

T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing

T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application)

T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval before tuber formation (3times application)

T₅: Volian flexi 300SC @ 1 ml/L at 15 days interval (3 times application)

T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application)

T₇: Untreated control

Earthing-up was commonly done for each treatment

3.10. Intercultural Operations

Various intercultural operations such as gap filling, weeding, earthing up, irrigation etc. were accomplished for better growth and development of the sweet potato.

3.10.1. Irrigation

Sweet potato vines are succulent and fragile and if sufficient moisture is not available in the soil immediately after planting it dries up. Hence, sufficient soil moisture at the time of planting is to be ensured for proper sprouting and establishment of vines. Sweet potato is mostly grown under rain fed conditions. Hence, planting is carried out on rainy day or immediately after rain. It is also grown in dry season under protective irrigation. Under such conditions, if sufficient moisture is not available after planting, irrigation need to be provided on alternate days initially for

the first fortnight and thereafter once in 7-10 days. Trials conducted in the sweet potato growing areas have conclusively proved the beneficial effect of irrigation. Sweet potato required on an average of 2 mm of water per day in the early parts of the growing season and gradually increased to 5-6 mm of water/day prior to harvest (Gomes and Carr 2003). Biswal (2008), found that when soil moisture was high or the soil was compacted sweet potato had luxuriant vegetative growth with little or no tuberization of root. Excessive irrigation should be avoided as poor aeration may cause poor storage root induction or development (Chua and Kays 1981).

Indira and Kabeerathumma (1988), imposed water stress for 20 days to sweet potato grown in lysimeter during early growth phase (10-30 days), root development phase (30-50 DAP) and root maturity phase (75-90 DAP) and observed significant reduction in root yield for the stress induced during early growth phase. Stress induced during root development phase however slightly improved the yield while stress during root maturity phase resulted in a slight reduction in yield. Martin (1988), stated that sweet potato could not tolerate dry conditions at planting. According to Yassen and Thompson (1988) irrigation increased the number of root and total marketable yield.

According to Goswami *et al.* (1995), three irrigations at root initiation, early bulking and late bulking stages were most favourable for higher number of roots/plant, root bulking rate and dry matter content resulting in higher root yield (27.8 t ha⁻¹).

In this experiment field light watering in every alternative day after planting to 20 days were done. After that when the root is developed then watering in the sweet potato field after 30 days, 60 days and 90 days.

3.10.2. Gap filling

The planted vines in the experimental plot were kept under careful observation. Very few seedlings were damaged after planting and that vines were replaced by new vines from the stock.

3.10.3. Weeding and Earthing Up

To protect the crop from weeds, at least two weedings followed by earthing up have to be given between 15 and 30 and 35 and 60 days after planting (Nedunchezhiyan and Ray, 2010).

3.11. Harvesting

Harvesting of sweet potato has been done in three steps from 5 selected vines in the plot. 1st harvest was done at 15 days early harvest, 2nd harvest was done at optimum time of 130 days and 3rd harvest was done after 15 days from optimum harvest for assess sweet potato weevil. Finally per hectare yield was calculated by converting the yield of optimum days harvest.

3.12. Data Collection

The data on the following parameters were recorded at different time intervals as given below:

- Identify different insect pests in sweet potato field
- Number of insect pests on leaves and branches
- Total number of sweet potato storage roots/plant
- Total number of infested sweet potato storage roots/plant
- Vine length (cm)
- Number of storage roots/vine
- Weight of individual storage root (g)
- Yield of storage roots (t/ha)

3.13. Calculation:

3.13.1 Determination of storage roots infestation in number

All the storage roots were counted from 5 randomly selected plants from middle rows of each plot and examined. The collected data were divided into early, mid and late harvesting stage. The healthy and damaged storage roots were counted and the percent storage root infestation was calculated using the following formula:

$$\% \text{ infestation} = \frac{\text{Number of infested storage roots}}{\text{Total number of storage roots}} \times 100$$

3.13.2 Determination of storage roots infestation in weight

All the storage roots were counted from 5 randomly selected plants from middle rows of each plot and examined. The collected data were divided into early, mid and late harvesting stage. The healthy and damaged storage roots were weighted and the percent storage root infestation was calculated using the following formula:

$$\% \text{ infestation} = \frac{\text{Weight of infested storage roots}}{\text{Total weight of storage roots}} \times 100$$

3.14. Insect Pests of Sweet Potato

3.14.1. Sweetpotato Bug

Physomerus grossipes

Description and biology: The sweet potato bug lays groups of eggs on the undersides of leaves or on the stem. The mother bug guards her eggs and the young gregarious nymphs.

Damage: The nymphs and adults pierce the stems and petioles of the sweet potato and suck the plant sap, thus causing wilting and stunting.



Plate 1. Photograph showing Sweet potato bug

3.14.2. Mealybug

Phenacoccus solenopsis

Description and biology: Females are white, waxy and 3-4 mm long. Bare areas on the abdomen appear as dark bands. Adult males are grey and 1 mm long.

Damage: Adult females and nymphs suck sap. Leaves become yellow, vigour is reduced and young plants may die.



Plate 2. Photograph showing Mealybug

3.14.3. Green vegetable bug

Nezara viridula

Description and biology: Green, shield-shaped bug around 15 mm long. Initially dark red and orange, then green with distinctive black, white and red patterning.

Damage: Young shoots are damaged by sap sucking.



Plate 3. Photograph showing green vegetable bug

3.14.4. Leaf hopper

Amrasca sp.

Description and biology: Look like tiny cicadas; torpedo-shaped and ranging in colour from yellowish to green and mottled brown.

Damage: Feeding leaves speckled yellow tracks on the leaves.



Plate 4. Photograph showing leaf hopper

3.14.5. Minor Leaf Feeders

Zonoceros variegatus

Description and biology: An Asian species, the polyphagous slant-faced grasshopper is bright green and characterized by a pointed conical head and short antennae. It measures 30-40 mm in length.

Damage: It bores through the petiole of the host plant to lay its eggs. These are covered with a reddish brown gummy substance.



Plate 5. Photograph showing minor leaf feeders

3.14.6. Sweet potato beetle

Colasposoma sellatum

Description and biology: Glossy black beetle, 6-9 mm long with thickened wing covers and thorax. White grub up to 12 mm long, rather hairy, with six small legs.

Damage: Larvae feed on storage roots, creating tracks on the surface of the root. Damage is similar to that caused by flea beetle larvae, but deeper. Adults chew on leaves.



Plate 6. Photograph showing sweet potato beetle

3.14.7. Tortoiseshell Beetle

Aspidomorpha spp.

Description and biology: Eggs are laid on the underside of the sweetpotato leaves or other Convolvulaceae in batches cemented to the leaves. The eggs of some species are concealed in a papery oothecum. Larvae are characteristically flattened and spiny. In some species, the tail is held up over the back and the larva may carry excreta and previous cast skins. The pupa is less spiny than the larva, and is fixed inert to the leaf



Plate 7. Photograph showing Tortoiseshell Beetle

The adults are broadly oval and may be bright and patterned. Larvae, pupae and adult are found on both sides of the foliage. Development from egg to adult takes 3-6 weeks depending on the species.

Damage: Both adults and larvae eat large round holes in the leaves. Attacks are sometimes sufficiently severe to completely skeletonize the leaves and peel the stems.

3.14.8. Whitefly

Bemisia tabaci

Description and biology: Snow white, around 1 mm long with wings held in a peak along the body. First instar nymphs are flat, greenish, mobile and around 0.3 mm long. Later instar nymphs are also flat but opaque white and stationary, appearing similar to soft scale insects but with pointed tails. Mature nymphs turn golden, their bodies thicken and eyes turn red.



Plate 8. Photograph showing whitefly

Damage: Nymphs and adults suck sap from plants, causing yellow or purplish stippling between the veins on leaves. Growth may be stunted. Whitefly can also transmit some viruses.

3.14.9. Sweet potato weevil

Cylas spp.

Description and biology: Three species of the genus *Cylas* are pests of sweet potato; they are commonly called sweet potato weevils. All three species—*Cylas formicarius*, *C. puncticollis*, and *C. brunneus*—are found in Africa. *C. formicarius* is present in Asia and in parts of the Caribbean. The elongated ant-like adults of the three species can be distinguished from each other. *Cylas puncticollis* is the easiest to distinguish because the adult is all black and larger than the other two. *C. formicarius* has a bluish black abdomen and a reddish brown thorax. *C. brunneus* adults are small and not uniform in coloring. The most common type can easily be confused with *C. formicarius*. In



Plate 9. Photograph showing sweet potato weevil larva

all species, the eggs are shiny and round. The legless larvae are white and curved, and the pupae are white.

Damage: Damage symptoms are similar for all three species. Adult sweetpotato weevils feed on the epidermis of vines and leaves. Adults also feed on the external surfaces of storage roots, causing round feeding punctures, which can be distinguished from oviposition sites by their greater depth. The developing larvae of the weevil tunnel in the vines and storage roots, causing significant damage. Frass is deposited in the tunnels. In response to damage, storage roots produce toxic terpenes, which render storage roots inedible even at low concentrations and low levels of physical damage. Feeding inside the vines causes malformation, thickening, and cracking of the affected vine.

3.15. Beneficial insect in the field

3.15.1. Ladybird beetle

Coccinella transversa

Description and biology: Most are brightly coloured, dome-shaped beetles 3-5 mm long with distinctive spots and stripes on their outer wing covers. Black with coloured markings a ‘crocodile like’ appearance.



Plate 10. Photograph showing Ladybird beetle

3.15.2. Spider

The importance of spiders as predators has been clearly demonstrated for rice, but their role has not been studied adequately in many other crops and little is known about their contribution to biological control of sweet potato pests. The lynx spider *Oxyopes* sp. and the wolf spider *Lycosa* sp. are abundant in sweet potato fields. These do not spin webs but rather hunt prey directly. Web-spinning spiders are also common



Plate 11. Photograph showing spider

CHAPTER IV

RESULTS AND DISCUSSION

The study was conducted for the identification of insect pests in sweet potato and their management. Data was recorded on insect pest incidence, number of healthy, infested leaves and branches for different insect pests and their infestation

level, healthy and infested storage roots of sweet potato and their infestation level and also yield contributing characters and yield of sweet potato. The analysis of

variance (ANOVA) of the data on different parameters has been presented in Appendix II-IX. The results have been discussed and presented under the following headings and sub-headings:

4.1 Incidence of different insect pest in sweet potato

4.1.1 Species of insect pests

During the entire growing period sweet potato competes with numerous insect pests under favorable condition which is the common phenomenon for the cultivation of this crop. Under the present study 9 different species of insect pests were found and they were belongs to 9 family under 3 orders. The common name, scientific name, order, family and nature of damage of these insect pests are presented in Table 1. Among the recorded insect pests 5 species belong to the order Hemiptera, 3 species belong to Coleoptera and only 1 species under Othoptera order and different family.

Among the insect pests it was observed that sweet potato weevil was more destructive for storage roots of sweet potato. The natures of the damage of these insects are presented in Table 1.

4.1.2 Insect population

Number of insect population of 5 selected vine/plot was observed in the experimental plot for the entire growing period. The number of observed insect pests was presented in Table 2.

Table 1. List of the insect pests found in the experimental sweet potato field during study period

Sl. No.	Common name	Scientific name	Order	Family	Nature of damage
1.	Sweet potato Bug	<i>Physomerus grossipes</i>	Hemiptera	Coreidae	The nymphs and adults pierce the stems and petioles of sweet potato and suck the plant sap
2.	Mealybug	<i>Phenacoccus solenopsis</i>	Hemiptera	Pseudococcidae	Adult females and nymphs suck sap. Leaves become yellow, vigour is reduced and young plants may die
3.	Green vegetable bug	<i>Nezara viridula</i>	Hemiptera	Pentatomidae	Young shoots are damaged by sap sucking
4.	Leaf hopper	<i>Amrasca</i> spp.	Homoptera	Cicadellidae	Feeding leaves speckled yellow tracks on the leaves
5.	Minor leaf feeders	<i>Zonoceros variegatus</i>	Orthoptera	Pyrgomoriphidae	It bores through the petiole of the host plant to lay its eggs.
6.	Sweet potato beetle	<i>Colasposoma sellatum</i>	Coleoptera	Chrysomelidae	Larvae feed and creating tracks on the surface on storage roots and adults chew on leaves
7.	Whitefly	<i>Bemisia tabaci</i>	Homoptera	Aleyrodidae	Nymphs and adults suck sap, causing yellow or purplish stippling between the leaf veins
8.	Tortoiseshell Beetles	<i>Aspidomorpha</i> spp.	Coleoptera	Chrysomelidae	Both adults and larvae eat large round holes in the leaves. Attacks are sometimes sufficiently severe to completely skeletonize the leaves and peel the stems
9.	Sweet potato weevil	<i>Cylas formicarius</i>	Coleoptera	Curculionidae	Adult sweet potato weevils feed on the epidermis of vines and leaves and also the external surfaces of storage roots, causing round feeding punctures. The developing larvae of the weevil tunnel in the vines and storage roots, causing significant damage.

4.1.2.1 Sweet potato bug

Data revealed that for the consideration of sweet potato bug, the lowest number (1.27) of sweet potato bug was observed from T₂ (Carbofuran 5G @ 15 kg/ha at 60 days after sowing) treatment which was closely followed (1.86, 1.93 and 2.07) by T₄ (Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval: 3 times application), T₃ (Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval: 3 times application) and T₅ (Voliam flexi 300SC @ 1 ml/L at 15 days interval: 3 times application), respectively and they were statistically similar, whereas the highest number of sweet potato bug (5.13) was recorded from T₇ (Untreated control) treatment (Table 2). Allard (1990) reported that deltamethrin and pirimiphos methyl gave good control of sweet potato pests.

4.1.2.2 Mealybug

In case of mealybug, the lowest number of mealybug was observed from T₂ (1.67) treatment which was closely followed by T₄ (2.27) treatment and then T₃ (2.87), whereas the highest number of mealybug was found from T₇ (6.93) treatment (Table 2).

4.1.2.3 Green vegetable bug

In consideration of green vegetable bug, the lowest number of green vegetable bug was recorded from T₂ (1.07) treatment which was statistically similar to T₄ (1.27) and closely followed by T₃ (1.87) treatment and T₅ (2.07) and they were statistically similar, while the highest number of green vegetable bug was observed from T₇ (4.20) treatment (Table 2). Misra *et al.* (2001) reported from earlier experiment that combination of vine dipping of 0.05% monocrotophos and three foliar sprays with 0.05% endosulfan proved very effective in controlling green vegetable bug.

Table 2. Number of identified insect pests in sweet potato during the growing period for different management practices

Treatments	Number of different insect pests								
	Sweet potato bug	Mealybug	Green vegetable bug	Leaf hopper	Minor leaf feeders	Sweet potato beetle	Whitefly	Tortoiseshell beetle	Sweet potato weevil
T ₁	2.93 b	3.94 b	2.93 b	4.80 b	2.33 b	3.87 b	3.93 b	2.53 b	4.20 b
T ₂	1.27 d	1.67 f	1.07 d	2.07 e	1.13 e	1.47 f	1.67 e	1.13 e	1.67 e
T ₃	1.93 c	2.87 d	1.87 c	3.47 d	1.87 d	2.33 e	2.53 d	1.80 cd	2.67 d
T ₄	1.86 c	2.27 e	1.27 d	2.53 e	1.33 e	2.13 e	2.27 d	1.47 de	2.33 d
T ₅	2.07 c	3.27 c	2.07 c	4.27 c	1.93 cd	2.87 d	3.27 c	1.87 c	3.13 c
T ₆	2.73 b	3.87 b	2.67 b	4.33 bc	2.20 bc	3.33 c	3.33 c	2.07 c	4.07 b
T ₇	5.13 a	6.93 a	4.20 a	6.40 a	4.73 a	6.33 a	6.73 a	4.33 a	6.73 a
LSD _(0.05)	0.323	0.356	0.347	0.471	0.276	0.347	0.347	0.342	0.369
CV(%)	6.95	5.66	8.47	6.64	7.05	6.12	5.76	8.88	5.84

In a column, numeric data represents the mean value of 3 replications; each replication is derived from 5 selected vine/plot of each treatment

In a column means having similar letter(s) are statistically identical and those having dissimilar letter(s) differ significantly as per 0.05 level of probability

T₁: Emamectin benzoate @ 2 ml/L of water

T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing

T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application)

T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval (3 times application)

T₅: Voliam flexi 300SC @ 1 ml/L at 15 days interval (3 times application)

T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application)

T₇: Untreated control

4.1.2.4 Leaf hopper

In case of leaf hopper, the lowest number of leaf hopper was recorded from T₂ (2.07) treatment which was statistically similar to T₄ (2.53) and closely followed by T₃ (3.47) treatment, whereas the highest number of leaf hopper was found from T₇ (6.40) which was followed by T₁ (4.80) and T₆ (4.33) treatment and they were statistically similar (Table 2).

4.1.2.5 Minor leaf feeders

Data revealed that for minor leaf feeders, the lowest number of minor leaf feeders was recorded from T₂ (1.13) treatment which was statistically similar to T₄ (1.33) and closely followed by T₃ (1.87) and T₅ (1.93) treatment and they were statistically similar, whereas the highest number of minor leaf feeders was found from T₇ (4.73) which was followed by T₁ (2.33) and T₆ (2.20) treatment and they were statistically similar (Table 2).

4.1.2.6 Sweet potato beetle

In consideration of sweet potato beetle, the lowest number of sweet potato beetle was recorded from T₂ (1.47) treatment which closely followed by T₄ (2.13) and T₃ (2.33) and they were statistically similar, whereas the highest number of was found from T₇ (6.33) which was followed by T₁ (3.87) treatment (Table 2).

4.1.2.7 Whitefly

In case of whitefly, the lowest number of whitefly was recorded from T₂ (1.67) treatment which closely followed by T₄ (2.27) and T₃ (2.53) and they were statistically similar, whereas the highest number of whitefly was found from T₇ (6.73) which was followed by T₁ (3.93) treatment (Table 2).

4.1.2.8 Tortoiseshell beetle

From the recorded data in case of tortoiseshell beetle, the lowest number of tortoiseshell beetle was found from T₂ (1.13) which was statistically similar to T₄ (1.47) and closely followed by T₃ (1.80) treatment, whereas the highest number from T₇ (4.33) which was followed by T₁ (2.53) treatment (Table 2).

4.1.2.9 Sweet potato weevil

Data revealed that for sweet potato weevil, the lowest number of sweet potato weevil was observed from T₂ (1.67) treatment which was closely followed by T₄ (2.33) and T₃ (2.67) treatment and they were statistically similar, while the highest number of sweet potato weevil was recorded from T₇ (6.73) which was followed by T₁ (4.20) and T₆ (4.07) treatment and they were statistically similar (Table 2). Parker *et al* (1990) found that the sweet potato weevil is one of the major pests of sweet potato worldwide.

4.2 Leaf and branches infestation by different insect pests in different stages

4.2.1 At early growth stages (1-45 days)

Statistically significant variation was recorded due to different treatments in terms of healthy, infested leaves and branches and also % of infestation caused by different insect pests at early growth stages of sweet potato (Table 3).

4.2.1.1 Leaves of sweet potato

In case of leaves of sweet potato at early growth stage, it was observed that the highest number of healthy leaves/vine was recorded from T₂ (46.20) treatment which was statistically similar with other treatment except T₇, while the lowest number of healthy leaves (36.73) was observed from T₇ treatment (Table 3). In case of infested leaves, the lowest number (1.87) was observed from T₂ treatment which was statistically similar to T₄ (2.00) and T₃ (2.33) treatment and closely followed by T₅ (2.73) treatment, while the highest number of infested leaves was found from T₇ (5.13) treatment which was followed by T₁ (3.87) treatment. In consideration of infestation level of leaves, the lowest infestation was recorded from T₂ (3.90%) treatment which was statistically similar to T₄ (4.38%) and T₃ (5.11%) treatment and closely followed by T₅ (5.93%) treatment, whereas the highest infestation (12.30%) was observed from T₇ treatment which was closely followed by T₁ (8.39%) and T₆ (6.83%) treatment and they were statistically similar. In case of leaf infestation reduction over control the highest value was recorded from T₂ (68.29) and the lowest value was found from T₁ (31.79) treatment.

Table 3. Incidence of insect pests on leaf and branches of sweet potato at early growth stage for different management practices

Treatments	Leaf of sweet potato				Branches of sweet potato			
	Healthy (No.)	Infested (No.)	Infestation (%)	Infestation reduction over control	Healthy (No.)	Infested (No.)	Infestation (%)	Infestation reduction over control
T ₁	42.33 a	3.87 b	8.39 b	31.79	4.53 cd	0.40 b	8.11 b	36.94
T ₂	46.20 a	1.87 e	3.90 e	68.29	5.27 a	0.13 d	2.39 e	81.42
T ₃	43.53 a	2.33 de	5.11 cde	58.46	5.00 abc	0.27 bcd	5.14 cde	60.03
T ₄	44.20 a	2.00 e	4.38 de	64.39	5.13 ab	0.20 cd	3.75 de	70.84
T ₅	43.40 a	2.73 cd	5.93 cd	51.79	4.87 abc	0.33 bc	6.37 bcd	50.47
T ₆	42.67 a	3.13 c	6.83 bc	44.47	4.67 bc	0.40 b	7.90 bc	38.57
T ₇	36.73 b	5.13 a	12.30 a	--	4.07 d	0.60 a	12.86 a	--
LSD _(0.05)	4.683	0.577	1.677	--	0.481	0.138	2.720	--
CV(%)	6.16	10.79	14.09	--	5.64	13.90	12.00	--

In a column, numeric data represents the mean value of 3 replications; each replication is derived from 5 selected vine/plot of each treatment

In a column means having similar letter(s) are statistically identical and those having dissimilar letter(s) differ significantly as per 0.05 level of probability

T₁: Emamectin benzoate @ 2 ml/L of water

T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing

T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application)

T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval (3 times application)

T₅: Voliam flexi 300SC @ 1 ml/L at 15 days interval (3 times application)

T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application)

T₇: Untreated control

4.2.1.2 Branches of sweet potato

For branches/vine of sweet potato at early growth stage, data revealed that the highest number of healthy branches was recorded from T₂ (5.27) treatment which was statistically similar to T₄ (5.13), T₃ (5.00) and T₅ (4.87) treatment and closely followed by T₆ (4.67) treatment, while the lowest number of healthy branches (4.07) was observed from T₇ treatment which was statistically similar to T₁ (4.53) treatment (Table 3). In case of infested branches, the lowest number (0.13) was observed from T₂ treatment which was statistically similar to T₄ (0.20) and T₃ (0.27) treatment, while the highest number of infested branches was found from T₇ (0.60) treatment which was followed by T₁ (0.40) and T₆ (0.40) treatment. In consideration of infestation level of branches, the lowest infestation was recorded from T₂ (2.39%) treatment which was statistically similar to T₄ (3.75%) and T₃ (5.14%) treatment and closely followed by T₅ (6.37%) treatment, whereas the highest infestation (12.86%) was observed from T₇ treatment which was closely followed by T₁ (8.11%) and T₆ (7.90%) treatment and they were statistically similar. In case of infestation reduction over control, the highest value was recorded from T₂ (81.42) and the lowest value was found from T₁ (36.94) treatment.

4.2.2 At mid growth stages (46-85 days)

Statistically significant variation was recorded for healthy, infested leaves and branches and also % of infestation caused by different insect pests at mid growth stages of sweet potato due to different treatments (Table 4).

4.2.2.1 Leaves of sweet potato

In case of leaves/vine of sweet potato at mid growth stage, it was observed that the highest number of healthy leaves was recorded from T₂ (88.67) treatment which was statistically similar with other treatment except T₇, while the lowest number of healthy leaves (74.47) was observed from T₇ treatment (Table 4). In case of infested leaves, the lowest number (2.47) was observed from T₂ treatment which was closely followed by T₄ (2.87) treatment, whereas the highest number of infested leaves was found from T₇ (8.33) treatment which was

Table 4. Incidence of insect pests on leaf and branches of sweet potato at mid growth stage for different management practices

Treatments	Leaf of sweet potato				Branches of sweet potato			
	Healthy (No.)	Infested (No.)	Infestation (%)	Infestation reduction over control	Healthy (No.)	Infested (No.)	Infestation (%)	Infestation reduction over control
T ₁	83.07 a	4.93 b	5.61 b	44.29	11.20 bc	0.93 b	7.68 b	34.58
T ₂	88.67 a	2.47 g	2.71 e	73.09	13.33 a	0.33 e	2.45 e	79.13
T ₃	85.73 a	3.27 e	3.68 d	63.46	12.27 ab	0.67 cd	5.14 d	56.22
T ₄	87.53 a	2.87 f	3.18 de	68.42	13.13 a	0.47 de	3.40 e	71.04
T ₅	85.73 a	3.80 d	4.26 c	57.70	12.27 ab	0.80 bc	6.10 cd	48.04
T ₆	84.80 a	4.13 c	4.66 c	53.72	11.73 abc	0.93 b	7.39 bc	37.05
T ₇	74.47 b	8.33 a	10.07 a	--	10.53 c	1.40 a	11.74 a	--
LSD _(0.05)	8.024	0.323	0.522	--	1.514	0.225	1.457	--
CV(%)	5.35	4.29	6.01	--	7.05	15.78	13.06	--

In a column, numeric data represents the mean value of 3 replications; each replication is derived from 5 selected vine/plot of each treatment

In a column means having similar letter(s) are statistically identical and those having dissimilar letter(s) differ significantly as per 0.05 level of probability

T₁: Emamectin benzoate @ 2 ml/L of water

T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing

T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application)

T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval (3 times application)

T₅: Voliam flexi 300SC @ 1 ml/L at 15 days interval (3 times application)

T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application)

T₇: Untreated control

followed by T₁ (4.93) treatment. In consideration of infestation level of leaves, the lowest infestation was recorded from T₂ (2.71%) treatment which was statistically similar to T₄ (3.18%) and closely followed by T₃ (3.68%) treatment, whereas the highest infestation (10.07%) was observed from T₇ treatment which was followed by T₁ (5.61%). In case of infestation reduction over control the highest value was recorded from T₂ (73.09) and the lowest value was found from T₁ (44.29) treatment.

4.2.2.2 Branches of sweet potato

For branches of sweet potato at mid growth stage, data revealed that the highest number of healthy branches/vine was recorded from T₂ (13.33) treatment which was statistically similar with other treatment except T₇ and T₁ treatment, while the lowest number of healthy branches (10.53) was observed from T₇ treatment which was statistically similar to T₁ (11.20) treatment (Table 4). In case of infested branches, the lowest number (0.33) was observed from T₂ treatment which was statistically similar to T₄ (0.47) and closely followed by T₃ (0.67) treatment, while the highest number of infested branches was found from T₇ (1.40) treatment which was followed by T₁ (0.93), T₆ (0.93) and T₅ (0.80) treatment and they were statistically similar. In case of infestation level of branches, the lowest infestation was recorded from T₂ (2.45%) treatment which was statistically similar to T₄ (3.40%) and closely followed by T₃ (5.14%) and T₅ (6.10%) treatment and they were statistically similar, whereas the highest infestation (11.74%) was observed from T₇ treatment which was closely followed by T₁ (7.68%) and T₆ (7.39%) treatment and they were statistically similar. In case of reduction over control, the highest value was recorded from T₂ (79.13) and the lowest value was found from T₁ (34.58) treatment.

4.2.3 At late growth stages (86-130 days)

Different treatments showed statistically significant differences in terms of healthy, infested leaves and branches and also % of infestation caused by different insect pests at late growth stages of sweet potato (Table 5).

Table 5. Incidence of insect pests on leaf and branches of sweet potato at late growth stage for different management practices

Treatments	Leaf of sweet potato				Branches of sweet potato			
	Healthy (No.)	Infested (No.)	Infestation (%)	Infestation reduction over control	Healthy (No.)	Infested (No.)	Infestation (%)	Infestation reduction over control
T ₁	135.27 a	7.00 b	4.92 b	45.87	19.33 bc	1.67 b	7.93 b	28.49
T ₂	143.33 a	3.80 e	2.58 e	71.62	22.53 a	0.60 d	2.61 d	76.47
T ₃	139.80 a	5.13 d	3.55 d	60.95	21.67 ab	1.00 c	4.42 c	60.14
T ₄	142.00 a	4.13 e	2.83 e	68.87	22.47 a	0.73 d	3.16 d	71.51
T ₅	138.73 a	5.67 d	3.92 d	56.88	20.47 abc	1.20 c	5.55 c	49.95
T ₆	136.87 a	6.40 c	4.47 c	50.83	19.67 bc	1.47 b	7.00 b	36.88
T ₇	122.73 b	12.27 a	9.09 a	--	18.13 c	2.27 a	11.09 a	--
LSD _(0.05)	12.03	0.540	0.410	--	2.397	0.252	1.263	--
CV(%)	4.94	4.78	5.16	--	6.54	11.08	11.91	--

In a column, numeric data represents the mean value of 3 replications; each replication is derived from 5 selected vine/plot of each treatment

In a column means having similar letter(s) are statistically identical and those having dissimilar letter(s) differ significantly as per 0.05 level of probability

T₁: Emamectin benzoate @ 2 ml/L of water

T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing

T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application)

T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval (3 times application)

T₅: Voliam flexi 300SC @ 1 ml/L at 15 days interval (3 times application)

T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application)

T₇: Untreated control

4.2.3.1 Leaves of sweet potato

In case of leaves/vine of sweet potato at late growth stage, it was observed that the highest number of healthy leaves/vine was recorded from T₂ (143.33) treatment which was statistically similar with other treatment except T₇, while the lowest number of healthy leaves (122.73) was observed from T₇ treatment (Table 5). In case of infested leaves, the lowest number (3.80) was observed from T₂ treatment which was statistically similar to T₄ (4.13) and closely followed by T₃ (5.13) and T₅ (5.67) treatment and they were statistically similar, while the highest number of infested leaves was found from T₇ (12.27) treatment which was followed by T₁ (7.00) treatment. In consideration of infestation level of leaves, the lowest infestation was recorded from T₂ (2.58%) treatment which was statistically similar to T₄ (2.83%) and closely followed by T₃ (3.55%) and T₅ (3.92%) treatment and they were statistically similar, whereas the highest infestation (9.09%) was observed from T₇ treatment which was closely followed by T₁ (4.92%) treatment and they were statistically similar. In case of reduction over control the highest value was recorded from T₂ (71.62) and the lowest value was found from T₁ (45.87) treatment.

4.2.3.2 Branches of sweet potato

For branches/vine of sweet potato at late growth stage, data revealed that the highest number of healthy branches/vine was recorded from T₂ (22.53) treatment which was statistically similar to T₄ (22.47), T₃ (21.67) and T₅ (20.47) treatment, while the lowest number of healthy branches (18.13) was observed from T₇ treatment which was statistically similar to T₆ (19.67) and T₁ (19.33) treatment (Table 5). In case of infested branches, the lowest number (0.60) was observed from T₂ treatment which was statistically similar to T₄ (0.73) and closely followed by T₃ (1.00) and T₅ (1.20) treatment and they were statistically similar, while the highest number of infested branches was found from T₇ (2.27) treatment which was followed by T₁ (1.67) and T₆ (1.47) treatment and they were statistically similar. In consideration of infestation level of branches, the lowest infestation was recorded from T₂ (2.61%) treatment which was

statistically similar to T₄ (3.16%) and closely followed by T₅ (5.55%) and T₃ (4.42%) treatment and they were statistically similar, whereas the highest infestation (11.09%) was observed from T₇ treatment which was closely followed by T₁ (7.93%) and T₆ (7.00%) treatment and they were statistically similar. In case of reduction over control, the highest value was recorded from T₂ (76.47) and the lowest value was found from T₁ (28.49) treatment.

4.3 Storage root infestation by sweet potato weevil in different stages

4.3.1 At 15 days early harvest

Healthy, infested storage roots of sweet potato in number and weight basis per vine and also % of infestation caused by sweet potato weevil at 15 days early harvest varied significantly for different treatment (Table 6).

4.3.1.1 Storage roots of sweet potato in number basis

In case of storage roots of sweet potato/vine in number basis at 15 days early harvest, it was observed that the highest number of healthy storage roots/vine was recorded from T₂ (6.87) treatment which was statistically similar with other treatment except T₇, while the lowest number (5.73) was observed from T₇ treatment (Table 6). In case of infested storage roots, the lowest number (0.27) was observed from T₂ treatment which was statistically similar to T₄ (0.40) and T₅ (0.33) closely followed by T₃ (0.47) treatment, while the highest number was found from T₇ (0.80) treatment which was followed by T₁ (0.60) and T₆ (0.53) treatment and they were statistically similar. In consideration of infestation level of storage roots in number basis, the lowest infestation was recorded from T₂ (3.69%) treatment which was statistically similar to T₄ (4.74%) and T₅ (5.95) and closely followed by T₃ (6.63%) treatment, whereas the highest infestation (12.25%) was observed from T₇ treatment which was closely followed by T₁ (8.67%) and T₆ (7.82%) treatment and they were statistically similar. In case of reduction over control the highest value was recorded from T₂ (69.88) and the lowest value was found from T₁ (29.22) treatment. Ebregt *et al.*, (2005) stated that harvesting two weeks earlier reduce the loss due to weevil from greater than 30 percent to less than five percent.

Table 6. Incidence of sweet potato weevil on storage roots of sweet potato at 15 days early harvest in number and weight basis for different management practices

Treatments	Storage roots of sweet potato in number basis				Storage roots of sweet potato in weight basis			
	Healthy (No.)	Infested (No.)	Infestation (%)	Infestation reduction over control	Healthy (g)	Infested (g)	Infestation (%)	Infestation reduction over control
T ₁	6.33 ab	0.60 b	8.67 b	29.22	678.14 a	69.48 b	9.29 b	28.43
T ₂	6.87 a	0.27 e	3.69 e	69.88	713.71 a	42.95 d	5.68 e	56.24
T ₃	6.53 a	0.47 bcd	6.63 bcd	45.88	701.02 a	54.69 c	7.24 c	44.22
T ₄	6.67 a	0.33 de	4.74 de	61.31	707.11 a	48.66 d	6.45 d	50.31
T ₅	6.33 ab	0.40 cde	5.95 cde	51.43	696.48 a	56.34 c	7.48 c	42.37
T ₆	6.33 ab	0.53 bc	7.82 bc	36.16	683.90 a	65.52 b	8.75 b	32.59
T ₇	5.73 b	0.80 a	12.25 a	--	608.97 b	90.85 a	12.98 a	--
LSD _(0.05)	0.577	0.159	2.164	--	61.73	5.937	0.748	--
CV(%)	5.06	18.34	17.12	--	5.07	5.45	5.09	--

In a column, numeric data represents the mean value of 3 replications; each replication is derived from 5 selected vine/plot of each treatment

In a column means having similar letter(s) are statistically identical and those having dissimilar letter(s) differ significantly as per 0.05 level of probability

T₁: Emamectin benzoate @ 2 ml/L of water

T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing

T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application)

T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval (3 times application)

T₅: Voliam flexi 300SC @ 1 ml/L at 15 days interval (3 times application)

T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application)

T₇: Untreated control

4.3.1.2 Storage roots of sweet potato in weight basis

In case of storage roots of sweet potato/vine in weight basis at 15 days early harvest, it was observed that the highest weight of healthy storage roots/vine was recorded from T₂ (713.71 g) treatment which was statistically similar with other treatment except T₇, while the lowest weight (608.97 g) was observed from T₇ treatment (Table 6). In case of infested storage roots, the lowest weight (42.95 g) was observed from T₂ treatment which was statistically similar to T₄ (48.66 g) and closely followed by T₃ (54.69 g) and T₅ (56.34 g) treatment and they were statistically similar, while the highest weight was found from T₇ (90.85 g) treatment which was followed by T₁ (69.48 g) and T₆ (65.52 g) treatment and they were statistically similar. In consideration of infestation level of storage roots in weight basis, the lowest infestation was recorded from T₂ (5.68%) treatment which was closely followed by T₄ (6.45%) treatment, whereas the highest infestation (12.98%) was observed from T₇ treatment which was closely followed by T₁ (9.29%) and T₆ (8.75%) treatment and they were statistically similar. In case of reduction over control the highest value was recorded from T₂ (56.24) and the lowest value was found from T₁ (28.43) treatment.

4.3.2 At optimum days harvest

Statistically significant variation was recorded due to different treatments for healthy, infested storage roots of sweet potato in number and weight basis per vine and also % of infestation caused by sweet potato weevil at optimum days harvest (Table 7).

4.3.2.1 Storage roots of sweet potato in number basis

In case of storage roots of sweet potato/vine in number basis at optimum days harvest, it was observed that the highest number of healthy storage roots/vine was recorded from T₂ (7.00) treatment which was statistically similar with other treatment except T₇, while the lowest number (5.93) was observed from T₇ treatment (Table 7). In case of infested storage roots, the lowest number (0.33) was observed from T₂ treatment which was statistically similar to T₄ (0.40), T₅ (0.53) and T₃ (0.53) treatment, while the highest number was found from

Table 7. Incidence of sweet potato weevil on storage roots of sweet potato at optimum days harvest in number and weight basis for different management practices

Treatments	Storage roots of sweet potato in number basis				Storage roots of sweet potato in weight basis			
	Healthy (No.)	Infested (No.)	Infestation (%)	Infestation reduction over control	Healthy (g)	Infested (g)	Infestation (%)	Infestation reduction over control
T ₁	6.60 a	0.73 b	9.97 b	26.47	692.78 a	76.26 b	9.91 b	30.01
T ₂	7.00 a	0.33 c	4.49 e	66.89	734.06 a	50.91 d	6.49 e	54.17
T ₃	6.67 a	0.53 bc	7.35 cd	45.80	723.43 a	62.09 c	7.92 cd	44.07
T ₄	6.80 a	0.40 c	5.56 de	59.00	726.26 a	55.63 cd	7.10 de	49.86
T ₅	6.60 a	0.53 bc	7.49 bcd	44.76	717.08 a	63.75 c	8.16 c	42.37
T ₆	6.60 a	0.67 b	9.21 bc	32.08	702.30 a	72.91 b	9.41 b	33.55
T ₇	5.93 b	0.93 a	13.56 a	--	627.41 b	103.46 a	14.16 a	--
LSD _(0.05)	0.557	0.195	2.411	--	61.69	7.931	0.928	--
CV(%)	4.74	18.60	16.46	--	4.93	6.43	5.78	--

In a column, numeric data represents the mean value of 3 replications; each replication is derived from 5 selected vine/plot of each treatment

In a column means having similar letter(s) are statistically identical and those having dissimilar letter(s) differ significantly as per 0.05 level of probability

T₁: Emamectin benzoate @ 2 ml/L of water

T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing

T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application)

T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval (3 times application)

T₅: Voliam flexi 300SC @ 1 ml/L at 15 days interval (3 times application)

T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application)

T₇: Untreated control

T₇ (0.93) treatment which was followed by T₁ (0.73) and T₆ (0.67) treatment and they were statistically similar. In consideration of infestation level of storage roots in number basis, the lowest infestation was recorded from T₂ (4.49%) treatment which was statistically similar to T₄ (5.56%) and closely followed by T₃ (7.35%) treatment, whereas the highest infestation (13.56%) was observed from T₇ treatment which was closely followed by T₁ (9.97%) and T₆ (9.21%) treatment and they were statistically similar. In case of reduction over control the highest value was recorded from T₂ (66.89) and the lowest value was found from T₁ (26.47) treatment.

4.3.2.2 Storage roots of sweet potato in weight basis

In case of storage roots of sweet potato/vine in weight basis at optimum days harvest, it was observed that the highest weight of healthy storage roots/vine was recorded from T₂ (734.06 g) treatment which was statistically similar with other treatment except T₇, while the lowest weight (627.41 g) was observed from T₇ treatment (Table 7). In case of infested storage roots, the lowest weight (50.91 g) was observed from T₂ treatment which was statistically similar to T₄ (55.63 g) and closely followed by T₃ (62.09 g) and T₅ (63.75 g) treatment and they were statistically similar, while the highest weight was found from T₇ (103.46 g) treatment which was followed by T₁ (76.26 g) and T₆ (72.91 g) treatment and they were statistically similar. In consideration of infestation level of storage roots in weight basis, the lowest infestation was recorded from T₂ (6.49%) treatment which was statistically similar to T₄ (7.10%) treatment, whereas the highest infestation (14.16%) was observed from T₇ treatment which was closely followed by T₁ (9.91%) and T₆ (9.41%) treatment and they were statistically similar. In case of reduction over control the highest value was recorded from T₂ (54.17) and the lowest value was found from T₁ (30.01) treatment.

4.3.3 After 15 days from optimum harvests

Statistically significant variation was recorded due to different treatments for healthy, infested storage roots of sweet potato in number and weight basis per vine and also % of infestation caused by sweet potato weevil after 15 days from optimum harvests (Table 8).

4.3.3.1 Storage roots of sweet potato in number basis

In case of storage roots of sweet potato/vine in number basis after 15 days from optimum harvest, it was observed that the highest number of healthy storage roots/vine was recorded from T₂ (7.13) which was statistically similar with other treatment except T₇, while the lowest number (6.13) from T₇ treatment (Table 8). In case of infested storage roots, the lowest number (0.40) was observed from T₂ treatment which was statistically similar to T₄ (0.47) and followed by T₅ (0.67), T₆ (0.67) and T₃ (0.73) treatment, while the highest number was found from T₇ (1.20) treatment which was followed by T₁ (0.87) treatment. In consideration of infestation level of storage roots in number basis, the lowest infestation was recorded from T₂ (5.31%) which was statistically similar to T₄ (5.98%) and followed by T₅ (8.92%) and T₆ (8.98), whereas the highest infestation (16.37%) was observed from T₇ treatment which was closely followed by T₁ (11.38%) treatment. In case of reduction over control the highest value was recorded from T₂ (67.56) and the lowest value was found from T₁ (30.48) treatment.

4.3.3.2 Storage roots of sweet potato in weight basis

In case of storage roots of sweet potato/vine in weight basis after 15 days from optimum harvest, it was observed that the highest weight of healthy storage roots/vine was recorded from T₂ (743.79 g) treatment which was statistically similar with other treatment except T₇, while the lowest weight (638.30 g) was observed from T₇ treatment (Table 8). In case of infested storage roots, the lowest weight (55.30 g) was observed from T₂ treatment which was statistically similar to T₄ (61.23 g) and followed by T₃ (67.45 g) and T₅ (70.10 g) treatment and they were statistically similar, while the highest weight from T₇ (131.23 g) treatment which was followed by T₁ (91.74 g) treatment. In consideration of

Table 8. Incidence of sweet potato weevil on storage roots of sweet potato after 15 days from optimum harvest in number and weight basis for different management practices

Treatments	Storage roots of sweet potato in number basis				Storage roots of sweet potato in weight basis			
	Healthy (No.)	Infested (No.)	Infestation (%)	Infestation reduction over control	Healthy (g)	Infested (g)	Infestation (%)	Infestation reduction over control
T ₁	6.73 a	0.87 b	11.38 b	30.48	710.19 a	91.74 b	11.43 b	32.96
T ₂	7.13 a	0.40 d	5.31 d	67.56	743.79 a	55.30 f	6.93 f	59.35
T ₃	6.87 a	0.73 bc	9.62 bc	41.23	733.64 a	67.45 de	8.43 de	50.56
T ₄	7.27 a	0.47 d	5.98 d	63.47	736.52 a	61.23 ef	7.69 ef	54.90
T ₅	6.80 a	0.67 c	8.92 c	45.51	726.03 a	70.10 cd	8.80 d	48.39
T ₆	6.73 a	0.67 c	8.98 c	45.14	713.52 a	76.96 c	9.74 c	42.87
T ₇	6.13 b	1.20 a	16.37 a	--	638.30 b	131.23 a	17.05 a	--
LSD _(0.05)	0.537	0.187	1.892	--	61.86	8.423	0.924	--
CV(%)	4.43	14.76	11.18	--	4.87	5.98	5.19	--

In a column, numeric data represents the mean value of 3 replications; each replication is derived from 5 selected vine/plot of each treatment

In a column means having similar letter(s) are statistically identical and those having dissimilar letter(s) differ significantly as per 0.05 level of probability

T₁: Emamectin benzoate @ 2 ml/L of water

T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing

T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application)

T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval (3 times application)

T₅: Voliam flexi 300SC @ 1 ml/L at 15 days interval (3 times application)

T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application)

T₇: Untreated control

infestation level of storage roots in weight basis, the lowest infestation was recorded from T₂ (6.93%) treatment which statistically similar to T₄ (7.69%) treatment, whereas the highest infestation (17.05%) was observed from T₇ which was closely followed by T₁ (11.43%) treatment. In case of reduction over control the highest value was recorded from T₂ (59.35) and the lowest value was found from T₁ (32.96) treatment. According to Stathers *et al.*, (2005) reported that timely harvesting to remove the largest storage root most at risk from weevil attack and subsequent hilling up of the soil around the roots prevent weevils from accessing the root through cracks in the soil.

4.4 Yield contributing characters and yield

Yield contributing characters and yield of sweet potato showed statistically significant variation due to different treatment (Table 9).

4.4.1 Vine length at harvest

In consideration of vine length at harvest, the longest vine was recorded from T₂ (205.54 cm) treatment, which was statistically similar with other treatment except T₇, whereas the shortest vine from T₇ (169.56 cm) treatment (Table 9).

4.4.2 Number of storage roots/vine

For number of storage roots/vine, the highest number was recorded from T₂ (7.00) treatment, which was statistically similar with other treatment except T₇ and the minimum number was found from T₇ (5.93) treatment (Table 9).

4.4.3 Weight of individual storage root

In case of weight of individual storage root, the highest weight was recorded from T₂ (148.48 g), which was statistically similar with other treatment except T₇, while the lowest weight was found from T₇ (127.01 g) treatment (Table 9).

4.4.4 Yield of storage roots

In consideration of yield of storage roots, the highest yield was recorded from T₂ (41.86 t/ha), which was statistically similar with other treatment except T₇ and the lowest yield was found from T₇ (30.27 t/ha) treatment (Table 9).

Table 9. Yield and yield contributing characters of sweet potato for different management practices

Treatments	Vine length (cm) at harvest	Number of storage roots/vine	Weight of individual storage root (g)	Yield of storage roots (t/ha)
T ₁	191.80 a	6.56 a	141.00 a	38.54 a
T ₂	205.54 a	7.00 a	148.48 a	41.86 a
T ₃	199.25 a	6.69 a	146.21 a	39.84 a
T ₄	202.01 a	6.91 a	147.01 a	40.14 a
T ₅	197.80 a	6.58 a	144.96 a	39.66 a
T ₆	194.85 a	6.56 a	142.26 a	39.68 a
T ₇	169.56 b	5.93 b	127.01 b	30.27 b
LSD _(0.05)	19.57	0.439	12.52	3.907
CV(%)	5.66	3.73	4.94	5.69

In a column, numeric data represents the mean value of 3 replications; each replication is derived from 5 selected vine/plot of each treatment

In a column means having similar letter(s) are statistically identical and those having dissimilar letter(s) differ significantly as per 0.05 level of probability

T₁: Emamectin benzoate @ 2 ml/L of water

T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing

T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application)

T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval (3 times application)

T₅: Voliam flexi 300SC @ 1 ml/L at 15 days interval (3 times application)

T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application)

T₇: Untreated control

CHAPTER V

SUMMARY AND CONCLUSION

The present study regarding documentation of insect pests in sweet potato and their management has been conducted during rabi season (November 2016 to March 2017) in the experimental fields of Sher-e-Bangla Agricultural University, Dhaka. The variety of BARI Sweet Potato 8 were used as test crop. Seven treatments with one untreated control were tested in this experiment. They were T₁: Emamectin benzoate @ 2 ml/L of water; T₂: Carbofuran 5G @ 15 kg/ha at 60 days after sowing; T₃: Diazinon 10G @ 12 kg/ha at 60 days after sowing + Azadiractin @ 3 ml/L at 10 days interval (3 times application); T₄: Sevin 85WP @ 3 g/L + Admire @ 1 ml/l at 15 days interval (3 times application); T₅: Voliam flexi 300SC @ 1 ml/L at 15 days interval (3 times application); T₆: Lamda cyhalothin @ 1 ml/L at 10 days interval (3 times application) and T₇: Untreated control. The experiment was laid out in Randomized Completely Block Design (RCBD) with three replications. Data were recorded on insect pest incidence, vine and branch infestation, infestation of storage roots in number and weight basis and statistically significant variation was recorded for different treatment.

Under the present study 9 different species of insect pests were found and they were belongs to 9 family under 3 orders. Among the recorded insect pests 5 species belong to the order Hemiptera, 3 species belong to Coleoptera and only 1 species under Orthoptera order and there is different family for each of the insect pests species. Among the insect pests it was observed that sweet potato weevil was more destructive for storage roots of sweet potato. Data revealed that for the consideration of sweet potato bug, the lowest number of sweet potato bug was observed from T₂ (1.27), whereas the highest number from T₇ (5.13). In case of mealybug, the lowest number of mealybug was recorded from T₂ (1.67), whereas the highest from T₇ (6.93) treatment. In consideration of green vegetable bug, the lowest number of green vegetable bug was recorded from T₂ (1.07), while the

highest number from T₇ (4.20) treatment. In case of leaf hopper, the lowest number of leaf hopper was recorded from T₂ (2.07) treatment, whereas the highest number from T₇ (6.40) treatment. Data revealed that for minor leaf feeders, the lowest number of minor leaf feeders was recorded from T₂ (1.13), whereas the highest number from T₇ (4.73) treatment. In consideration of sweet potato beetle, the lowest number of sweet potato beetle was recorded from T₂ (1.47), whereas the highest number from T₇ (6.33) treatment. In case of whitefly, the lowest number of whitefly was recorded from T₂ (1.67), whereas the highest number from T₇ (6.73) treatment. From the recorded data in case of tortoiseshell beetle, the lowest number of tortoiseshell beetle was found from T₂ (1.13), whereas the highest number from T₇ (4.33) treatment. Data revealed that for sweet potato weevil, the lowest number of sweet potato weevil was observed from T₂ (1.67), while the highest number from T₇ (6.73) treatment.

In case of leaves of sweet potato at early growth stage, it was observed that the lowest infestation was recorded from T₂ (3.90%), whereas the highest infestation (12.30%) was observed from T₇ treatment. In case of leaf infestation reduction over control the highest value was recorded from T₂ (68.29) and the lowest value from T₁ (31.79) treatment. At mid growth stage, the lowest infestation was recorded from T₂ (2.71%), whereas the highest infestation (10.07%) was observed from T₇ treatment. In case of infestation reduction over control the highest value was recorded from T₂ (73.09) and the lowest value was found from T₁ (44.29) treatment. At late growth stage, the lowest infestation was recorded from T₂ (2.58%), whereas the highest infestation (9.09%) was observed from T₇ treatment. In case of reduction over control the highest value was recorded from T₂ (71.62) and the lowest value was found from T₁ (45.87) treatment.

For branches/vine of sweet potato at early growth stage, the lowest infestation was recorded from T₂ (2.39%), whereas the highest infestation (12.86%) from T₇ treatment. In case of infestation reduction over control, the highest value was recorded from T₂ (81.42) and the lowest value was found from T₁ (36.94) treatment. At mid growth stage, the lowest infestation was recorded from T₂

(2.45%), whereas the highest infestation (11.74%) from T₇ treatment. In case of reduction over control, the highest value was recorded from T₂ (79.13) and the lowest value was found from T₁ (34.58) treatment. At late growth stage, the lowest infestation was recorded from T₂ (2.61%), whereas the highest infestation (11.09%) from T₇ treatment. In case of reduction over control, the highest value was recorded from T₂ (76.47) and the lowest value from T₁ (28.49) treatment.

In case of storage roots of sweet potato/vine in number basis at 15 days early harvest, the lowest infestation was recorded from T₂ (3.69%), whereas the highest infestation (12.25%) was observed from T₇ treatment. In case of reduction over control the highest value was recorded from T₂ (69.88) and the lowest value was found from T₁ (29.22) treatment. At optimum days harvest, it was observed that the lowest infestation was recorded from T₂ (4.49%), whereas the highest infestation (13.56%) from T₇ treatment. In case of reduction over control the highest value was recorded from T₂ (66.89) and the lowest value from T₁ (26.47) treatment. After 15 days from optimum harvest, the lowest infestation was recorded from T₂ (5.31%), whereas the highest infestation (16.37%) from T₇ treatment. In case of reduction over control the highest value was recorded from T₂ (67.56) and the lowest value from T₁ (30.48) treatment.

In case of storage roots of sweet potato/vine in weight basis at 15 days early harvest, it was observed that the lowest infestation was recorded from T₂ (5.68%), whereas the highest infestation (12.98%) was observed from T₇ treatment. In case of reduction over control the highest value was recorded from T₂ (56.24) and the lowest value was found from T₁ (28.43) treatment. At optimum days harvest, the lowest infestation was recorded from T₂ (6.49%), whereas the highest infestation (14.16%) from T₇ treatment. In case of reduction over control the highest value was recorded from T₂ (54.17) and the lowest value was found from T₁ (30.01) treatment. After 15 days from optimum harvest, the lowest infestation was from T₂ (6.93%), whereas the highest infestation (17.05%) from T₇ treatment. In case of reduction over control the highest value was recorded from T₂ (59.35) and the lowest from T₁ (32.96) treatment.

In consideration of vine length at harvest, the longest vine was recorded from T₂ (205.54 cm), whereas the shortest vine from T₇ (169.56 cm) treatment. For number of storage roots/vine, the highest number was recorded from T₂ (7.00), while the minimum number from T₇ (5.93) treatment. In case of weight of individual storage root, the highest weight was recorded from T₂ (148.48 g), whereas the lowest from T₇ (127.01 g) treatment. For yield of storage roots, the highest yield was recorded from T₂ (41.86 t/ha), whereas the lowest from T₇ (30.27 t/ha) treatment.

Conclusion:

From the above discussion it can be concluded that among the different control measures, Carbofuran 5G @ 15 kg/ha at 60 days after sowing was the best for the controlling insect pests of sweet potato and also attaining highest yield of storage roots.

Recommendation:

Due to some limitations a few number of control measures were used for the controlling insect pests of sweet potato. So, more bio-pesticides and chemical pesticides can be included for the further studies to find out the more profitable yield of sweet potato storage roots. However, further study of this experiment is also needed in different agro-ecological zones of Bangladesh for accuracy of the results obtained from the present experiment.

REFERENCES

- Abidin, P.E. (2004). Sweetpotato breeding for northeastern Uganda: farmer varieties, farmer-participatory selection, and stability of performance., Dissertation, Wageningen University.
- Adam, S. (1992). Sweet potato can ward off weeds. *Agricultural Research* November, 22.
- Ahmed, M.T., Nath, S.C., Sorwar, M.A. and Rashid, M.H. (2015). Cost-effectiveness and resource use efficiency of sweet potato in Bangladesh. *J. of Agril. Economics and Rural Development*, volume 2(2), pp. 26-31.
- Allard, G. B. (1990). Integrated control of arthropod pests of root crops November 1988 -December 1989. Midterm report. Nairobi, Kenya. 22 pp.
- Allard, G. B., Cock, M. J. W. and Rangi, D. K. (1991). Integrated control of arthropod pests of root crops, final report. Nairobi, Kenya. 18 pp.
- Allard, G. B., Cock, M. J. W. and Rangi, D. K. (1991). Integrated control of arthropod pests of root crops, final report. Nairobi, Kenya. 18 pp.
- Ames, T., Smit, N. E. J. M., Braun, A. R., O'Sullivan J. N. and Skoglund, L. G. 1996. Sweet potato: Major pests' diseases, and nutritional disorders. International Potato Center (CIP), Lima, Peru.
- Ames, T., Smit, N. E. J. M., Braun, S. R. O., Sullivan, J. N. and Skoglund, L. G. (1996). Sweet potato. Major Pest, Disease and Nutritional Disorders. International potato center (CIP), Lima, Peru. 152 pp.
- Andrade, M., Barker, I., Cole, D., Dapaah H, Elliott, H., Fuentes, S., Grüneberg, W., Kapinga, R., Kroschel, J., Labarta, R., Lemaga,B., Loechl, C., Low, J., Lynam, J., Mwanga,R., Ortiz, O., Oswald, A. and Thiele, G. (2009). Unleashing the potential of sweetpotato in sub-Saharan Africa: current challenges and way forward. International Potato Center (CIP), Lima, Peru.
- Anonymous. (1989). *Textile Outlook International*. The Economist Intelligence Unit. 40, Duke Street, London, U.K. pp. 3-4.
- Anota, Y. and Odebisi, J. A. (1984). Resistance in sweet potato to *Cylas puncticollis*. Boheman (Coleoptera: Curculionidae). *J. of Biologia Africana* 1(1): 21 – 30.
- AVRDC. (1987). Correlation of root characteristics and root infestation by sweet potato weevil. Progress report. Taipei, Taiwan. 13pp.

- Azerad, R. (2000). Regio and stereoselective microbial hydroxylation of terpenoid compounds. In: Patel, R.N. (Eds.), *Stereoselective Biocatalysis*. Marcel Dekker, New York, pp. 153–180.
- Bashaasha, B., Mwanga, R.O.M., Ocitti p'Obwoya, C. and Ewell, P.T. (1995). Sweetpotato in farming and food systems of Uganda. A farm survey report., Sub-Saharan African region, Nairobi, Kenya/National Agricultural Research Organization, Kampala, Uganda.
- BBS. (2012). Statistical Year Book of Bangladesh. Bangladesh Bureau of Statistics Division, Ministry of Planning, Government of the People Republic of Bangladesh, Dhaka, Bangladesh.
- Bhuiyan, M.K.R., Hossain, M., Hussain, M.A.E. and Attaluri, S. (2006). Research and development activities of sweet potato in Bangladesh. In: 14th Triennial Symposium of International Society of Tropical Root Crops, 20-26 November 2006, Central Tuber Crops Research Institute, Thiruvananthapuram, India, 17pp.
- Biswal, S. (2008). Response of sweet potato (*Ipomoea batatas* L.) to irrigation and fertility levels. PhD thesis, Orissa Agricultural University and Technology, Bhubaneswar, India.
- Bohlen, E. 1973. Crop pests in Tanzania and their control. Verlag Paul Parey. Berlin, Germany.
- Buchanan, G. O., Williams, L. A. D. and Reese, P. B. (2000). Biotransformation of cadinane sesquiterpenes by *Beauveria bassiana* ATCC 7159. *J. of Phytochemistry* **54**: 39-45. Labo Lima, M. L. S. (1990). Bioassays with *Metarhizium anisopliae* and *Beauveria bassiana* against adults of the sweet potato weevil *Cylas puncticollis* in Investiga Cao Agraria, Cape Verde. *J. of insect* **3**(2): 50 – 52.
- Burri, B.J. (2011). Evaluating sweetpotato as an intervention food to prevent vitamin A deficiency. *Compr Rev Food Sci Food Saf* **10**:118–130.
- Chalfant, R, B., Jansson, R. K., Seal, D. R. and Schalk, J. M. (1990). Ecology and management of sweet potato insects. *Annual Review of Entomol.* **35**:157 – 189.
- Chen, K. and Wey, H. (1990). Dissolution enzyme kinetics of 11-hydroxylation of cortexolone by *Curvularia lunata*. *J. of Enzyme Microbiological Tech.* **12**: 616 -621.

- Cheng, K. W. (1981). Screening for varietal resistant to sweet potato weevil. Scientific Research report in Republic of China Part I 1980. Nankang, Taiwan. 32pp.
- Chua, L.K. and Kays, S.J. (1981). Effect of soil oxygen concentration on sweet potato storage root induction and or development. *Hort Science* **16**: 17-73.
- Collins, D. O., Buchanan, G. O., Reynolds, W. F. and Reese, P. B. (2001). Biotransformation of squamulosone by *Curvularia lunata* ATCC 12017. *J. of Phytochemistry* **57**: 377–383.
- Daiber, K. C. (1994). Injurious insects, spider mites and nematodes on sweet potatoes in Southern Africa. *J. of Insect* **101**(5): 550 – 557.
- Davies, H. G., Green, R.H., Kelly, D. R. and Roberts, S. M., (1989). Biotransformations in preparative organic chemistry: the use of isolated enzymes and whole cell system in synthesis. Academic Press, London. 145 pp.
- Downham, M. C. A., Smit, N. E. J. M., Laboke, P. O., Hall, D. R. and Odongo, B. (2001). Reduction of Pre-harvest infestations of Africa sweet potato weevils *Cylas brunneus* and *Cylas puncticollis* (Coleoptera: Apionidae) using a pheromone mating disruption technique. *J. of Crop Prot.* **20**:163 – 166.
- Ebregt, E., Struik, P. C., Abidin, P. E. and Odongo, B. (2005). Pest damage in sweet potato, groundnut and maize in North – Eastern Uganda with special reference to damage by millipedes (Diplopoda). NJAS – Wageningen. *J. of Life Scis* **53**:49 – 69.
- Edris, K.M., Islam, D.K., Chowdhury, M.S. and Haque, A.K.M.M. (1979). Detailed Soil Survey of Bangladesh, Dept. Soil Survey, Govt. People's Republic of Bangladesh. pp. 118.
- FAO (Food and Agriculture Organization) (2011). Production quantity of primary crops, country statistics for eastern Africa. <http://faostat3.fao.org/faostatgateway/go/to/download/Q/QC/E>. Accessed 29 April 2013.
- FAO. (1988). Production Year Book. Food and Agricultural of the United Nations Rome, Italy. 42: 190-193.
- FAO. (2012): Food and Agricultural Organization. <http://faostat.fao.org/site/567/DesktopDefault.aspx?PageID=567#ancor>.
- FAOSTAT. (2011). Food and Agricultural Organization <http://faostat.fao.org/site/567/DesktopDefault.aspx?PageID=567#ancor>.

- Geisthardt, M. and Van Hasten, A. (1992). Noxious beetles of the Cape verde Islands with a additional reference to West Africa report. Wiesbaden Germany: Verlagchrista Hemmen. 198 pp.
- Gomes, F. and Carr, M.K.V. (2003). Effects of water availability and vine harvesting frequency on the productivity of sweet potato in southern Mozambique. II. Crop water use. *Experimental Agriculture* 39, 39-54.
- Goswami, S.B., Sen, H. and Jama, P.K. (1995). Rootization and yield potential of sweet potato cultivars as influenced by water management practices. *J. of Root Crops* 21 (2), 77-81.
- Hahn, S. K. and Leuschner, K. (1981). Resistance of sweet potato cultivars to African sweet potato weevil. *J. of Crop Sci.* 2(4): 499 – 503.
- Haider, J., Marutomo, K. and Azad, A. K. (1991). Estimation of microbial biomass carbon and nitrogen in Bangladesh. *Soil Sci. Plant Nutr.* 37(4): 591-599.
- Harrison, H. F. Jr., Peterson, J. K., Snook, M. E., Bohac, J. R. and Jackson, D. M. (2003). Quantity and potential biological activity of caffeic acid in sweet potato (*Ipomoea batatas* L.) storage root periderm. *J. of Agri. Food and Chemistry* 51: 2943-2948.
- Harrison, H.F. Jr. and Peterson, J.K. (1991). Evidence that sweet potato (*Ipomoea batatas*) is allelopathic to yellow nutsedge (*Cyperus esculentus*). *Weed Sci.* 39, 308-312.
- Hill, D. 1975. *Agricultural Insect Pests of the Tropics and their control.* Cambridge University Press, London, UK.
- Holland, H. L. (1992). *Organic synthesis with oxidative enzymes.* VCH, New York. 151 pp.
- Holland, H. L. and Reimland, E. (1985). Microbial hydroxylation of steroids. Rearrangement during epoxidation and hydroxylation, and the stepwise nature of these enzyme reactions. *Canadian J. of Chemistry* 63: 1121–1126.
- Hwang, J. S. and Hung, C. C. (1991). Evaluation of the effect of integrated control of sweet potato weevil, *Cylas formicarius* (Fabricius) with sex pheromone and insecticide. *Chinese J. of Entomol.* 11: 140-146.
- Indira, P. and Kabeerathumma, S. (1988). Physiological response of sweet potato under water stress. I. Effect of water stress during the different phases of tuberization. *J. of Root Crops* 14 (2), 37-40.

- Jaarsveld, P.J., Faber, M., Tanumihardjo, S.A., Nestel, P., Lombard, C.J. and Benadé, A.J.S. (2005). β -carotene-rich orange-fleshed sweet potato improves the vitamin A status of primary school children assessed with the modified-relative-dose-response test^{1,2,3}. *Am J Clin Nutr* **81**:1080–1087.
- Jackson, D. M. and Peterson, J. K. (2000). Sublethal effects of resin glycosides from the periderm of sweet potato storage roots on *Plutella xylostella* (Lepidoptera: Plutellidae). *J. of Economic Entomol.* **93**: 388-393.
- Jalal, F., Nesheim, M.C., Agus, Z., Sanjur, D. and Habicht, J.P. (1998). Serum retinol concentrations in children are affected by food sources of β -carotene, fat intake, and anthelmintic drug treatment. *Am J Clin Nutr* **68**:623–629.
- Jansson, R. K. and Lecrone, S. H. (1991). *Euderus purpureas* (Hymenoptera: Eulophidae) a parasitoid of sweet potato weevil (Coleoptera: Apionidae) *J. of Southern Florida Entomol.* **74**: 596-598.
- Jansson, R. K. and Lecrone, S. H. (1997). Effect of sweet potato cultivar on efficacy and persistence of entomopathogenic nematodes for control of *Cylas formicarius*. *Nematropica. J. of Nematode* **27**: 41-52.
- Jansson, R. K., Lecrone, S. H. and Gaugler, R. (1991). Comparison of single and multiple releases of *Heterorhabditis bacteriophora* Poinar (Nematoda: Heterorhabditidae) for control of *Cylas formicarius* (Fabricius) (Coleoptera: Apionidae). *J. of Biol. Control* **1**: 320-328.
- Jeong, B.C. (2000). Current role of sweet potato and its earlier harvesting by improved PE film mulch cultivation in Korea. In: Potential of Root Crops for Food and Industrial Resources, Proceedings of 12th Symposium of International Society of Tropical Root Crops, September 10-16, 2000. Tsukuba, Japan, pp 350.
- Kabi, S., Ocenga – Latigo, M. W., Smit, N. E. J. M., Stathers, T. E. and Rees, D. (2001). Influence of Sweetpotato Rooting characteristic on infestation and damage by *Cylas* spp. *J. of African Crop Sci.* **9**:165 – 174.
- Kaguongo, W., Ortmann, G., Wale, E., Darroch, M. and Low, J. (2012). Factors influencing adoption and intensity of adoption of orange flesh sweetpotato varieties: evidence from an extension intervention in Nyanza and western provinces, Kenya. *Afr J. Agric Res* **7**:493–503.
- Kasasian, L. (1978). Pest control in tropical roots crops. PANS manual no. 4. Ibadan, Nigeria 235pp.
- Kassasian, L. and Seeyave, J. (1969). Critical periods for weed competition. PANS 15, 208-212.

- Kays, S. J., Harrison, J. A., Wildon, D. D. and Severson, R. F. (1993). Semi-artificial Diet for the Sweet potato weevil. (Coleoptera; Curculionidae). *J. of Econ. Entomol.* 86:957-961.
- Kibata, G. N. 1973. Studies on varietal susceptibility and pest control of sweet potato (*Ipomoea batatas*) in Central Kenya. Advances in Medical Veterinary and Agricultural entomology in eastern Africa, Nairobi, Kenya.
- Komi, K. (2000). Eradication of sweet potato weevil, *Cylas formicarius* (Fabricius) from Muroto City, Kochi. Japan Extension Bulletin **493**: 15-22.
- Lagnaoui, A., Cisneros, F., Alcazar, J. and Morales, F. (2000). A sustainable pest management strategy for sweet potato weevil in Cuba: a success story. South American Extension Bulletin **493**: 1-7.
- Lamare, V., Fourneron, J. D. and Furstoss, R. (1991). Microbial transformations 9. Biohydroxylation of α -cedrene and cedrol: synthesis of an odoriferous minor component of Cedar wood essential oil. *J. of Biochemistry* **28**: 6269-6272.
- Li, Z. Z. (1998). Control of *Cylas formicarius* (Fabricius) with pheromone. *J. of Plant Prot.* **24**: 28-30.
- Low, J.W., Arimond, M., Osman, N., Cunguara, B., Zano, F. and Tschirley, D. (2007). A foodbased approach introducing orange-fleshed sweetpotatoes increased vitamin A intake and serum retinol concentrations in young children in rural Mozambique. *J Nutr* **137**:1320–1327.
- Lunaa, F. and Wilson, P.N. (2015). An Economic Exploration of Smallholder Value Chains: Coffee Transactions in Chiapas, Mexico, *International Food and Agribusiness Management Review*, **18**(3).
- Macfarlane, R. (1987). Sweet potato weevil (*Cylas formicarius*) insecticide trial Annual Report 1985. Ministry of Agriculture and Lands, Research Department, Solomon Islands. 36pp.
- Maeto, K. and Uesato, T. (2007). A new species of Bracon (Hymenoptera: Braconidae) parasitic on alien sweet potato weevils in the south-west islands of Japanese. *Japanese J. of Entomol. Sci.* **10**: 55-63.
- Mannion, C. M. and Jansson, R. K. (1992). Comparison of ten entomopathogenic nematodes for control of sweet potato weevil (Coleoptera: Apionidae). *J. of Econ. Entomol.* **85**: 1642-1650.
- Mao, L., Jett, L. E., Story, R. N., Hammond, A. M., Peterson, J. K. and Labonte, D. R. (2004). Influence of drought stress on sweet potato resistance to sweet potato weevil, *Cylas formicarius* (Coleoptera: Apionidae), and storage root chemistry. *Florida J. of Entomol.* **87**: 261-267.

- Marti, H. R., Mills, H. A., Severson, R. F. and Kays, S. J. (1993). Nutritional effects on sweet potato storage root surface chemistry. *J. of Plant Nutrient* **16**: 653-665.
- Martin, F.W. (1988). Planting dates and morphology and growth of four sweet potato clones in Puerto Rico. *J. of Agril. University Puerto Rico* **72** (3), 437-444.
- Mason, L. J. and Jansson, R. K. (1991). Disruption of sex pheromone communication in *Cylas formicarius* (Coleoptera: Apionidae) as a potential means of control. *Florida J. of Entomol.* **74**: 469-472.
- Mason, L. J., Seal, D. R. and Jansson, R. K. (1991). Response of sweet potato weevil (Coleoptera: Apionidae) to selected insecticides. *Florida J. of Entomol.* **74**: 350-355.
- Ministry of Agriculture. 1992a. Comparative Estimate of All-Island Domestic Crop Production and Area Reaped 1991/1990. Data Bank & Evaluation Division, Jamaica, 32 pp.
- Misra, A. K., Singh, R. S. and Pandey, S. K. (2001). Relative efficacy of chemicals and botanical insecticide against sweet potato weevil, *Cylas formicarius* (Fabricius). *Annual J. of Plant Prot. Sci.* **9**:201-204.
- Mutuura, J., Ewell, P.T., Abubaker, A., Munga, T., Ajanga, S., Irungu, J., Omari, F. and Maobe, S. (1992). Sweet potato in the food systems of Kenya. Results of a socio-economic survey. In: Proceedings of KARI/CIP technical workshop on collaborative research, Nairobi.
- Mwanga, R.O.M., Niringiye, C., Odongo, B., Opio, A.F., Lemaga, B., Kyaddondo, B., Sserunjogi, L., Owori, C. and Kapinga, R. (2003a). Partnership: an approach to promote orange-fleshed sweetpotato, experiences from Uganda. In: Kapinga, R., Kingamkono, R., Msabaha, M., Ndunguru, J., Lemaga, B. and Tusiime, G. (eds). Proceedings of the 13th international society for tropical root crops (ISTRC) symposium. Tropical root and tuber crops: opportunities for poverty alleviation and sustainable livelihoods in the developing world. November 10–14, 2003, Arusha., pp 770–779.
- Nair, O. M. (2006). Agro- techniques and planting material production in sweet potato. In: Quality planting material production in Tropical tuber crops, Central Tuber Crop Research Institute. (Edited by Byju, G.) Thiruvananthapurain, India. pp 55-58.
- Nath, R., Kundu, C.K., Majumder, A. Gunri, S. Chattopadhyay, A. and Sen, H. (2006). Productivity of sweet potato as influenced by cultivar, season and staggered harvesting in laterite ecosystem of West Bengal. In: 14th Triennial Symposium of International Society of Tropical Root Crops, 20-26 November 2006, Central Tuber Crops Research Institute, Thiruvananthapuram, India, pp 213.

- Nawale, R.N. and Salvi, M.J. (1983). Effects of season on yield of sweet potato. *J. of Root Crops* **9**, 55-58.
- Nderitu, J., Sila, M., Nyamasyo, G. and Kasina, M. (2009). Insect species associated with sweet potatoes in Eastern Kenya. *International J. Sustainable Crop Production* **4**:14 -18.
- Nedunchezhiyan, M. and Byju, G. (2005). Effect of planting season on growth and yield of sweet potato (*Ipomoea batatas* L.) varieties. *J. of Root Crops* (**2**), 111-114.
- Nedunchezhiyan, M. and Ray, R.C. (2010). Sweet potato growth, development production and utilization: overview. In: Ray RC, Tomlins KI (Eds) Sweet Potato: Post Harvest Aspects in Food, Nova Science Publishers Inc., New York, pp 1-26.
- Nedunchezhiyan, M. and Reddy, D.S. (2006). Effect of time of planting and varieties on growth and yield of sweet potato (*Ipomoea batatas* L.) under rainfed conditions. In: Naskar SK, Nedunchezhiyan M, Rajasekhara Rao K, Sivakumar PS, Ray RC, Misra RS, Mukherjee A (Eds) Root and Tuber Crops: in Nutrition, Food Security and Sustainable Environment, Regional Centre of Central Tuber Crops Research Institute, Bhubaneswar, India, pp 123-127.
- Ngoan, T.N. (2006). Status of root crops production, utilization and marketing in Vietnam. In: 14th Triennial Symposium of International Society of Tropical Root Crops, 20-26 November 2006, Central Tuber Crops Research Institute, Thiruvananthapuram, India, 9 pp.
- Nottingham, S. F., Son, K. C., Wilson, D. D., Severson, R. F. and Kays, S. J. (1989). Feeding and oviposition preferences of sweet potato weevil, *Cylas formicarius elegantulus* (Summers) on storage roots of sweet potato cultivars with differing surface chemistries. *J. of Chemistry and Ecol.* **15**: 895-903.
- Nottingham, S. F., Wilson, D. D., Severson, R. F. and Kays, S. J. (1987). Feeding and oviposition preferences of the sweet potato weevil, *Cylas formicarius elegantulus* on the outer periderm and exposed inner core of storage roots of selected sweet potato cultivars. *J. of Entomol.* **45**: 271-275.
- Okonyo, N. J. (2013). Effect of variety, planting material and in-ground storage on sweet potato weevil (*Cylas* spp.) population, damage and yield of sweet potato (*Ipomea batatas* Lam.). Dissertation for Award of MSc Degree at Egerton University, Kenya. 98pp.
- Otto, N., Russel, M. and Eric, C. (2006). Sweet potato weevil. *A review of recent management advances and appraisal of previous research in Papua. New Guinea and Australia.* 39 pp.

- Palaniswami, M. S. and Mohandas, N. (1994). Reridging as a cultural method for the management of sweet potato weevil *Cylas formicarius* (Fabricius). *J. of Root Crops* **20**: 101-105.
- Palaniswami, M. S. and Mohandas, N. (1996). Effect of soil drenching with insecticides at different stages of sweet potato on the control of *Cylas formicarius* and persistent toxicity of insecticides to adult weevil. In: Tropical tuber crops: Problems, Prospects and future strategies. (Edited by Kurup, G.T. *et al.*), New York, USA. pp. 340-346.
- Palaniswami, M. S. and Rajamma, P. (1986). Biological control of sweet potato weevil and cassava spider mites. Annual progress report 1986 for the period January-December 1986. Central Tuber Crops Research Institute, Trivandrum, India. 102pp.
- Palaniswami, M. S., Visalakshi, A., Mohandas, N. and Das, L. (2002). Evaluation of soil application method of insecticides against *Cylas formicarius* (Fabricius) and its impact on soil microflora in sweet potato ecosystem. *J. of Root Crops* **28**: 55-60.
- Panizzi, A. R. and Slansky, Jr. F. 1985. Legume host impact on performance of adult *Piezodorus guilinii* (Westwood) (Hemiptera: Pentatomidae). *Environ. Entomol.* **14**: 237-242.
- Parker, B. L. (1990). Report on the distribution of *Cylas formicarius* Fabricius (Coleoptera: Apionidae) in Africa. Kenya. 49 pp.
- Parker, B. L., Wolfe, G. W. and Gitonga, W. (1990). Report on the distribution of *Cylas formicarius* Fabricius (Coleoptera: Apionidae) in Africa. South Africa. 63pp.
- Petro, R. de., Lightfoot, C., Apura, D., Acaba, M. and Cabling, J. (1986). Screening of sweet potato varieties by subsistence farmers in, Philippines: a case of traditional experimentation in upland agriculture. *J. of Tropical Research* **8**(4): 201 – 207.
- Pfeiffer, H. J. (1982). Sweet potato improvement in Cameroon. Root crops in eastern Africa. Proceedings of root crops workshop, Kigali, Rwanda, 23-27 November 1980. 33-38 pp.
- Pillai, K. S., Palaniswami, M. S., Rajamma, P., Ravindran, C. S. and Premkumar, T. (1996). An IPM approach for sweet potato weevil. In: Tropical tuber crops: Problems, Prospects and future strategies. (Edited by Kurup, G.T. *et al.*) New York, USA. p. 329-339.
- Powell, K. S., Hatlemink, A. E., Eganae, J. F., Walo, C. and Polomas, S. (2001). Sweet potato weevil (*Cylas formicarius*) incidence in the Humid Lowland of PNG. In : Proceedings of food security in Papua New Guinea. (Edited by Bourke, R. M. *et.al.*), 6 – 12 July 1998. 736 – 745pp.

- Rajamma, P. (1990). Evaluation of some newer insecticides for the control of the sweet potato weevil, *Cylas formicarius* (Fabricius). *J. of Entomol.* **15**: 241-243.
- Rajamma, P. and Pillai, K. S. (1991). Efficacy of insecticides and pruning on the control of sweet potato weevil. *J. of Root Crops* **17**: 174-179.
- Rajasekhara Rao, K. (2002). Induced host plant resistance in the management of sucking insect pests of groundnut. *J. of Plant Prot.* **10**: 45-50.
- Rajasekhara Rao, K. (2005). Systems approach for management of insect pest problem in tuber crops by farmers of Meghalaya. *J. of Plant Prot.* **22**: 34.
- Rajasekhara Rao, K., Naskar, S. K., Misra, R. S., Mukherjee, A., Thakur, N. S. A. and Yadav, D. S. (2006). Distribution of Major insect pests of Tuber crops and their natural enemies in North Eastern Hill Region of India. In: Annual Report 2005-2006, Central Tuber Crops Research Institute. (Edited by Premkumar, T. *et al.*), Thiruvananthapuram, Kerala, India. pp. 106-108.
- Ravindran, C.S. and Mohankumar, C.R. (1985). Effect of method of land preparation and depth of planting on the yield of sweet potato. In: Proceedings of National Symposium on Production and Utilization of Tropical Tuber Crops, 1 November 27-28, Central Tuber Crops Research Institute, Trivandrum, India, pp 129-133.
- Ravindran, C.S., Ravi, V., Nedunchezhiyan, M., George, J. and Naskar, S.K. (2010). Weed management in tropical tuber crops: An overview. *J. of Root Crops* **36** (2), 119-131.
- Sajjapongse, A. (1989). The effect of climatic factors on the yield of sweet potato. IBSRAM Newsletter **5** (13), 7.
- Sato, K., Uritani, I. and Saito, T. (1981). Characterization of the terplue inducing factor isolated from the larva of the sweet potato weevil, *Cylas formicarius* Fabricius (Coleoptera: Brentidae). *J. of Appl. Entomol. and Zool.* **16** (2): 103 – 112.
- Schmutterer, H. (1969). Report of Pests of crops in Northeast and Central Africa with particular reference to Sudan. Stuttgart, Germany. 45 pp.
- Shakoor, A., Kiarie, A.W., Njunguna, G. M. and Mihiu, S. G. (1984). Breeding for early maturity, drought, disease and insect resistance in sweet potato. *East African Agricultural and Forestry J.* **44**:318 – 323.
- Singh, B., Yazdani, S. S. and Singh, R. (1993). Relationship between biochemical constituents of sweet potato cultivars and resistance to weevil *Cylas formicarius* (Fabricius) damage. *J. of Entomological Research* **17**: 283-288.

- Sinha, A. K. (1994). Studies on comparative residual toxicity of insecticides to sweet potato weevil. Central Agricultural Station, Mon Ropos, Guyana (South America). *Indian J. of Entomol.* **56**: 123-128.
- Smit, N. E. J. M. (1997a). Integrated Pest Management for sweet potato in Eastern Africa. Thesis for Award of PhD at Wageningen University, Wageningen. 151pp.
- Smit, N. E. J. M. and Matengo, L. O. (1995). Farmers' cultural practices and their effects on pest control in sweet potato in South Nyanza, Kenya. *International J. of Pest Management* **41**(1): 2 – 21.
- Stathers, T. E., Rees, D., Nyango, A., Kiozya, H., Mbilinyi, L., Jeremiah, S., Kabi, S. and Smit, N. (2003). Sweet potato infestation by *Cylas* spp. in East Africa. *International J. of Pest Management* **49**(2); 141-146.
- Stathers, T., Namanda, S., Mwanga, R.O.M., Khisa, G. and Kapinga, R. (2005). Manual for sweet potato Integrated Production and Pest Management Farmer Field Schools in Sub Sahara Africa. International Potato center Kampala, Uganda. 168 pp.
- Stathers, T., Namanda, S., Mwanga, R.O.M., Khisa, G. and Kapinga, R. (2005). Manual for sweet potato Integrated Production and Pest Management Farmer Field Schools in Sub-Sahara Africa. International Potato center Kampala, Uganda. 168 pp.
- Stevenson, P. C., Muyinza, H., Hall, H., Porter, D. R., Farman, D., Talwana, H. and Mwanga, R.O. (2009). Chemical Basis for Resistance in sweetpotato (*Ipomoea batatas*) to the sweetpotato weevil (*Cylas puncticollis*). *J. of Pure Appl. Chemistry* **81**:141-151.
- Su, C. Y. (1991a). Field application of *Beauveria bassiana* for control of sweet potato weevil, *Cylas formicarius*. *Chinese J. of Entomol.* **11**:162-168.
- Su, C. Y. (1991b). Screening of soils pernicious to sweet potato weevil, *Cylas formicarius*, and use of *Beauveria bassiana*. *Chinese J. of Entomol.* **11**: 198-203.
- Talata, R.L., Mariscal, A.M. and Secreto, A.C. (1978). Critical periods for weed control in sweet potatoes. *Philippines J. of Weed Sci.* **5**, 1-6.
- Talekar, N. S. (1987). Influence of cultural pest management techniques on the infestation of sweet potato weevils. *International J. of Tropical Insect Sci.* **8**:809-814.
- Teli, V. S. and Salunkhe, G. N. (1994). Efficacy and economics of some insecticides against sweet potato weevil, *Cylas formicarius* (Fabricius). *J. of Insect Sci.* **7**: 202-204.

- Teli, V. S. and Salunkhe, G. N. (1996). A search for sources of resistance to sweet potato weevil. Morphological traits. *J. of Maharashtra Agril. Univ.* **20**: 400-403.
- Tesdale, J.R. and Mohler, C.L. (1993). Light transmittance, soil temperature and soil moisture under residue of hairy vetch and rye. *Agronomy J.* **85**: 673-680.
- Uritani, I., Saito, T., Honda, H. and Kim, W. K. (1975). Induction of furano – terperiods in sweet potato roots by larval components of the sweet potatoes weevils. *J. of Biochemistry* **37**:1857 – 1862.
- Vietmeyer, N. D. (1986). Lesser known plants of potential use in agriculture and forestry Science in Netherland. *J. of Forestry* **232**:1379 –1384.
- Wilson, D. D., Severson, R. F., Son, K. C. and Kays, S. J. (1988). Oviposition stimulant in sweet potato periderm for the sweet potato weevil, *Cylas formicarius elegantulus* (Coleoptera: Curculionidae). *J. of Environmental Entomol.* **17**: 691-693.
- Wolfe, G. W. (1991). The origin and dispersal of the pest species of *Cylas* with a key to the pest species groups of the world (Coleoptera: Apionidae). In: Sweet potato pest management, a global perspective (Edited by Jansson, R. K. and Raman, K.V.) West view Press, Boulder. 65pp.
- Yassen, A.A. and Thompson, P.G. (1988). The effect of cultivar plant population, density and irrigation on yield and quality of sweet potato. *HortScience* 23 (5), 825-828.
- Yasuda, K. (1995). Mass trapping of the sweet potato weevil *Cylas formicarius* (Fabricius) (Coleoptera: Brentidae) with a synthetic sex pheromone. *J. of Appl. Entomol. and Zool.* **30**: 31-36.
- Yasuda, K., Hongo, T. and Etoo, N. (2004). A new type of pheromone Formulation with visual stimulation for control of sweet potato weevil, *Cylas formicarius* (Fabricius). *Japanese J. of Appl. Entomol. and Zool.* **48**:87-93.
- Yasuda, K., Sugie, H. and Heath, R.R. (1992). Field evaluation of synthetic sex attractant pheromone of the sweet-potato weevil *Cylas formicarius* (Fabricius). (Coleoptera: Brentidae). *Japanese J. of Appl. Entomol. and Zool.* **36**: 81-87.
- Zaharah, A. and Tan, S.L. (2006). Performance of selected sweet potato varieties under different growing seasons on bris sandy soil in Malaysia. In: 14th Triennial Symposium of International Society of Tropical Root Crops, 20-26 November 2006, Central Tuber Crops Research Institute, Thiruvanantha puram, India, pp 225-226.

APPENDICES

Appendix I. Monthly record of air temperature, relative humidity and rainfall of the experimental site during the period from November, 2016 to March 2017

Month	Air temperature (⁰ C)		Relative humidity (%)	Rainfall (mm)
	Maximum	Minimum		
November, 2016	25.8	16.0	78	00
December, 2016	22.4	13.5	74	00
January, 2017	24.5	12.4	68	00
February, 2017	27.1	16.7	67	30
March, 2017	31.4	19.7	54	11

Source: Bangladesh Meteorological Department (Climate & weather division) Agargoan, Dhaka-1212

Appendix II. Analysis of variance of the data on number of identified insect pests in sweet potato during the growing period for different management practices

Source of variation	Degrees of freedom	Mean square								
		Number of different insect pests								
		Sweet potato bug	Mealybug	Green vegetable bug	Leaf hopper	Minor leaf feeders	Sweet potato beetle	Whitefly	Tortoiseshell beetle	Sweet potato weevil
Replication	2	0.017	0.013	0.013	0.048	0.013	0.025	0.025	0.017	0.023
Treatment	6	4.769**	8.693**	3.488**	6.377**	4.252**	7.642**	8.229**	3.310**	8.402**
Error	12	0.033	0.040	0.038	0.070	0.024	0.038	0.038	0.037	0.043

** : Significant at 0.01 level of significance

Appendix III. Analysis of variance of the data on incidence of insect pests on leaf and branches of sweet potato at early growth stage for different management practices

Source of variation	Degrees of freedom	Mean square					
		Leaf of sweet potato			Branches of sweet potato		
		Healthy (No.)	Infested (No.)	Infestation (%)	Healthy (No.)	Infested (No.)	Infestation (%)
Replication	2	0.310	0.008	0.007	0.002	0.002	0.890
Treatment	6	25.709*	4.060**	25.286**	0.500**	0.071**	35.585**
Error	12	6.930	0.105	0.889	0.073	0.006	2.337

** : Significant at 0.01 level of significance;

* : Significant at 0.05 level of significance

Appendix IV. Analysis of variance of the data on incidence of insect pests on leaf and branches of sweet potato at mid growth stage for different management practices

Source of variation	Degrees of freedom	Mean square					
		Leaf of sweet potato			Branches of sweet potato		
		Healthy (No.)	Infested (No.)	Infestation (%)	Healthy (No.)	Infested (No.)	Infestation (%)
Replication	2	2.749	0.0001	0.041	0.173	0.013	0.927
Treatment	6	66.048*	11.709**	18.487**	3.018**	0.371**	28.629**
Error	12	20.342	0.033	0.086	0.724	0.016	0.671

** : Significant at 0.01 level of significance;

* : Significant at 0.05 level of significance

Appendix V. Analysis of variance of the data on incidence of insect pests on leaf and branches of sweet potato at late growth stage for different management practices

Source of variation	Degrees of freedom	Mean square					
		Leaf of sweet potato			Branches of sweet potato		
		Healthy (No.)	Infested (No.)	Infestation (%)	Healthy (No.)	Infested (No.)	Infestation (%)
Replication	2	41.202	0.023	0.021	0.196	0.013	0.221
Treatment	6	141.253**	24.397**	14.464**	8.469**	1.002**	26.492**
Error	12	45.697	0.092	0.053	1.816	0.020	0.504

** : Significant at 0.01 level of significance

Appendix VI. Analysis of variance of the data on incidence of sweet potato weevil on storage roots of sweet potato at 15 days early harvest in number and weight basis for different management practices

Source of variation	Degrees of freedom	Mean square					
		Storage roots of sweet potato in number basis			Storage roots of sweet potato in weight basis		
		Healthy (No.)	Infested (No.)	Infestation (%)	Healthy (No.)	Infested (No.)	Infestation (%)
Replication	2	0.023	0.006	1.339	36.800	0.741	0.014
Treatment	6	0.382**	0.097**	24.138**	3763.029**	761.459**	17.596**
Error	12	0.105	0.008	1.480	1204.061	11.139	0.177

** : Significant at 0.01 level of significance

Appendix VII. Analysis of variance of the data on incidence of sweet potato weevil on storage roots of sweet potato at optimum days harvest in number and weight basis for different management practices

Source of variation	Degrees of freedom	Mean square					
		Storage roots of sweet potato in number basis			Storage roots of sweet potato in weight basis		
		Healthy (No.)	Infested (No.)	Infestation (%)	Healthy (No.)	Infested (No.)	Infestation (%)
Replication	2	0.040	0.008	0.916	22.698	9.829	0.112
Treatment	6	0.324*	0.126**	27.418**	3969.564*	918.037**	19.683**
Error	12	0.098	0.012	1.837	1202.432	19.877	0.272

** : Significant at 0.01 level of significance;

* : Significant at 0.05 level of significance

Appendix VIII. Analysis of variance of the data on incidence of sweet potato weevil on storage roots of sweet potato after 15 days from optimum harvest in number and weight basis for different management practices

Source of variation	Degrees of freedom	Mean square					
		Storage roots of sweet potato in number basis			Storage roots of sweet potato in weight basis		
		Healthy (No.)	Infested (No.)	Infestation (%)	Healthy (No.)	Infested (No.)	Infestation (%)
Replication	2	0.013	0.0001	0.009	64.860	1.613	0.012
Treatment	6	0.393*	0.212**	40.660**	3833.876*	1992.259**	35.242**
Error	12	0.091	0.011	1.131	1209.132	22.418	0.270

** : Significant at 0.01 level of significance;

* : Significant at 0.05 level of significance

Appendix IX. Analysis of variance of the data on yield and yield contributing characters of sweet potato for different management practices

Source of variation	Degrees of freedom	Mean square			
		Storage roots of sweet potato in number basis		Weight of individual storage roots (g)	Yield of storage roots (t/ha)
		Vine length (cm) at Harvest	Number of storage roots/vine		
Replication	2	14.704	0.008	1.432	0.008
Treatment	6	420.515*	0.357**	159.059*	0.145**
Error	12	120.994	0.061	49.524	0.029

** : Significant at 0.01 level of significance;

* : Significant at 0.05 level of significance