

**EFFECTIVENESS OF SOME PLANT MATERIALS
AGAINST JUTE YELLOW MITE, *Polyphagotarsonemus latus*
ON *Corchorus olitorius***

BENJIR AKTER



**DEPARTMENT OF ENTOMOLOGY
SHER-E-BANGLA AGRICULTURAL UNIVERSITY
DHAKA-1207**

JUNE, 2016

**EFFECTIVENESS OF SOME PLANT MATERIALS
AGAINST JUTE YELLOW MITE, *Polyphagotarsonemus latus*
ON *Corchorus olitorius***

BY

BENJIR AKTER

REGISTRATION NO. 15-06873

A thesis

Submitted to the Faculty of Agriculture,
Sher-e-Bangla Agricultural University,
Dhaka in partial fulfillment of the
requirements for the degree of

MASTER OF SCIENCE

IN

ENTOMOLOGY

SEMESTER: JANUARY-JUNE, 2016

Approved by

Dr. Mohammed Ali

Professor

Department of Entomology

Sher-e-Bangla Agricultural University

Supervisor

Dr. Md. Nazrul Islam

P.S.O.

Entomology Department

Pest management division

Bangladesh Jute Research Institute

Co-Supervisor

Dr. Mst. Nur Mohal Akhter Banu

Associate Professor

Department of Entomology

Sher-e-Bangla Agricultural University

&

Chairman

Examination Committee

**Dedicated To
Almighty 'Allah'
&
loving Parents and
Teachers**



DEPARTMENT OF ENTOMOLOGY
Sher-e-Bangla Agricultural University
Sher-e-Bangla Nagar, Dhaka-1207

Memo No: SAU/Entomology/

CERTIFICATE

This is to certify that thesis entitled, “**EFFECTIVENESS OF SOME PLANT MATERIALS AGAINST JUTE YELLOW MITE, *Polyphagotarsonemus latus* ON *Corchorus olitorius***” submitted to the Faculty of Agriculture, Sher-e-Bangla Agricultural University, Dhaka, in partial fulfillment of the requirements for the degree of **Master of Science (MS) in Entomology**, embodies the result of a piece of bona fide research work carried out by **Benjir Akter, Registration No 15-06873** under my supervision and guidance. No part of the thesis has been submitted for any other degree or diploma.

I further certify that any help or source of information, as has been availed of during the course of this investigation has duly been acknowledged.

Dated: June, 2016

Place: Dhaka, Bangladesh

Prof. Dr. Mohammed Ali

Supervisor

ACKNOWLEDGEMENTS

"Absolute trust and faith in the Almighty Allah will be the basis of all actions". First of all the author would like to express her deepest sense of gratefulness to Almighty Allah, who enables her to complete her research work and submit her thesis for degree of Master of Science (MS) in Entomology.

The author expresses her sincerest gratitude, heartfelt respect immense indebtedness and profound respect to her reverend Supervisor **Dr. Mohammed Ali**, Professor, Department of Entomology, Sher-e-Bangla Agricultural University, Dhaka for his scholastic guidance, affectionate feelings, invaluable suggestions, continuous encouragement and blessing in conducting the research work and constructive criticism in preparing the manuscript of the thesis like a pathfinder pole-star.

The author is honored to express her respect to her profound Co-supervisor **Dr. Md. Nazrul Islam**, Principal Scientific Officer & Head of the Department of Entomology, Pest management Division, Bangladesh Jute Research Institute, Dhaka for his encouragement, co-operation, kind comments and invaluable suggestion in improving the manuscript and in preparation of their thesis.

The author expresses her sincere respect to **Dr. Mst. Nur Mohal Akhter Banu**, Associate Professor, Department of Entomology, Sher-e-Bangla Agricultural University, Dhaka and chairman, Examination committee for her valuable suggestions and pleasant cooperation during the study period.

The author also expresses heartfelt thanks to all the teachers of the Department of Entomology, SAU, for their valuable suggestions, instructions, cordial help and encouragement during the period of the study.

Finally, the author would like to express her gratefulness to her beloved father Amir Hossain, mother Shamima Sultana and all the well-wishers for their blessings, sacrifices, inspirations and moral support, which opened the gate and paved the way to her higher studies.

June, 2016

The Author

**EFFECTIVENESS OF SOME PLANT MATERIALS AGAINST JUTE
YELLOW MITE, *Polyphagotarsonemus latus* ON *Corchorus olitorius***

BY

BENJIR AKTER

ABSTRACT

In order to study the effectiveness of some plant materials against jute yellow mite, *Polyphagotarsonemus latus* (Banks) on *Corchorus olitorius* (O-9897) experiments were conducted in the greenhouse premises and in the field of Bangladesh Jute Research Institute during the period from March to October 2016. In the greenhouse premises, the highest percent mortality 69.39%, 67.77%, 63.86%, 62.43% and 61.47% @ 1:20 were recorded from neem seed kernel extract, mahogany seed extract, pithraj seed extract, turmeric powder extract and green neem leaf extract, respectively. In field experiment, highest percent reduction of infestation over control (70.20%), lowest number of nodes per plant (52.50), highest plant height (3.10 m), and highest percentage of yield increased (38.60%) over control were observed in the plot treated with neem seed kernel extract. In conclusion, neem seed kernel extract, mahogany seed extracts, pithraj seed extract, green neem leaf extract and turmeric powder extract @ 1:20 can be used safely by the farmers as component of Integrated Pest Management (IPM) Program of jute.

CONTENTS

CHAPTER	TITLE	PAGE
	ACKNOWLEDGEMENTS	I
	ABSTRACT	II
	TABLE OF CONTENTS	III-IV
	LIST OF TABLES	V
	LIST OF PLATES	VI
I	INTRODUCTION	1-3
II	REVIEW OF LITERATURE	4-24
2.1	Yellow mite (Broad mite)	4
2.2	Description	4
2.3	Biology and life cycle	5
2.4	Origin and distribution of Yellow mite	6
2.5	Host range	7
2.6	Nature and extent of damage	7
2.7	Behavior of Broad mite	8
2.8	Effect of different plant extracts for controlling mite pest	8
2.9	Effect of different plant extracts on different insect pests	10
2.10	Effect of neem oil on insect growth responses	19
III	MATERIALS AND METHODS	25-37
3.1	Experimental site	25
3.2	Soil	25
3.3	Climate	25
3.4	Land preparation	26
3.5	Experimental design and layout	26
3.6	Plot size	26
3.7	Fertilizer Application	26
3.8	Planting materials	27

CONTENTS (CONTD.)

CHAPTER	TITLE	PAGE
3.9	Pot preparation	27
3.10	Inter cultural operation	27
3.11	The test pests	27
3.12	Details of the treatments	30
3.13	Extraction and formulation of some plant materials	31
3.13.1	Preparation of green neem leaf extract	32
3.13.2	Preparation of mahogany seed extracts	32
3.13.3	Preparation of neem seed kernel extract	32
3.13.4	Preparation of pithraj seed extract	32
3.13.5	Preparation of Turmeric powder extracts	33
3.14	Experimental details	36
3.14.1	Pot experiment	36
3.14.2	Experiment in field condition field	36
3.15	Statistical analysis	37
IV	RESULTS AND DISCUSSION	38-52
4.1	Effect of aqueous extraction of different plant materials on jute yellow mite	38
4.2	Effect of different plant materials on yellow mite (1 st spray) under field condition.	40
4.3	Effect of different plant materials on yellow mite (2 nd spray) under field condition.	41
4.4	Effect of different plant materials on internodes of jute plant.	42
4.5	Effect of different plant materials on plant height.	47
4.6	Effect of different plant materials on base diameter of jute.	48
4.7	Effect of different plant materials on fibre yield of jute	49
VII	SUMMERY AND CONCLUSION	53-54
VIII	REFERENCES	55-66

LIST OF TABLES

TABLE NO.	TITLE	PAGE
1	Effect of plant materials (aqueous extraction) on % mortality of jute yellow mite in the pot.	39
2	Effect of different plant materials on % reduction of yellow mite in field condition (First spray) over control.	44
3	Effect of different plant materials on % reduction of yellow mite in field condition (2nd spray) over control.	45
4	Effect of different plant materials on number of inter nodes of Jute plant.	46
5	Effect of different plant materials on plant height.	50
6	Effect of different plant materials on Base Diameter.	51
7	Effect of different plant materials on fibre yield.	52

LIST OF PLATES

PLATES NO.	TITLE	PAGE
1	Magnified picture of mite	5
2	Egg and nymph of jute yellow mite	5
3	A partial view of field experiment	28
4	Egg & nymph of yellow mite	28
5	Adult Female (A) and male(B) jute yellow mite	29
6	Yellow mite infested plant	29
7	Pot experiment	30
8	Green neem leaf extract	33
9	Pithraj seed extract	34
10	Mahogany seed extract.	34
11	Turmeric Powder extract.	35
12	Neem seed Kernel extract.	35
13	Severely yellow mite infested jute plant showing internodes and increased number of nodes.	47

CHAPTER I

INTRODUCTION

Jute, a fibre crop of international eminence, is the most important cash crop and foreign exchange earner of Bangladesh. It is cultivated for its phloem fibre, which yields the strongest and most durable fibre of commerce. Jute fibre is extensively used in the world for its versatility, durability and fineness as it is used for the production of newsprint, carpet making, hessians, gunny bags, ropes etc. Our agricultural community is dependent to a large extent on jute and jute products. Jute was described as a very important commodity concerning agriculture industry and trade in Bangladesh.

Jute is mostly grown in the Indo-Bangladesh region and in some countries of the South East Asia. Jute ranks second only to cotton among all the natural fibre in case of production (Talukder *et al.* 1989). It has been reported that about 90% of World's jute produced in Bangladesh and India (Atwal, 1976). With respect to production, Bangladesh ranks second among the jute growing countries of the world. The land and climatic conditions of Bangladesh are congenial to the production of good quantity of jute and two species *Corchorus capsularies L.* and *Corchorus olitorius L.* are cultivated for fibre. In Bangladesh, about 6.73 lac hectares of land are under jute cultivation and the total yield is 1633152.65 metric ton (BBS, Agriculture 2015). It is worthy to note that 100 thousand traders and 250 thousand industrial labours earn their livelihood from the jute business (Khandaker, 1987).

Jute is liable to damage by about 40 species of insects and mites at all stages of the growth from seedling to harvest (Kabir, 1975). Among them yellow

mite *Polyphagotarsonemus latus* (Banks) is one of the most common and a serious pest of jute (*Corchorus spp.*). In Bangladesh, jute crop is frequently attacked by yellow mite, and as a result of its infestation, the plant is severely affected. About 38% loss in fibre yield by yellow mite was estimated under field condition (Kabir, 1975). The yellow mite of jute commonly known as yellow tea mite is a very destructive pest and causes damage to both fibre and seed crops. Its damage is better known as “Telenga” or “Telchita” disease in Bangladesh. It appears on jute at the end of April when the plants are about one feet tall. The jute yellow mite becomes active in mid-May (Kabir, 1975). The damage of the terminal shoots is seldom visible before June. Initial mite attacks are usually seen near dwellings and shady places of leaves. It seems that the mite is carried from plot to plot of the jute planting by wind. The adult mites also play an important role in the dispersal and distribution by carrying female nymphs to younger leaves.

The nymphs are held above the males body by means of a sucker like organ near the tip of Posterior terminus. Both yield and quality of fibre are reduced due to attack of this pest. Management of mite is based mainly on its chemical control. But the use of chemical acaricides may cause pest resurgence and their residual effect resulting in environmental, social and other problems. To minimize the use of these in mite control programs, alternative substances are now strongly felt in many developed countries. The biologically active natural plant products can play a significant role in this regard. These products may help to keep the drawbacks of conventional methods within bounds. Plant products are environmentally safe, less hazardous, less expensive, biodegradable and readily available.

Many plants exhibiting pesticidal properties have been known since time

of immemorial. Over the past 50 years, more than 2000 plant species belonging to different families and genera have been reported to contain toxic compounds and a multitude of chemical compounds possessing diverse and novel types of structural patterns have been isolated from various plants (Adityachaudhury *et al*, 1985). In recent years the derivatives of neem (*Azadirachta indica*) have come under close scrutiny of scientist around the world as the most promising source of natural insecticides (Saxena, 1989). However, reports on the use of neem oil in jute pest management are scanty in Bangladesh. Neem has been reported to have antifeedant, repellent, toxicant, insect growth inhibitors, chemosterillant and anti oviposition activity (Gujar, 1992).

In Bangladesh, few studies have been conducted on the efficacy of different plant materials against yellow mite attack. Therefore, an attempt has been made to provide information for using plant materials as input for the control of yellow mite. Thus, the study was undertaken to fulfill the following objectives:

- To identify the effective plant materials for the control of jute yellow mite.
- To evaluate the efficacy of different plant materials against jute yellow mite and
- To determine the effective doses of selected plant materials against jute yellow mite infestation.

CHAPTER II

REVIEW OF LITERATURE

2.1 Yellow mite (Broad mite)

The yellow mite, *Polyphagotarsonemus latus*, was first described by Banks, (1904) as *Tarsonemus latus* from the terminal buds of mango in a greenhouse in Washington, D.C., USA (Denmark, 1980). This species has a large host range world-wide.

2.2 Description

Eggs: The eggs are colorless, translucent and elliptical in shape. They are about 0.08 mm long and are covered with 29 to 37 scattered white tufts on the upper surface (Denmark, 1980; Pena and Campbell, 2005; Baker, 1997).

Larvae: Young broad mites have only three pairs of legs. They are slow moving and appear whitish due to minute ridges on the skin (Peña and Campbell, 2005). As they grow they range in size from 0.1 to 0.2 mm long (Anonymous, 2005).

Nymph: After one day, the larva becomes a quiescent nymph that is clear and pointed at both ends. The nymphal stage lasts about a day. Nymphs are usually found in depressions. Female nymphs are often carried by males (Pena and Campbell, 2005).

Adults: Female mites are about 0.2 mm long and oval in outline. Their bodies are swollen in profile and a light yellow to amber or green in color with an indistinct, light, median stripe that fork near the back end of the body. Males are similar in color but lack the stripe. The two hind legs of the adult females are reduced to whip-like appendages. The male is smaller

(0.11mm) and faster moving than the female. The male's enlarged hind legs are used to pick up the female nymph and place her at right angles to the male's body for later mating (Pena and Campbell, 2005).

2.3 Biology and life cycle

The yellow mite has four stages in its life cycle: egg, larva, nymph and adult. Adult females lay 30 to 76 eggs (averaging five per day) on the undersides of leaf surface and in the depressions of small fruit over an 8 to



Plate 1: Egg and nymph of jute yellow mite



Plate 2: Magnified picture of mite

13-day period and then die. Adult males may live 5 to 9 days. While unmated females lay male eggs, mated females usually lay four female eggs for every male egg. The eggs hatch in two or three days and the larvae emerge from the egg shell to feed. Larvae are slow moving and do not disperse far. After two or three days, the larvae develop into a quiescent larval (nymph) stage. Quiescent female larvae become attractive to the males which pick them up and carry them to the new foliage. Males and females are very active, but the males apparently account for much of the dispersal of a mite population in their frenzy to carry the quiescent female

larvae to new leaves, when females emerge from the quiescent stage, males immediately mate with them (Anonymous, 2005; Baker, 1997; Pena and Campbell, 2005). There are also reports of the broad mite using insect hosts, specifically some whiteflies species, to move from plant to plant (Palevsky *et al*, 2001).

Female *P. latus* lays 30 to 76 eggs on the leaf surface over an 8- to 13-days oviposition period. The larvae hatch in 2 or 3 days and emerge from the egg shell to feed. Larvae are slow moving and do not disperse far. In 2 or 3 days, the larvae develop into a quiescent larval stage. Quiescent female larvae become attractive to the males which pick them up and carry them to the new foliage. Males live 5 to 9 days; females live 8 to 13 days.

2.4 Origin and distribution of yellow mite

P. latus has a worldwide distribution and is known by a number of common names. It is found in Australia, Asia, Africa, Europe, North America, South America, and the Pacific Islands. In India and Sri Lanka it is called the "yellow tea mite," while those in Bangladesh call it the "yellow jute mite." In some European countries it is called the "broad spider." In parts of South America it is called the "tropical mite" or the "broad rust mite" (Anonymous, 2005).

2.5 Host range

The yellow mite has a wide host range in tropical areas. It attacks greenhouse plants in temperate and subtropical regions (Pena and Campbell, 2005). Crops listed as hosts include: apple, avocado, cantaloupe, castor, chili, citrus, coffee, cotton, eggplant, grapes, guava, jute, papaya, passion fruit, pear, potato, sesame, string or pole beans, mango, tea, tomato (Pena and Campbell, 2005). It was identified for the first time on

watermelons in the U.S.A in 2006 (Pons, 2007). Broad mites infest many ornamentals, including African violet, ageratum, azalea, begonia, chrysanthemums, cyclamen, dahlia, gerbera, gloxinia, ivy, jasmine, impatiens, lantana, marigold, peperomia, pittosporum, snapdragon, verbena, and zinnia (Baker, 1997).

The broad mite is considered a serious pest of *Pittosporum* spp. in Florida (Johnson and Lyon, 1991).

2.6 Nature and extent of damage

This mite is considered a sub-major pest at low elevations in summer months. Mites feed by piercing plant cells and sucking up the sap that oozes from the wound (Waterhouse and Norris, 1987). Reduction in photosynthesis and instability of water balance are some of the damaging effects to plants. Feeding damage also causes terminal leaves and flower buds to become cupped and distorted. As a result of feeding injury, corky brown areas appear between the main veins on the underside on the leaf. Young foliage sometimes becomes rust colored and nearly always is deformed. Blooms abort, and the plant growth is stunted. Damaged leaves often become discolored, thickened and brown (Lacob, 2007).

Damage of flowers and fruits differ among plants. On macadamia, flowers and leaves may appear corky brown or bronze colored. On lemon fruits, feeding results in silvering of the fruit that greatly reduces the market value even though the fruit is otherwise unaffected (Waterhouse and Norris, 1987). On beans, extensive feeding on fruit results in a purplish or dark brown appearance of the pods.

The broad mite does not vector of any known plant virus diseases (Waterhouse and Norris 1987; Higa and Namba, 1970).

2.7 Behavior of broad mite

Larvae and adults prefer to feed on the undersides of leaves usually in the vicinity of the egg. The adult males have a specialized last pair of legs and are often seen carrying a female pupa over its body. Male pupae are usually not moved, but migrate to new leaf growth carrying a developing female pupa soon after the adult male emerges (Hill, 1983). The males often carry the pupae to newly opened leaves. Copulation occurs immediately after the female has emerged from the pupal skin.

Relative to the males, females are relatively sedentary (Waterhouse and Norris, 1987), but the species is generally considered fast moving (Brown and Jones, 1983).

2.8 Effect of different plant extracts for controlling mite pest

Banu (2007) conducted an experiment to compare some non-chemical approaches to control jute yellow mite and jute hairy caterpillar in greenhouse and field condition during 2004-2006, In greenhouse condition, double spray of green neem leaf extract @ 1:20 and dry neem leaf @ 1:50 was found to be effective and gave 74.63% and 70.83% mortality 72 hr after treatment on potted plants. However, in field condition, double spray of green neem leaf extract and dry neem leaf extract gave 67.70 % and 72.20% reduction of infestation 7th day after spray. The fibre yield was also increased by 19.9% for green leaf extract and 35.9% for dry neem leaf over the control treatment, respectively. In addition, hand picking method was found to be effective, easiest, economic and eco-friendly.

Banu and Singh (2007) studied the effect of neem leaf extracts (dry and green) in green house and field condition during the period from 2003-05

against jute yellow mite. Dry neem leaf @ 1:50 and green neem leaf @ 1:20 gave 62% and 64% mortality, respectively. Fibre yield was increased by 39.95% and 35.93% over the control treatment.

Pasini et al. (2003) Observed that the effect of a commercial formulation of neem oil at different stages of the life cycle of red mite of paraguaytea showed that the formulation was efficient in controlling adults. Azadirachtin also affected the fecundity of the female mites.

Palaniswamy and Ragini (2003) sprayed 5% aqueous extracts of *Adathoda vasica*, *Vernonia negundo*, *Azadirachta indica*, *Aristolochia bracteata*, *Lippia nodi* (flora), *Argemone mexicana*, *Sansevieria* sp. and *Aloe* sp. on chilies 30 days after transplanting in Tamil Nadu, India. The polyphagotersonemus latus populations were reduced and *Aloe* sp. also recorded to be the lowest population at 0.67 mites leaf⁻¹.

Sanguanpong and Schmutterer (1992) found that pentane extract and cold pressed neem oil reduced the fecundity of the mites on treated plants and the survival of nymph hatched from treated eggs. Application of pentane extract or neem oil in sublethal concentration caused growth disrupting effects on the nymphal stages and ovicidal effects.

Pande et al. (1987) they found neem leaf extract 1% and neem seed kernel 5% to be effective against *Tetranychus neocaledonicus* and *Tetranychus urticae* respectively.

2.9 Effect of different plant extracts on different insect pests

Jagjeet et al (2005) treated pigeon pea seeds with 11 seed protectants, i.e. neem seed kernel powder at 20g, neem oil at 10 ml, mustard oil and groundnut oil each at 7.5ml, turmeric powder at 3.5g, mustard oil+turmeric powder at 3.75ml+1.75g, groundnut oil+turmeric powder at 3.75ml+1.75g each per one kg of seed, covering 4m with each of seed, dung cake ash,

sawdust and wheat husk and mixed them with half kg of seed by shaking it manually. All the seed protectants, except for sawdust and turmeric powder, recorded significantly higher adult mortality than the control after the first day of treatment. Neem oil was effective (64.33% adult mortality) up to 35 DAT and it was followed by mustered oil+turmeric powder; which recorded only 16.33% adult mortality. While other treatments were not effective.

Zhu *et al* (2004) observed biological activity of azadirachtin on rice stem borer, *Chilo suppressalis*. After feeding on water-oats treated with 0.75 and 0.50mg azadirachtin/litre, the third instar larvae were completely dead in 3 and 6 days respectively. Mortality of the newly hatched *C. suppressalis* reached 100% within 24h after treatment with 6, 3, and 2mg azadirachtin/lit.

Maisary *et al*. (2004) examined the effect of neem oil on the 2nd and 4th instars and eggs of *Culex pipiens* under laboratory condition. 46.98% of *C. pipiens* were killed upon exposure to 1000ppm of neem oil. The lower concentration (10ppm, 100ppm) showed little efficiency on the eggs. The continuous treatment of the 2nd and 4th instars with neem oil (100ppm) caused high mortality and complete inhibition of the formulation of mature instars. It is concluded that in general, exposure to neem oil for a short period (24 and 48 hours) is less effective as compared with continuous.

Rahman *et al* (2003) conducted an experiment to evaluate five indigenous plants seed oils viz Castor, Neem, Pithraj, Safflower and Sesame at concentration of 1, 2, 3, 4, and 5% revealed that all the plant seed oils have grain protectant value against lesser meal worm. The results showed that sesame and neem oil were more effective than Castor, Pithraj and Safflower oils. The tested seed oils provided good protection for wheat grains.

Eungwijarupanya *et al.* (2002) tested neem extracts containing 0.185% azadirachtin at 3 concentrations 100ml, 200ml and 300ml diluted in 5 litres of water, These were applied using a thermal fogger to a 15 years old teak (*Tectona grandis*) for control of teak defoliator, *Hybiaea puera*. After application larvae were collected and reared in the laboratory to observed mortality. One day after fogging mortality started to increase for these treated with 200ml and 300ml/5L concentrations and all larvae died within 6 days when treated with 300ml/5L.

Padmasheela and delvi (2002) tested a commercial formulation of neem oil EC (Nimbex, 0.03%) at different concentration viz. 25ppm, 50ppm, 75ppm, and 100ppm for mortality effects against grubs of *Oryctes rhinoceros* (a coconut pest) at laboratory conditions In feeding toxicity test, neem oil at concentrations of 50ppm, 75ppm, and 100ppm caused 20%, 45% and 90% mortality respectively on exposure up to 96hr and 100ppm caused 90.67% mortality of *Oryctes rhinoceros* grubs.

Qureshi *et al* (2002) investigated the direct effect of neem extracts on the adult glass beetle, *Costelytra zealandica* where laboratory bioassay showed that neem caused only low mortality even at the highest dose.

Karmakar and Bhole (2001) observed the efficacy and persistent toxicity of some neem products- neem oil, and nimbecidine against adult of *Epilachna dodecastigma*, the treatments with 2% neem oil and 2% nimbecidine resulted 90.69% and 71.90% mortality respectively.

Shaminathan and Jayaraj (2001) conducted two experiments to evaluated botanical pesticides like Ipomoea leaf extracts, neem oil and madhuca oil (at 0.3% or 3.0% each) against *Perrisia virgate*. The leaf dip method was used in both experiments and pest mortality was recorded at 24hr, 48hr and 72 hr after treatments. In experiments 1, treatments with 3% neem oil

recorded the highest mortality (43.13%). Neem recorded 50% mortality at 72hr, and in experiment 2, at 48hr, fortified (0.3%) neem oil recorded a maximum mortality of 49.3% and at 72hr, fortified neem recorded 63.6% mortality. Arcos *et al.* (2001) conducted that the effect of neem oil was evaluated by using concentrations of 0.5, 1.0, 1.5, 2.0, and 3.0%. They mainly showed that mortality recorded by ingestion was attributed to starvation. Filter paper soaked with neem oil inhibited feeding of *I. marginipennis*.

Rani *et al.* (2000) investigated the efficacy of cotton seed, neem, palm, rice bran and soybean oils as seed coating against pulse beetle, *Callosobruchus chinensis* infesting Chickpea and found that neem oil at 1 ml/kg of seeds gave the highest adult mortality (65%) three days after treatment.

Ranjana *et al.* (2000) tested five plant extracts from *Azadirachta indica* kernels, Bulbs of *Allium sativum* and *Citrus sinensis* rech, *Citrus limm* peels and *Mangirfera indica* leaves each having three concentration (1%, 1.5% and 2%) against pulse beetle, *C. maculatus*. The petroleum ether extract of neem kernel was most effective as 1.5% and 2% level showed 50% and 61.11% mortality.

Sharma (1999) reported that neem seed (*Azadirachtin indica*) kernel powder at 4% and neem leaf powder at 5% protected maize for 5 moths against *Sitophilus oryzae*, *Sitotroga cerealella*, *Rhyzopertha dominica* and *Trogoderma granarium*. Neem oil (Neembicidine 2%) effectively reduced the emergence of F1 and F2 progeny of all the pests and completely protected maize up to 9 months and suggested that neem products can be mixed with stored maize to protect the grains up to 9 months from the attack of this major pest.

Reddy *et al* (1999) stated that application of neem oil (*Azadirachtin*), Karanja oil (*Pongamia glubra*), Mahua oil (*Madhuca lalifolia*) and palmolein oil (*Elaeis gaineenis*) at dosages of 0.5% and 1.0% level effectively protected green gram from *C.chinensis*. Neem oil at 1% level was the best protected followed by palmolein, Karanja and mahua oils. These oils also exhibited contact toxicity and no adults could survive in neem treated green gram at 5% concentration.

Mayabini (1997) studied the efficacy of neem bark decoction, neem based chemicals and neem derivatives (neem oil, leaf extract and leaf decoction) against rice leaf folder *Cnaphalocrosis medinalis*. Ail were applied as foliar sprays to pot-grown rice plants Leaf area fed by the larvae was recorded after 48 hours. Neem bark decoction appeared to be a very effective botanical for controlling the rate of feeding and reducing the rate of population.

Naganagouda *et al* (1997) conducted a field study to determine the efficacy of various insecticides and neem products for the control of *Nilapaevata lugens*, *Sogatella furcifera*, *Cnaphalocrosis medinahs* and *Scirpophaga incertulas* on rice Monocrotophos was the most effective insecticide in terms of giving the highest yield followed by neem oil and Nimbicidin.

Deka and Hazarika (1997) observed that neem (*Azadirachta indica*) seed oil (NSO) acted as a potential antifeedant against adult of the chrysomelid, *Dicladispa armigera* Under laboratory conditions, daily consumption of fresh rice leaves was 1.05 g, 0.08 mg, which was reduced by 50% when leaves were treated with 6.46% NSO.

Raguraman and Rajasekaran (1997) stated the effect of neem oil and neem seed bitters applied at different concentrations as either high volume, low volume or ultra-volume sprays to the rice brown plants hopper,

Nilaparvata lugens. All neem products affected the orientation, probing and feeding time, food ingested and growth and development of *Nilaparvata lugens*.

In a laboratory study Haque *et al* (1996) found that when first and third instars larvae and adults of *Epilachna dodecastigma* were exposed to 0.25, 0.50, 1.0 or 2.0% neem (*Azadirachta indica*) oil applied on brinjal leaf discs, all the first instar larvae were killed at the concentrations before feeding and the feeding activity of third instar larvae and adults decreased with increasing oil concentration.

Braman (1993) observed antifeedant effects of azadirachtin in nymphal tawny mole crickets, *Scapteriscus vicinus* Scudder, in laboratory tests. Crickets surviving treatment grow more slowly and tunneled less than their untreated counterparts.

Islam (1993) determined the comparative efficacy of azadirachtin, the major active ingredient in neem, as a feeding deterrent for six species of noctuidae of economics importance, the black army worm, *Actebta femtica* Tansch, bertha army worm, *Manestra configweata* Walker, variegated cutworm, *Peridroma sancia* Hubner, zebra caterpillar, *Melanra picta* Harr, Asian armyworm, *Spodoptera litura* Fab and the cabbage looper, *Trichoplusia ni* Hubner Fourth instar larvae of *Spodoptera litura* was the most sensitive to the antifeedant effects of azadirachtin whereas *Actebia fermica* was the least.

Nesseh *et al.* (1993) tested the repellent effect of neem oil on adults of *Schistocerca gregaria*. They found that *Schistocerca gregaria* consumed 100% of the leaves of the untreated plant, while the adults started feeding on treated plant after 24 hours of the application.

The antifeedant properties of the seeds of some meliaceous plants were reported by Chiu-ShinFoon and Qiu-Yu-Tong (1993). In their experiments with neem seed oil and petroleum ether extracts of the seed kernels of two species of china berry they demonstrated their potentials as strong antifeedant against nymphs of brown plant hopper, *Nilaparvata lugens*.

Salsoloy and Embuido (1992) evaluated neem oil for its insecticidal action on cotton boll worm, *Helicoverpa armigera* Hubner. The oil applied along sprayed on cotton and the effects were compared. Neem oil; sprayed on cotton gave poor control of the pest.

Salem (1991a) found that 100 ppm concentration of neem oil extract was the most effective extract against larval feeding of potato tuber moth, *Phthorimaea operculella* Zell.

Salem (1991b) tested pure neem seed oil against the cotton boll worms, *Pectinophora gossypiella* Saund and *Earias insulana* Boisd. The most active concentration caused reduction in the percentage of infestation nearest to 150 ppm. The percentages of infestation decreased with the increase of neem seed oil concentrations.

Rovesti and Deseo (1990) stated that neem, (*Azadirachta indica*) and its oil, extracts and derivatives including azadirachtin are used as antifeedants, repellents, ovicides and growth regulators, they can also reduce adult fecundity and egg viability.

Gonzalez-Gomez *et al.* (2006) conducted an experiment to evaluate the acute toxicity of crude neem seed extract and neem based commercial product (0, 1, 2, 3 and 4%) on *Varroa destructor* (Aceri: Varroidae) and *Apis mellifera* and repellence of varroa mites. They found neem based products had a persistent repellency effect that lasted approximately 48 h.

Islam (2006) conducted the experiment on the exploration of plant materials extracts for the control of jute yellow mite, *Polyphagotarsonemus latus* (Banks) was conducted at the Department of Entomology, Bangladesh Jute Research Institute, Manik Mia Avenue, Dhaka, during the period from May to August, 2005. The treatments were green neem leaf extract (1:10), green neem leaf extract (1:20), neem oil 1%, neem oil 0.5%, onion bulb extract (1:10), onion bulb extract (1:20), green mahua leaf extract (1:10), green mahua leaf extract (1:20), green nayantara leaf extract (1:10), green nayantara leaf extract (1:20), green karabi leaf extract (1:10), green karabi leaf extract (1:20) and untreated control. Neem oil was emulsified at 0.5 & 1.0% with 0.1% Nikalin. The minimum number of yellow mite was observed in T₃ (neem oil at 1%) during all stages of the plant and it reduced the height percent of mite population over control. Neem oil (1%) significantly reduced the population of yellow mite, increased plant height and numbers of leaves per plant and decreased the number of increased leaves and nodes per plant.

Gupta (2003) examined the efficacy of neem products viz., neem kernel extract in water or cow urine, neem leaf extract in cow urine and neem oil in controlling the major pests (*Antigastra catalaunalis* and *Dasineura seasami*) of sesame cv. Flower. The capsule damage was the lowest with the application of neem oil and endosulfan, respectively. Application of neem oil resulted lowest bud and flower damage (13.3%) and highest grain yield (6.55 kg/ha) gave net profit (Rs. 2633/ha).

Jyothi and Sannaveerappanavar (2003) conducted an experiment on contact toxicity of aqueous neem seed kernel extract and neem seed oil emulsion to egg, larval, pupal and adult stages of *Chellomenes sexmaculata* (Fabricius) (Coleoptera: Coccinellidae) under laboratory condition. Neem oil emulsion at 2 and 4% and NSKE at 2 and 4% significantly affected the

egg hatching (80.0, 75.0 and 82.5%, respectively) compared to the control (95.0%). Aqueous NSKE at 4% was relatively more toxic (22.0% mortality) to the 3rd instar larvae.

Imtiaz *et al* (2001) studied the effects of neem leaf extracts on adult rice weevil, *Sitophilus oryzae*. Glass film method was adopted to determine the LC₅₀ rate. After plotting a graph between mortality and concentration, the LC₅₀ was found to be 0.44 µg/sq. cm.

Jeyakumar and Uthamasamy (1997) carried out an experiment in the laboratory to study the efficiency of some botanicals against *Liriomyza trifoli* on cotton. Application of some botanicals insecticides such as neem oil (3%) and neem seed kernel extract (5%) resulted higher larval mortality. Whereas *Pongamia pinnata* oil (3%), neem oil (2%), neem seed kernel extract (2.5%) were found least effective.

Behera and Satapathy (1996) evaluated 7 plant materials viz. *Azadirachta indica*, *Armona reticulate*, *Datura anoxia*, *Callophyllum inophyllum*, *P. pinnata*, *Calotropis gigantean*, *Thevelia peruviana*. They found that aqueous extract of neem seed kernel was the most effective causing 100% mortality of *Spodoptera litura* at 10 days after treatment among other 6 indigenous plant extract.

Kareem and Durairaj (1987) showed that extracts of Neem Seed Kernel (NSK) when evaluated at field, 4% NSK significantly reduced Green Leaf Hopper (GLH), Brown Plant Hopper (BPH) and White Backed Plant Hopper (WBPH) population and Leaf Folder (LF) damage. This crude water extract caused greater BPH and LF feeding inhibitor and WBPH nymphaal emergence.

Kareem and Durairaj (1987) evaluated crude extracts of neem seed kernel (NSK) and neem cake (NC) in water and neem oil (NO) emulsion along with two synthetic insecticides in fields as foliar sprays for the control of

major insect pests of rice. NSK 4% significantly reduced green leafhopper (GUI). Brown Plant Hopper (BPH) and White Backed Plant Hopper (WBPH) populations and leaf folder (I J^r) damages in two fields trials proving either as per with or next in efficacy to fethion and phosphamidon spraying.

Jotwani and Srivastava (1984) reported that antifeeding properties of neem seed kernel suspensions against desert locust, *Schitocerca gregaris* Forsk; Castor hairy caterpillar, *Cuproctis lunata* W., Tobacco caterpillar, *Spodoptera litura* F., Bihar hairy caterpillar, *Utethesia pulchella* L. Grasshoppers, *Acrida exaltata* W. Antifeedant properties of *A. indica*, *M. azadirach*, *M. toosendan* were investigated by Shin-Foon and Zhang (1984) on fifth instar larvae of *S. litura* and were sensitive to neem treated leaf discs in choice tests.

Islam (1993) revealed that oil neem, extracts of leaves and seeds of *A. indica*, *M. azadirach*, *A. ruhituka* and *A. reticulate* with hexen diethyl ether. 95% ethanol and acetone showed potential as antifeedant or feeding deterrents for the control of brown rice plant hopper (BPH), rice green leaf hopper (RGLH), rice hispa (RH) and lesser rice weevil (LRW). The young seedling of rice sprayed with 8-12% of crude and emulsified neem oil also significantly reduced feeding in brown hopper and green rice leaf hopper. Aqueous and methanol extracts of neem and chinaberry also deterred feeding in adult pulse beetle & early instar larvae of jute hairy caterpillar.

2.10 Effect of neem oil on insect growth responses

Sudipta and Sanjib (1998) reported that larvae of rice moth, *Corcyra cephalonica* (Stainton) were maintained in neem oil (Azadirachtin, 0.03%) absorbed crushed jowar (sorghum) grains in four doses (0.25, 0.50, 0.75, and 1.0 ml, each dose in 20 g of food), with an initial population of 50 newly hatched larvae/100 g of neem absorbed food in each replication

Deformed adults with a prolonged period of development were obtained. Growth inhibition, developmental disturbances and mortality increased markedly with increased doses.

Lowerry *et al.* (1996) reported that neem (*Azadirachta indica*) seed oil (NSO) added to meridic diet at concentration as low as 0.016% reduced pupation and prevented adult eclosion rate of obliquebanded leaf roller (*Choristoneura rosaceana*). At a rate of 0.0016% NSO reduced the fitness of *Choristoneura rosaceana*, resulting in longer developmental times, lower adult eclosion rates, and reduced egg production compared with controls. Pupation was completely inhibited at concentration of 0.25 and 1.0% for larvae exposed in the fifth or sixth instar, respectively; rates as low as 0.016% reduced pupal weights and eclosion rates. For larvae transferred to treated diet in the fifth instar, physical abnormalities in the wings of adults occurred at a rate of 0.004% NSO and increased with increasing treatment rates.

Nauman and Islam (1995) found that applications of 3 concentration of oil-free neem seed extracts to cabbage plants in cages did not deter oviposition by individuals of 3 species of noctuid moths, *Trichoplusia sp*, *Peridroma saucia* and *Spodoptera litura*. 1% crude oil emulsion significantly reduced the proportion of eggs laid by *S. litura* on treated plants.

Freisewinkel (1993) found the contact effects of neem oil topically sprayed on third instar nymph of *Locusta migratoria migratorioides*. In parallel experiment neem oil was applied directly to the abdomen of the nymphs. Feeding larvae at the beginning of the third instar with maize leaves treated with neem oil tested the effectiveness of neem oil given orally. The mortality in feeding experiments was much higher than in spraying or direct application experiments. Treated locusts showed prolonged nymphal developmental and reduced increase in weight.

Nicol (1993) studies the effects of the neem seed oil in third instar nymphs of *Schistocerca gregaria*. In cages, which were sprayed with neem oil, the locust showed higher mortality rates, delayed nymphal development and morphogenetic effects of antennae, eyes and wings. Moreover, the adults derived from treated nymphs were smaller in size than those in the control.

In laboratory experiments Venkateswarlu *et al.* (1993) studied the effect of neem oil (0.1, 0.25, 0.50, 1.0 and 1.25%) on growth and development of *Lipaphis erysimi*. At concentration of 1.0, 1.25 and 1.50% all the nymphs reared on treated Indian mustard leaves diet before reaching the adult stage. At the lower concentration nymphal survival, fecundity and growth index of the aphid decreased and developmental period increased.

Rao *et al.* (1993) tested, Neemark, Biosol, Repelin and neem oil at 0.5-3.0% against larvae of *Spodoptera litura* in the laboratory. Repellency, antifeedant activity and developmental period increased with increase in concentration of all pesticides. Adult's emergence, growth, survival, larval and pupal weight, number of eggs laid and hatchability of eggs decreased with increase in concentration. Neem oil had the greatest effect, followed by Neemark, Biosol and Repelin.

Islam (1993) stated the efficacy of azadirachtin, the major active ingredient in the botanical insecticide neem, as a larval growth inhibitor and feeding deterrent for six species of noctuids of economic importance: the black army cutworm, *Actebia fennica*, Tansch, the bertha army worm, *Mamestra configurata* Walker, the variegated cutworm, *Pendroma saucia* Hubner, the zebra caterpillar, *Melanchra pieta* Harr, the Asian armyworm, *Spodoptera litura* Fab and the cabbage looper, *Trichoplusia ni* Hubner. When added to an artificial diet, azadirachtin inhibited normal growth of all species in a dose dependent fashion.

Becker *et al.* (1992) observed that natural insecticides, neem, contains the active chemical azadirachtin, which disrupt the hormonal changes in *Bemisia tabaci* causing death during moulting.

In laboratory experiments Schmutterer (1992) applied concentration of 10 and 20 ppm/litre of azadirachtin, of an azadirachtin-free fraction and of 100 ppm/litre or an enriched, formulated seed kernel extract of *Azadirachta indica*, against the 5th larval of *Pieris brassicae*. Application of neem products against young (1st-3rd) larval instar of *Pieris brassica*, which may be typical under practical conditions, led to the death of the caterpillars.

Freisewinkel and Schmutterer (1991) showed that the topical application of neem extract at 0.25 to 1.0 ml/m² to the 5 nymphal instars of the gregarious phase of *Iocusta migratoria migratorioides* led to increased mortality during moults, prolonged development and reduced fitness. Morphogenetic effects were observed on the legs, wings and antennae. A reduction in weight corresponded to reduced feeding activity. Color changes and supernumerary moults suggested tendencies towards soliterization. The earlier the nymphs treated, and the higher the amounts applied, the more distinct the effects.

Salem (1991a) found that larval mortality ranged between 14.28% to 78.57% and the percent of eggs hatching ranged between 57.5% to 89.4% when different concentration from neem seed oil extract, were tested against the potato tuber moth, *Phthorimaea operculella* Zell.

Loke *et al.* (1990) evaluated six concentrations (1.25, 2.25, 5.0, 10.0, 20.0, and 40.0 percent) of neem oil in acetone, for contact toxicity against 2nd and 3rd larval instars of *Plutella xylostella* L. Significant mortalities of both larval stages were observed with neem oil concentration of 10 percent

and above. Although the lower concentration of neem oil appeared to be sub-lethal with regard to contact toxicity effect, physiological and growth disruptive effects, such as retardation of growth (prolonged), delayed adult emergence and abnormal adults but the effects were more pronounced in the younger instar. Subsequent treatment of pupae and adults of *Plutella xylostella* with neem oil concentrations of 1.25, 2.25, 5.0 and 10.0 percent showed that pupae were generally not affected by the concentrations tested. However, male and female adult moths treated with 2.5 percent and higher concentrations of neem oil had significantly higher mortalities in 48 hours and shorter longevities than the adults in control.

Mishra *et al.* (1990) reported that brinjal leaves treated with 0.025 and 0.05% neem oil to *Epilachna vigintioctopunctata* increased the duration of life stages in the subsequent generation.

Saxena (1985) found that insects fed far less, grew poorly and laid fewer eggs on rice plants treated with the oil, cake, extracts such as azadirachtin, and their formulation. Contact with or ingestion of neem seed derivatives disrupted growth of insect pests. Neem oil alone or in combination with seed oil of custard apple (*Annona squamosa* L.) was effective in reducing the survival of *N. virescens* and its transmission of grassy and ragged stunt viruses.

Schmutterer *et al.* (1984) investigated that topical application of neem oil on last instars *N. lugens*, *S. furcifera* and *N. virescens* nymphs resulted in their premature death. Seventy seven to 100% mortality of *S. furcifera* was caused by neem oil (Saxena *et al.* 1983).

Mariappan and Saxena (1983) reported that custard-apple oil, neem oil and their mixtures were effective in reducing the survival of the green leaf

hopper, *N. virescens* Distant and its transmission of the rice tungro virus (RTV).

Heyde *et al.* (1983) found that 2 to 4 days exposure of *Sogatella furcifera* to plants treated with 500 ppm of neem seed kernel extracts resulted in 75% mortality whereas in the control, mortality was only 5%. On third instar, *N. lugens* nymphs, a combination of foliar and topical application induced higher mortality (75%) than either application alone (30%).

Schmutterer *et al.* (1983) studied the morphogenetic effects of four partially purified fractions of neem seed extracts and two methanolic seed extracts on larvae of rice ear cutting caterpillar, *Mythimna separata* Walker, and the rice leaf folder, *C. medinalis*, larvae fed for 24 hours. On rice leaf cuts dipped in different solution of the partially purified fractions and methanolic extracts exhibited pronounced development abnormalities and mortalities in succeeding larval instars and in pupal and adults stages.

CHAPTER III

MATERIALS AND METHODS

The experiments were carried out in laboratory, greenhouse and in the field during March to October, 2016. The details of different materials used and methodology followed during the experimental period are described below.

3.1 Experimental site

The research work was carried out at laboratory and Greenhouse premises of BJRI, Dhaka and in the experimental field of Dhaka.

3.2 Soil

The experimental area belonging to the Agro-Ecological Zone (AEZ-7) "Active Brahmaputra and Jamuna Floodplain". The soil texture was sandy loam.

3.3 Climate

The experimental area was under the subtropical climate. Usually the rainfall was heavy during Kharif season and scantily in Rabi season. The atmospheric temperatures increased as the growing period proceeded towards Kharif season. The weather conditions of crop growth period such as monthly mean rainfall (mm), mean temperature ($^{\circ}\text{C}$), sunshine hours and humidity (%) were collected.

3.4 Land preparation

The land was prepared at 'JOE' condition by deep ploughing and harrowing followed by laddering and leveled properly. The seeds were sown after final preparation of land.

3.5 Experimental design and layout

The experiment was laid out in a Randomized Complete Block Design (RCBD) with three replications following the methods described by Gomez and Gomez. (1984).

3.6 Plot size

The size of the individual plot was 3 m × 2.1 m. The space between plot to plot and line to line was 0.75 m and 0.3 m, respectively.

3.7 Fertilizer Application

The following fertilizers (As prescribed fertilizer recommendation of Soil science department of BJRI) were used.

Name of the fertilizers	Rate (kg/ha)
Urea	200
TSP	50
MP	60
Gypsum	95
ZnSO ₄	11

The total amount of TSP, MP, Gypsum, Zinc sulphate and the half of urea were applied at the time of final land preparation. The remaining half of the urea was applied after 45 days of seed sowing.

3.8 Planting materials

The variety 0-9897 of *Corchorus olitorius* L. was used. The seeds were collected from the Breeder Seed Department of BJRI. Before sowing, seeds were tested for germination in the laboratory and the percentage of germination was found to be over 90%.

3.9 Pot preparation

Earthen pots (12" diameter) were brought from the market and filled with dairy soil and sand.

3.10 Inter cultural operation

Weeding, mulching and irrigation were done as and when necessary but no plant protection measures were taken.

3.11 The test pests

Test pests was jute yellow mite, *Polyphagotarsonemus latus* (Order: Acarina, Family: Tarsonemidae).



Plate 3: A partial view of field experiment



Plate 4: Egg & nymph of yellow mite under stereo microscope

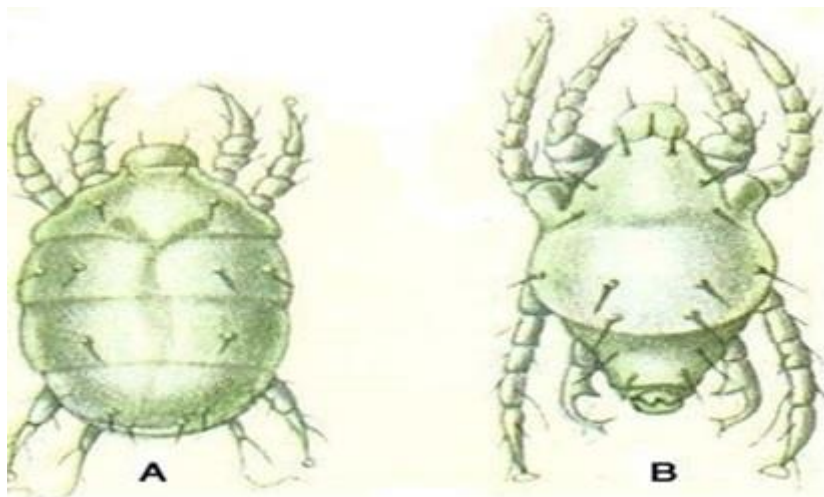


Plate 5: Adult Female (A) and male (B) jute yellow mite



Plate 6: Yellow mite infested plant



Plate 7: Pot experiment

3.12 Details of the treatments

Effectiveness of some indigenous plant materials were evaluated against yellow mite of jute in potted plants grown in green house premises. The selected plants were neem, mahogany, pithraj and turmeric. Mortalities of jute yellow mite were calculated by observing under microscope in greenhouse experiment. In the field condition, infestation level was determined by calculating deformed leaves.

A) Efficacy of specific plant part of a selected plants with three doses (Aqueous extraction) was determined. The treatments were as follows:

T₁ = Green neem leaf extract @ 1:10

T₂ = Green neem leaf extract @ 1:20

T₃ = Green neem leaf extract @ 1:30

T₄ = Pithraj seed extract @ 1:10

T₅ = Pithraj seed extract @ 1:20

T₆ = Pithraj seed extract @ 1:30

T₇ = Mahogany seed extract @ 1:10

T₈ = Mahogany seed extract @ 1:20

T₉ = Mahogany seed extract @ 1:30

T₁₀ = Turmeric powder extract @ 1:10

T₁₁ = Turmeric powder extract @ 1:20

T₁₂ = Turmeric powder extract @ 1:30

T₁₃ = Neem seed kernel extract @ 1:10

T₁₄ = Neem seed kernel extract @ 1:20

T₁₅ = Neem seed kernel extract @ 1:30

T₁₆ = Control (untreated)

B) Field experiments was conducted with the following treatments:

T₁ = Green neem leaf extracts @ 1:20

T₂ = Pithraj seed extracts @ 1:20

T₃ = Mahogany seed extracts @ 1:20

T₄ = Turmeric powder extracts @ 1:20

T₅ = Neem seed kernel extracts @ 1:20

T₆ = Control (untreated)

3.13 Extraction and formulation of some plant materials

3.13.1 Preparation of green neem leaf extract

Fresh green leaves were collected from the campus of BJRI, Dhaka. The collected leaves were washed under running tap water. Hundred grams of Air dried leaves were taken into an electric blender with 1000 ml of water. After blending the leaves well, the mixture were filtered through a fine cloth then the extract was considered as 1:10 neem leaf extract. Similarly 1: 20 and 1:30 neem leaf extracts were prepared by adding 2000 ml and 3000 ml of water with 100g of leaves, respectively.

3.13.2 Preparation of mahogany seed extracts

Matured mahogany fruits were collected from BJRI campus. Mahogany fruits were dried in the sun. Then seeds were separated from fruits. One hundred gram of seeds were crushed and dissolved in 1000 ml, 2000 ml and 3000 ml of water separately for overnight for the concentration of 1:10, 1:20 and 1:30 of mahogany seed extracts. The suspensions was filtered through linen cloth.

3.13.3 Preparation of neem seed kernel extract

Matured neem fruits were collected from neem plants of BJRI, Dhaka then they dipped in water for 48 hours for easy removal of the shell. After removing the hard shell the seed kernels were dried in air which was then made into dust with the help of a grinder. Then the whole dust was passed through a sieve (25-mesh diameter) to obtain fine and uniform dust. One hundred gram of dusts was dissolved in 1000 ml, 2000 ml and 3000ml of water separately for overnight to prepare the neem seed kernel extract of concentration 1:10, 1:20 and 1:30. The suspensions were filtered through linen cloth for spraying.

3.13.4 Preparation of pithraj seed extract:

Matured pithraj seeds were collected from Manikganj. Pithraj seeds were dried in the sun. One hundred gram of seeds water crushed and dissolved in 1000 ml, 2000 ml and 3000 ml of water separately for overnight to for the concentration of 1:10, 1:20 and 1:30 of pithraj seed extracts. The suspensions were filtered through linen cloth.

3.13.5 Preparation of Turmeric powder extracts:

One hundred gram of turmeric powder were soaked separately in 1000 ml, 2000 ml and 3000 ml of water and kept for 2 hours, then filtered through linen cloth to prepare 1:10, 1:20 and 1:30 (materials : water) turmeric powder extracts.

Aqueous extraction of seeds and leaves of different indigenous plants were prepared as described by Zebitz (1986) with few modifications.



Plate 8: Green neem leaf extract.



Plate 9: Pithraj seed extract.

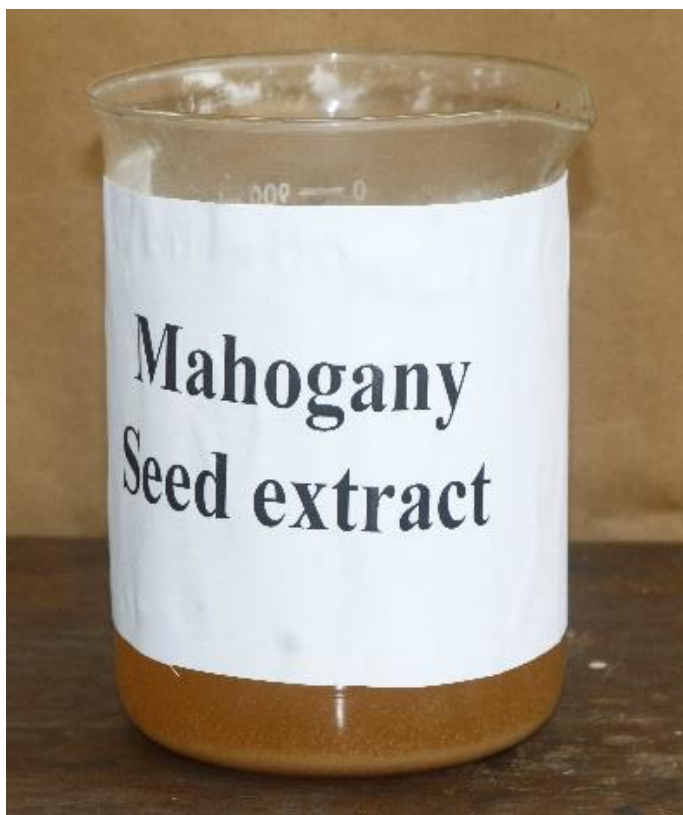


Plate 10: Mahogany seed extract.



Plate 11: Turmeric Powder extract



Plate 12: Neem seed Kernel extract.

3.14 Experimental details

3.14.1 Pot experiment

Pot experiment was conducted to determine the optimum dose for field experiment with aqueous extraction of plant materials. When sufficient infestations of yellow mite infestation were built up in all the pots, then extracts of green neem leaves, pithraj seed, mahogany seed, turmeric powder and neem seed kernel were sprayed on the infested plants. Each treatment was replicated three times. Yellow mite population were recorded by examining the leaf samples under binocular microscope before spray and after 24, 48 and 72 hrs. After spray for calculating % mortality. Percent mortality was calculated as per following formula:

$$\% \text{ corrected mortality} = \left(1 - \frac{T_a \times C_b}{T_b \times C_a}\right) \times 100$$

Where,

T_a= No. of mites after treatment

T_b=No. of mites before treatment

C_a= No. of mites after treatment in control

C_b= No. of mites before treatment in control

3.14.2 Experiment in field conditions

The efficacy of plant materials (aqueous extraction) were evaluated in field. After sufficient natural infestations built up in all the plots, green neem leaf, pithraj seed, mahogany seed, turmeric powder and neem seed kernel extracts were sprayed on infested plots with a repetition of 2nd spray

after 20 days of 1st spray. The control plots were kept untreated and each treatment was replicated 3 times. Number of infested plants were recorded before spray and at 7th day after spray for calculation of % reduction of infestation over control. Percent reduction was calculated as per following formula:

$$\% \text{ Reduction} = \frac{\text{No. of infested plants in control plot} - \text{No. of infested plants in treated plot}}{\text{No. of infested plants in control plot}} \times 100$$

3.15 Statistical analysis

The experiment were conducted in RCBD & CRD. The data were statistically analyzed to obtain the level of significance using the MSTAT software for analysis of variance developed by Russel (1986). The treatment means were separated by Duncan's Multiple Range Test (DMRT).

CHAPTER IV

RESULTS AND DISCUSSION

4.1 Effect of aqueous extraction of plant materials on jute yellow mite

Experiments were conducted to study the effect of different concentrations of plant materials such as green neem Leaf extract, Pithraj seed extract, mahogany seed extract, turmeric powder extract, neem seed kernel extract on percent mortality of yellow mite in pot. Results are shown in Table 1. Among the three different concentrations of neem seed kernel extract, the highest percent mortality (69.39%) was recorded in neem seed kernel extract @ 1:20 (T₁₄) followed by (T₁₃) 1:10 (65.34%) and (T₁₅) 1:30 (64.08%), respectively. In case of mahogany seed extract, the highest percent mortality (67.77%) was recorded at the dose 1:20 (T₈) followed by (T₇) 1:10 (65.31%) and (T₉) 1:30 (64.36%) respectively. Pithraj seed extract @ 1:20 (T₅) provided the highest mortality (63.86%) followed by (T₆) 1:30 (61.40%). In green neem leaf extract, the highest percent mortality (61.47%) was recorded at the dose 1:20 (T₂) followed by T₃ (60.59%). In case of Turmeric powder extract, all dose provided similar percent mortality T₁₁, T₁₀, T₁₂ (62.43%, 60.97% and 60.61%) respectively. Accordingly, pande *et al.* (1987) reported 1% neem leaf extract and 5% neem seed kernel which were very much effective against *Tetranychus neocaledomcus* and *Tetranychus urticae*. Based on the above findings, neem seed kernel extract @ 1:20, Mahogany seed extract @ 1:20, Pithraj seed extract @ 1:20, green neem leaf extract @ 1:20 and turmeric powder extract @ 1:20 were selected for field trial.

Table 1: Effect of plant materials (aqueous extraction) on percent mortality of jute yellow mite in the pot

Treatment	No. of mite/sq. cm leaf before treatment	% mortality over control at		
		24 hrs	48 hrs	72 hrs
T1	51	55.48 f	56.49 d	58.31 b
T2	48	59.43 cdef	60.58 abcd	61.47 ab
T3	52	56.16 f	57.59 cd	60.59 ab
T4	49	58.98 cdef	58.43 bcd	60.13 ab
T5	48	61.40 bcd	61.48 abcd	63.86 ab
T6	45	56.80 ef	58.07 bcd	61.40 ab
T7	54	65.34 ab	62.44 abcd	65.31 ab
T8	50	63.30 bc	65.71 ab	67.77 ab
T9	43	60.52 cde	63.19 abcd	64.36 ab
T10	45	58.72 def	60.59 abcd	60.97 ab
T11	45	62.35 bcd	62.52 abcd	62.43 ab
T12	52	63.33 bc	61.95 abcd	60.61 ab
T13	51	65.07 ab	65.96 ab	65.34 ab
T14	52	68.56 a	67.96 a	69.39 a
T15	49	65.56 ab	64.89 abc	64.08 ab
T16	40	–	–	–
LSD(0.05)	–	3.811	6.651	7.962
CV (%)	–	4.65	8.05	9.47

Means within a column followed by same letter(s) are not significantly different at 5% level.

T₁ = Green neem leaf extract @1:10, T₂ = Green neem leaf extract @1:20, T₃ = Green neem leaf extract @1:30, T₄ = Pithraj seed extract @1:10, T₅ = Pithraj seed extract @1:20, T₆ = Pithraj seed extract @1:30, T₇ = Mahogany seed extract @1:10, T₈ = Mahogany seed extract @1:20, T₉ = Mahogany seed extract @1:30, T₁₀ = Turmeric powder extract @1:10, T₁₁ = Turmeric powder extract @1:20, T₁₂ = Turmeric powder extract @1:30, T₁₃ = Neem seed kernel extract @1:10, T₁₄ = Neem seed kernel extract @1:20, T₁₅ = Neem seed kernel extract @1:30, T₁₆ = Control.

4.2 Effect of different plant materials on yellow mite (1st spray) under field condition

The effect of different plant materials on percent reduction over control of plant infestation by yellow mite was determined in the field. Results of Table 2 indicated that after 1st spray, all different plant materials reduced considerable amount of plant infestation. The highest reduction of plant infestation was 68.65% recorded in neem seed kernel extract (T₅) treated plots followed by mahogany seed extract, T₃ (63.07%) treated plots. Pithraj seed extract (T₂) provided reduction of plant infestation (57.38%). The lowest plant infestation reduction was recorded in turmeric powder, T₄ (53.54%) followed by green neem leaf extract, T₁ (54.68%). The results indicated that neem seed kernel extract and mahogany seed extract were most effective, while Pithraj seed extract and turmeric powder extract were moderately effective against jute yellow mite.

It was reported by Anonymous (2009) that the highest 60% reduction of yellow mite infestation was recorded in 1:20 ratio of mahogany seed extract.

Banu et al. (2007) reported that green leaf extract and dry neem leaf extract reducing yellow mite infestation over control in jute. It has also been reported that different plant materials were controlled various insect pests

(Pasini et al 2003, Karmakar and Bhole, 2001. Karecm and Durairaj, 1987, Naganagouda et al. 1997).

In the present study, the results of field trial clearly indicates that neem seed kernel extract, mahogany seed extract and pithraj seed extract were most effective plant materials while green neem leaf extract and turmeric powder extract were moderately effective against jute yellow mite.

4.3 Effect of different plant materials on yellow mite (2nd spray) under field condition

After 2nd spray in the field (20 days after 1st spray) the effect of different plant materials of percent reduction of plant infestation over control was studied and results are shown in Table 3. It was found that all different plant materials reduced remarkable amount of plant infestation significantly over control. The highest reduction of plant infestation (70.20%) was recorded from neem seed kernel extract (T₅) treated plot followed by mahogany seed, T₃ (68.46%). Pithraj seed extract, T₂ treated plots provided 65.38% reduction. The lowest plant infestation reduction 61.72% and 62.37% were recorded in green neem leaf extract (T₁) and turmeric powder extract (T₄) over control. Which were statistically similar. These results indicated that neem seed kernel extract and mahogany seed extract were most effective, while Pithraj seed extract was moderately effective. Islam (2006) showed 1% neem oil and green neem leaf extract at the dose 1:20 is very much effective for reducing mite population in jute.

4.4 Effect of different plant materials on internodes of jute plant

The effect of different plant materials on number of internodes per plant at three growth stages of jute plant are presented in (Table 4). At early stage, the lowest number of nodes/plant (20.05) was recorded in neem seed kernel

extract (T₅) followed by 21.30 (mahogany seed extract, T₃) which were statistically similar but significantly different from other treatment. The untreated control, T₆ (36.00) had the highest number of nodes per plant, which was significantly different from other treatments. At the middle stage, the lowest number of nodes per plant 33.85 was recorded from treatment neem seed kernel extract (T₅) followed by 35.85 mahogany seed extract (T₃) which was statistically similar but different from other treatment. The highest number of nodes per plant 80.25 was observed in untreated control (T₆) which was significantly different from other treatments.

On the other hand, the late stage of plant, the lowest 52.50 number of nodes per plant was recorded from neem seed kernel extract (T₅) which was statistically similar to 54.50 (T₃) and 57.50 (T₂). The highest 92.50 number of nodes per plant was observed in treatment T₆ (untreated control), which was significantly different from other treatments. The results indicate that neem seed kernel extract was most effective followed by mahogany seed extract, pithraj seed extract, green neem leaf extract and turmeric powder extract. It was also clear that different plant materials showed the better performance on reducing the number of nodes per plant at all stages of jute plant growth than untreated control plant. It should be noted that severe infestation of mite on jute plant causes defoliation, stunting of plant growth, increased the number of nodes per plant (Plate 13) and decreased the fibre quality. Because application of different plant materials reduced mite infestation and increased plant height that decreased the number of nodes with in a unit area. The effect of different plant materials in the present study was in accordance with the findings observed by Pande *et al.* (1987) and Devraj (1990). They found that, neem leaf extract 1% and neem

seed kernel extract at 5% were effective against mites. So the effectiveness of different plant materials against mites proved promising in the present study. Therefore different plant materials can be used for the control of yellow mite on jute.

Table 2: Effect of different plant materials on % reduction of yellow mite in field condition (First spray) over control

Treatment	Average no. of mite infested plant/plot before spray	No. of mite infested plant after 7days of spray	% reduction over control
T ₁	74.25	26	54.68 C
T ₂	62.5	23	57.38 BC
T ₃	81	21	63.07 ab
T ₄	88.5	25	53.54 C
T ₅	68.25	20	68.65 A
T ₆	75	67.25	0.000 D
LSD(0.05)	-	-	4.897
CV (%)	-	-	6.6

Means within a column followed by same letters are not significantly different at 5% level.

T₁=Green Neem Leaf Extract @ 1:20

T₂=Pithraj Seed Extract@ 1:20

T₃=Mahogany Seed Extract@ 1:20

T₄=Turmeric powder extract@ 1:20

T₅=Neem Seed Kernel Extract@ 1:20

T₆=Control

Table 3: Effect of different plant materials on % reduction of yellow mite in field condition (2nd spray) over control

Treatment	Average no. of mite infested plant/plot before spray	No. of mite infested plant after 7days of spray	% reduction over control
T ₁	48.75	15.75	61.72 c
T ₂	43	14.25	65.38 bc
T ₃	37	13	68.46 ab
T ₄	34.25	15.5	62.37 c
T ₅	35.25	12.25	70.20 a
T ₆	49.25	41.25	0.000 d
LSD(0.05)	-	-	4.216
CV (%)	-	-	5.12

Means within a column followed by same letters are not significantly different at 5% level.

T₁=Green Neem Leaf Extract @ 1:20

T₂=Pithraj Seed Extract@ 1:20

T₃=Mahogany Seed Extract@ 1:20

T₄=Turmeric powder extract@ 1:20

T₅=Neem Seed Kernel Extract@ 1:20

T₆=Control

Table 4: Effect of different plant materials on number of inter nodes of Jute plant

Treatment	No. of inter nodes at		
	Early stage (35 DAS)	Middle stage (80 DAS)	Late stage (120 DAS)
T ₁	25.50c	46.50b	62.25b
T ₂	24.25cd	37.70c	57.50c
T ₃	21.30de	35.85c	54.50c
T ₄	27.25b	47.50b	66.25b
T ₅	20.05e	33.85c	52.50c
T ₆	36.00a	80.25a	92.50a
LSD(0.05)	2.71	4.19	4.09
CV (%)	8.95	5.83	4.18

Means within a column followed by same letter(s) are not significantly different at 5% level.

T₁=Green Neem Leaf Extract @ 1:20

T₂=Pithraj Seed Extract@ 1:20

T₃=Mahogany Seed Extract@ 1:20

T₄=Turmeric powder extract@ 1:20

T₅=Neem Seed Kernel Extract@ 1:20

T₆=Control



Plate 13: Severely yellow mite infested jute plant showing internodes and increased number of nodes

4.5 Effect of different plant materials on plant height.

The effect of different plant materials on height of jute is presented in Table 5. At the time of harvest, the highest plant height 3.10 m. was recorded in the treatment neem seed kernel extract (T₅) followed by mahogany seed extract, T₃ (3.03) and pithraj seed extract, T₂ (3.02 m) respectively. These are statistically similar. The lowest plant height was recorded from untreated control plot, T₆ (2.90m) which was significantly lower than all other treatments treated plots. The above results indicate that different plant materials had significant effect on increasing height of jute plant.

Mite infestation causes growth stunting and finally reduces the plant height. Application of different plant materials decreased mite infestation and ultimately increased plant height. Therefore, different plant materials were reduced yellow mite infestation are increased plant height. Islam (2007) reported that the plant height 314 cm and 299 cm were recorded from @ 1:20 neem seed kernel extract and @1:20 green neem leaf extract. The effect of plant materials on increasing plant height as observed in the present study was in conformity with finding reported by Palaniswamy and Ragini (2000) against yellow mite on chili. They observed that 5% aqueous extract of neem leaf reduced mite population on chili and increased height.

4.6 Effect of different plant materials on base diameter of jute

The effect of different plant materials on base diameter of jute is presented in Table 6. At the time of harvest, the highest base diameter 15.93mm was recorded in the treatment neem seed kernel extract (T₅) followed by mahogany seed extract, T₃ (15.86) which was statistically similar. The lowest plant base diameter was recorded from untreated control plot, T₆ (14.50mm) which was significantly lower than all other treatments treated plots. The above results indicate that different plant materials had significant effect on increasing base diameter of jute plant.

4.7 Effect of different plant materials on fibre yield of jute

Effect of different plant materials on yellow mite has influence on yield of jute fibre. The highest , (38.60%) fibre yield was increased over untreated control, T₆ as shown in Table 7 followed by 37.04% from mahogany seed extract, T₃ treated plot which was statistically similar. The lowest (27.12%) fibre yield was recorded from turmeric powder extract, T₄ treated plots, which were significantly different from other treatment, pithraj seed extract

(T₂) and green leaf extract (T₁) significantly increased fibre yield by 33.25% and 29.41% respectively over control having no significant difference between them. It was reported by anonymus (2009) that 27% fibre increased over control was from magony seed extract treated plot and 23.5% fibre yield increased over control was reported from jute seed extract treated plot. The present study indicate that plant materials performed increased fibre yield over control. Similar findings were reported by Banu *et al.* (2007). They reported that the use of green neem leaf extract and dry neem leaf extract against yellow mite gave increased fibre yield over control.

Table 5: Effect of different plant materials on plant height.

Treatment	Plant Height (m.)
T ₁	3.00b
T ₂	3.02ab
T ₃	3.05ab
T ₄	2.98b
T ₅	3.10a
T ₆	2.90c
LSD(0.05)	8.025
CV (%)	4.77

Means within a column followed by same letter(s) are not significantly different at 5% level.

T₁=Green Neem Leaf Extract @ 1:20

T₂=Pithraj Seed Extract@ 1:20

T₃=Mahogany Seed Extract@ 1:20

T₄=Turmeric powder extract@ 1:20

T₅=Neem Seed Kernel Extract@ 1:20

T₆=Control

Table 6: Effect of different plant materials on base diameter

Treatment	Plant Base Diameter (mm)
T ₁	15.24 b
T ₂	15.34 b
T ₃	15.86 a
T ₄	15.39 b
T ₅	15.93 a
T ₆	14.50 c
LSD(0.05)	0.452
CV (%)	1.94

Means within a column followed by same letter(s) are not significantly different at 5% level.

T₁=Green Neem Leaf Extract @ 1:20

T₂=Pithraj Seed Extract@ 1:20

T₃=Mahogany Seed Extract@ 1:20

T₄=Turmeric powder extract@ 1:20

T₅=Neem Seed Kernel Extract@ 1:20

T₆=Control

Table 7: Effect of different plant materials on fibre yield

Treatment	Yield (t/ha)	% Increased yield over control
T ₁	2.629 cd	29.41 bc
T ₂	2.706 bc	33.25 ab
T ₃	2.782 ab	37.04 a
T ₄	2.581 d	27.12 c
T ₅	2.815 a	38.60 a
T ₆	2.033 e	0.000 d
LSD(0.05)	0.095	5.29
CV (%)	2.57	12.73

Means within a column followed by same letter(s) are not significantly different at 5% level.

T₁=Green Neem Leaf Extract @1:20

T₂=Pithraj Seed Extract@1:20

T₃=Mahogany Seed Extract@1:20

T₄=Turmeric powder extract@1:20

T₅=Neem Seed Kernel Extract@ 1:20

T₆=Control

CHAPTER V

SUMMARY AND CONCLUSION

In order to study the effectiveness of plant materials against yellow mite of jute, several experiments were conducted in the pot and field condition of Bangladesh Jute Research Institute during the period of March 2016 to October 2016.

Initially, aqueous extraction of different doses of plant materials including green neem leaf, neem seed kernel, mahogany seed, pithraj seed and turmeric powder were evaluated against yellow mite in the pot of greenhouse premises to find out suitable dose for the field experiment. The highest percent mortality 69.39%, 67.77%, 63.86%, 62.43% and 61.47% @ 1:20 were selected for field trial.

In case of first and second spray (field experiment), the highest percent reduction of infestations over control (68.65 and 70.20) were observed by neem seed kernel extract. However, they were statistically different than other treatments. The lowest percent reduction of infestations over control (53.54 and 61.72) were recorded by turmeric powder extract (1:20) and green neem leaf extract.

At the late stage of plants, highest number of nodes (92.50) per plant was found in control plots due to shortening of internodes caused by severe mite infestation which was statistically different than other treatments. The lowest number of nodes (52.50) per plant was found in neem seed kernel which was statistically similar with mahogany seed extract. Plant height

was found to be the highest (3.10 meter) in the plot treated by neem seed kernel extract and the lowest (2.90 meter) in control plot. The highest 15.86mm base diameter was found in neem seed kernel treated plot and the lowest 14.50 mm base diameter was recorded in control plot. Maximum percent yield increase over control (38.60) was found in the plot treated with neem seed kernel extract and the lowest percent yield increase over control (27.12) was found in the plot treated with turmeric powder extract.

As the dose 1:20 of different plant materials showed efficacy, it was used for further field trial. The result of field trial clearly indicate that neem seed kernel extract, mahogany seed extracts were highly effective plant materials while pithraj seed extract, green neem leaf extract and turmeric powder extract were moderately effective against jute yellow mite. These plant materials can be safely used by the farmers as components of IPM, a low cost alternative and a promising known botanical pest control agent and to save the crops from different insect pests effectively and economically without hampering the ecological balance agro-ecosystem. The present study will presumably help in future research to improve the effectiveness of plant materials as pesticides for the benefit of farmers and it can be used as a components of IPM program.

REFERENCE

- Adityachaudhury, N.A., Bhattacharyya, A., Chowdhury and Pal, S. (1985). Chemical constituents of plants exhibiting insecticidal, anti feeding and insect growth regulating activities *J. Sci. Industr Res.*, 44: 85-101.
- Al Maisary, M. F. and Al Rahawi. H.M.H. (2004). The effect of different concentrations of neem oil extract of some immature stages of *Culex pipiens* L. mosquito. University of Aden Journal of Natural and Applied Sciences, 8 (2): 213-218.
- Anonymous, (2005). Tips on Managing Common Bedding Plant Insects and Mites Pests. University of Connecticut Integrated Pest Management. 3: 350-351.
- Anonymous, (2009). Bangladesh Bureau of Statistics, Yearbook of Agricultural Statistics of Bangladesh. 29th Edition, Planning Division, Ministry of Planning, Government of the People's Republic of Bangladesh. 584 pp.
- Atwal. A. S. (1976). Agricultural pests of India and south East Asia. Kalyani Publishers, New Delhi. 502 p.
- Baker, J.R. (1997). Cyclamen mite and broad mite. Ornamental and Turf Insect Information Notes, 6: 41-46.
- Banks, N. (1904). Class III, Arachnida, Order 1, Acarina, four new species of injurious mites. Journal of the New York Entomological Society 12: 53-56.
- Banu, H. and Singh, P.K. (2007). Integrated Management of Jute Pest. *Appl Environ. Ecol.* 28(h): 186-203.

- Banu, IE, Sultana, K., Islam, M.N., Polan, M.S. and Kamrutfzaman, A.S.M. (2007). Comparative study of non-chemical approach and chemical pesticides for the management of jute pests. *Int. J. Sustain. Agril. Tech.* 3(5): 37-40.
- Barman, S.K. (1993). *Azadirachtin* affects growth and survival of immature tawny mole cricket. *Fla. Entomol.* 76(3): 526-530.
- BBS (Bangladesh Bureau of Statistics). (2015). Statistical Yearbook of Agricultural Statistics-2015, Ministry of Planning, Government of the People's Republic of Bangladesh, pp. 165-167.
- Becker, H., Corliss, J. and De Quattro, J. (1992). Control of *Bemisia tabaci* and the potential of *paecilomyces fumosoroseus* as a biopesticide. In *Bio-control News and Information*, 14 (4): 71-78.
- Behera, U.K. and Satapathy. C.R. (1996). Screening indigenous plants for their insecticidal properties against *Spodoptera litura* Fab. *insect Environ.* 2 (2): 43-44.
- Brown, R.D. and Jones, V.P. (1983). The Broad Mite on Lemons in Southern California. *California Agriculture*, 37(7/8): 21-22.
- Chiu-Shin-Foon and Qiu-Yu-Tong. (1993). Experiments on the application of the botanical insecticides for the control of diamondback moth in South China. *J. Appl. Entomol.*
- Deka, N. and Hazarika, L. K. (1997). Effect of neem seed oil on food consumption and utilization by rice hispa. *Pesticide Res. J.* 9 (1): 113-116.
- Denmark, H.A. (1980). Broad mite, *Polyphagotarsonemus latus* (Banks). FDACS DPI Bureau of Entomology Circular No, 213. 2.

- Devraj Urs, K.C. (1990). Efficacy of certain plant products in the control of brinjal red spider mite, *Tetranychus urticae*. Fourth Nat. Sym, In Acarology, Kerala, 12-14 Oct. 1990. Division of Acarology, Department of Zoology, University of Calicut, KTC Offset Press, Calicut.
- Eungwijarupanya, S., Yimchareon, S., Sumrid, Y. and Wylie, F.R. (2002). Control of teak defoliator *Myblaea parea* Gamer (Lepidoptera: Hybleidae). by thermal fogger application of neem extract. Preceding of the IUFRO-FAO workshop on pest management in tropical forest plantation. Chanthaburi. Thailand. 25- 29 May, 1998. FORSPA-Pblication. 30: 123125.
- Freisewinkel, D. C. (1993). Oral and topical effects of seed oil from the neem tree, *Azadirachta indica* (A. Juss.) on nymphs of the African migratory locust, *Locusta migratoria Migratorioides* (R.et. F.) *Deutsche Gesellschaft fur Allagemine and Wandte Entomologie*, pp. 785-789.
- Freisewinkel, D. C. and Schmutterer, H. (1991). Contact action of neem oil on the African migratory locust, *Locusta migratoria Migratorioides*. *Zeitschrift -fur- Angewandte-Zool.* 78 (2): 189-203.
- Gonzalez Gomez, R., Otero Colina, G., Villanueva Jimenez, J, A., Perez Amaro, J.A. and Soto Hernandez, R.M. (2006). *Azadirachta indica* toxicity repellence of *Varroa destructor* (Acari: Varroidac). *Agrocieneia-Montecillo.* 40 (6): 741-751.

- Gujar, G.T. (1997). *Neem (Azadirachta indica. A. Juss)*, a natural insecticide: Status and need for bioassay standards. *J. Pestic. Res*, 4 (ii): 69-79.
- Gupta, M.A. (2003), Comparative efficacy of neem products and endosulfan against major insect pests of sesame. *Indian J, Plant Protect.* 31 (2): 96-97.
- Haque, M.A. and Islam, B.N. (1988). Effects of neem oil on the food consumption and survival of *Epilachna dodecastigma* (Wied). *Bangladesh J, Entomol.* 6 (1-2): 1-5.
- Haque, M.A., Parvin, N. and Ahmed, K.S. (1996). Effect of neem oil on the food consumption and survival of *Epilachna dodecastigma* (Wied). *Bangladesh J. Entomol.* 6. 1-2.
- Heyde, J. Vd., Saxena, R. C. and Schmutterer, H. (1983). Neem oil and neem extracts as potential insecticides for control of Hemipterous rice pests. *Abs. 2nd Ini. Neem Conf.* Rausch Holzhausen Castle, FRG, May 1983, p. 27.
- Higa, S.Y. and Namba, R. (1970). Vectors of the Papaya Mosaic Virus in Hawaii.
- Hill, D. S. (1983). *Polyphagotarsonemus latus* (Banks). In: *Agricultural Insect Pests of the Tropics and Their Control*. Cambridge University Press. 746 pages.
- Imtiaz, A., Farzana, S., Azmi, M.A., Kahlaghan, A., Rizvi, S.A., Muzaffer, A. and Shakoori, A.R. (2001). Comparative toxicological studies of neem leaf extract and cyhalothria (Pyrethroid) against *Sitophilus oryzae* and their effects on

alkaline phosphatase activity. Proc, Pakistan congr, 2001. 21:255-261.

Islam, M.B. (1993). Growth inhibitory and antifeedant effects of azadirachtin on six noctuids of regional economic importance. *Pestic. Sci.* 38 (1): 57-63.

Islam, M.S. (2006). Exploration of plant materials extracts for the control of jute yellow mite, *Polyphagotarsonemus latus* Banks. M. S. Thesis. Department of Entomology, Sher-e-Bangla Agricultural University, Sher-e-Bangla Nagar. Dhaka.

Jagjeet, S., Chhilar, B.S. and Kashyap, R.K. (2005). Effectiveness of various seed protectants on adult mortality on pulse beetle, *Callasobrachus maculatus* (F) infesting pigeon pea seeds. *Annals Biol.* 21(1): 65-67. S

Jeyakumar, P. and Uthamasamy, S. (1997). Bio- efficiency of some synthetic insecticides and botanicals against *Liriamyza trifoli*. *Indian J. Entomol.* 59 (4): 347-350.

Johnson, W.T., and Lyon, H.H. (1991). *Insects that Feed on Trees and Shrubs*. 2nd ed., rev. Comstock Publishing Associates, 560 p.

Jotwani, M.G. and Srivastava, K.P. (1984). Neem in insect control. *Neem newsletter* 1 (I): 5.

Jyothi, M. and Sannaveerappanavar, V.T. (2003). Contact toxicity of aqueous neem seed kernel extract and neem seed oil to different life stages of *Cheilomenes sexmacidata* (Fabricius) (Coleoptera: Coccinellidae). *Biological control of Lepidoptera*

pests proceedings of the symposium of Biological control Lepidoptera pests, July- 17-18. Bangalore, India, pp. 47-48.

Kabir, A. K. M. F. (1975). In jute pest of Bangladesh. Annual Report 1975. Bangladesh Jute Research Institute. Dacca, pp 28-33.

Kareem, A.A. and Durairaj, C. (1987). Evaluation of neem derivatives for control of major insect pests in rice in Tamil Nadu. India. Agril. Coll. and Res. Inst., Maduraj (Tndia).

Karmakar, M.S. and Bhole, S.R. (2001). Efficacy and persistent toxicity of some neem products against adult of *Epilachna dodecastigma* Wcid. Plant Proc. Bull, Faridabad. 53 (1-2): 22-25.

Khandakar, A.L. (1987). Jute seed at farm level. Agril. Econ. and Soc. Sci Prog. Bangladesh Agril. Res. Coun. Dhaka. Bangladesh.

Lacob, Van T. (2007). Organophosphate insecticides and acaricides antagonise bifenthrin toxicity through esterase inhibition in *Tetranychus urticae*. Chichester, UK: John

Loke, W. H., Heng. C. K., Norlaili - Basirum and Azman- Rejab. (1990). Non-target effects of neem (*Azadirachta indica* A. Juss.) on *Plutella xylostella* L., Cabbage, Sawi and Padi. Malaysian Plant Protection Society (Malaysia). *Proc. (If the 3rd Int. Conf. on Plant Protection in the Tropics*. 11; 108-110.

Lowery, D.T., Bellerose, S., Smirle, M.J., Vincent, C. and Pilon, J.G. (1996). Effect of neem oil on the growth and development of the obliquebanded leaf roller, *Choristoneura rosaceana*. *Appl. Entomol* 79 (2): 203-209.

- Mariappan, V. and Saxena, R.C. (1983). Effect of custard apple oil and neem oil on survival of *Nephotettix virescens* (Homopters: Cicadcllidae) and on rice tungro virus transmission. *J. Econ. Entomol* 76: 573-576.
- Mayabini, J. (1997). Efficacy of Some neem products against rice leaf folder. *Cnaphalocrosis medinalis Guenee. Indian J. Plant Prot.* 25(2): 160-161.
- Mirsha, B.K., Mishra, P.R. and Patnaik, N.C. (1990). Effect of neem oil on the growth and development of Epilachna beetle, *Epilachna vigintioctopunctata* (FTbst). *OrissaJ. Agril. Res.* 2 (3-4): 169-172.
- Naganagouda, A., Sreenivasa, A.G., Rahman, S.M. and Patil, B.V. (1997). Evaluation of some plant products against major pests of paddy in Tungabhadra project area. *KarnatakaJ. Agril. Sci.* 10 (4): 1193-1196.
- Nauman, K. and Isman, M.B. (1995). Evaluation of neem (*Azadirachta indica*) seed extracts and oils as oviposition deterrent to noctuid moths. *Entomol. Expt.-et- Appl.* 76(2): 115-120.
- Nesseh, O.M., Freress, T. and krall, S. (1993). Repellent effect of *Azadirachta indica* (A. Juss.) On akults of *Schistocerca gregaria* Forskal investigations in the Republic of Niger. *Deutsche Gesells Chaft fur Allgemeine and Ahge Wandte Entomlogie*, p.835-938.
- Nicol, C.M.Y. (1993). Further studies on contact effects of seed oil from the neem tree, *Azadirachta indica* (A. Juss) on nymphs of the gregarious phase of the desert locust, *Schistocerca* F. *Deutsche*

Gesellschaft für Allgemeine and Ange Wandte Entomologic. 791-793.

Padmasheela, N. C. and Delvi, M. R. (2002). Antifeedant and mortality effects of neem oil (0.03% azadirachtin) against 3rd instar grubs of *Orytes rhinoceros* L. (Coleoptera: Scarabaeidae). *J. Ent. Res.* 26 (3): 239-244.

Palaniswamy, S., and Ragini, JC. (2003). Influence of certain plant extracts on yellow mite *polyphagotersonemus Latus* (Banks) on chillies. Department of Agricultural Entomology. Tamil Nadu Agricultural University. *Insect Environment.* 6:1, 25-26.

Palevsky, E., Soroker, V., Weintraub, P., Mansour, F., Abo-Moch, F. and Gerson, U. (2001). How species-specific is the phoretic relationship between the broad mite, *Polyphagotarsonemus latus* (Acari: Tarsonemidae), and its insect hosts? *Experimental & Applied Acarology* 25: 217-24.

Pande, Y.D., Majumdar, M. and Roy, H.L. (1987). Laboratory evaluation of various concentration of neem (*Azadirachta indica*) leaf extract against *Tetranychus neocaledonicus* Andre infesting okra in tripura. First Nat. Sem. On acarology, Kalyani, West Bengal, Oct. 29-31, Abstract no. 43.

Pasini, A., Capelo. S.M.J, and Oliveira, R.C. (2003). Preliminary assays for efficiency evaluation of neem oil for control of *Olygonychus yothersi* (Acari: Tetranychidae). *Setnina londrina.* 24 (2): 315-316.

Pena, J.E. and Campbell, C.W. (2005). Broad mite. EDIS. New Delhi, India: Malhotra Publishing House, 25: 944-949.

- Pons, L. (2007). Prompt progress made against a new threat to watermelon. *USDAARS News & Events*, 53, 67–107.
- Qureshi, M.S., Jackson, T.A., Townsend, R.J., Saville, D.J. and Zydenbos, S.M. (2002). Toxicity of neem and pyrethrum extracts to adult grass grub, Proc. Conf. Central Hotel, Routers, New Zealand, 11-15 August 2002. pp. 298-302.
- Raguramans, S. and Rajasckaran, B- (1997). Behavioural and physiological effects of neem on rice brown plant hopper, *Nilaparavata lugens*, *Madras Agril. J.* 84 (6): 326-328.
- Rahman, M.A., Ahmad, M., Shahjahan. M. and Ahamad, M. (2003). Laboratory evaluation of plant seed oils against the lessed meal worm, *Alphitohius diaperinus* (Panzer). *J. Asiat, Soc. Bangladesh Sci.* 29 (1): 11-18.
- Rani.C.S., Vijaya Lakhmi. K. and Rao, P.A. (2000). Vegetable oils of surface protectants against bruchid, *Callasohrachus chinensis(h.)* infestation on chickpea. *Indian J. Plant Protect.* 28(2): 184-186.
- Ranjana,S., Becnam.S., Saxena.R. and Saxena, B. (2000). Bioactivity of certain plant extracts against *Callasohrachus maculates* F. *J. App. Zool Res.* 11 (1): 29-32.
- Rao, G.R., Raghavaiah. G. and Nagalingam. B. (1993). Effect of botanicals on certain behavioural responses and on the growth inhibition of *Spodoptera litura* F. *Botanical Pesticides in Integrated pest management*, pp. 175-182

- Reddy, M.U., Bharati, S.R. and Reddy, D.D.B. (1999). Efficacy of some vegetable oils as protectants against the pulse beetle, *Callosobruchus chinensis* L. in green gram, *Phaseolus aureus* during storage. *Indian J, Nutr. Diet.* 36 (10): 436-442.
- Rovesti, L. and Deseo, K.V. (1990). *Azadirachta indica* A Juss. (neem) and its potentiality in the control of insects. *Informatore-Fitopatologica.* 40 (11): 27- 32.
- Salem, S.A. (1991 a). Response of cotton boll worms, *Pectinophora gossypiella* Saund and *Earias insulana* Boisd. to seed neem kernel pure oil. *Ann. Agric. Sci. Moshtohor.* 29 (1): 597-608.
- Salem,S.A. (1991b). Response of cotton bollworms, *Pectinophora gossypiella* Saund and *Earias insulana* Boisd to seed neem kernel pure oil. *Ann Agric. Sci., Moshtohor.* 29(1): 597-608.
- Salem. S.A. (1991a). Evaluation of neem seed oil as tuber protectant against *Phthorimaea operculella* Zell. *Ann. Agric. Sci. Moshtohor.* 29(1): 589-596.
- Sanguanpong, U. and Sehmutterer, H. (1992). Laboratory studies on the effect of neem oil and neem seed extracts on the spider mite, *Tetranychus urticae* Koch (Acari: Tetranychidae). Institute fur phytopathologic and Angewandte zoologie der Justus- Liebig-Universitat giessen. *Zeitschrift-fur- pflanzenkrankheiten-und-Pflanzenschutz.* 99:6, 637-646; 25.
- Saxena, R. C. (1989). In: *Insecticides of plant origin* Eds. B. J. R. Philogene and P. Morand. ASC Symposium Series 387, Washington, DC, pp. 110-135.

- Saxena, R.C. (1985), Evaluation and utilization of plant derivatives for control of insect pests in rice. *Philippine. J. Sc L* 10 (1): 15.
- Schmulterer, H., Saxena, R.C. and Heyde, V. (1983). Morphogenetic effects of some partially purified fractions and methanol ic extracts of neem seeds on *Mythimna separate* Walker and *Cnaphatocrosis medinalis*. *G. Z. Angew. Ent.* 95: 230-237.
- Schmuttercrer, H., Hedyde, J.V.D. and Saxena, H. (1984). Effects of neem oil on plant hopper and leaf hopper pests of rice, *Oryza sativa*. College, Laguna (Philippines).
- Shaminathan. V.R. and Jayaraj, S. (2001). Evaluation of botanical pesticides against the mealybug. *Perrisia virgata* Cockrell (Homoptera: Pseudococcidae) on cotton. *Madras Agril. J.* 88 (7-9): 535-537.
- Sharma, R. K. (1999). Efficacy of neem products against storage pests in maize. *Annals Agril. Res.* 20 (2): 198-201.
- Shin- Foon, C. and Zhang, Y.G. (1984), Effects of some plant materials of Meliaceae on fifth instar larvae of *Spodoptera litura* as feeding inhibitors. *Neem News Let.* 1(3):23.
- Solsoloy, A.D. and Embuido, A,G, (1992), Efficacy of neem, *Azadirachta indica* A. JLISS. For controlling cotton boll worm, *Helicoverpa armigera* (Hubn). *Cotton Res. J.* 5 (1-2): 76-77.
- Sudipta, C. and Sanjib, C. (1998). Food with neem oil affects life and development of rice moth, *Corcyra cephalonica* (Stainton) (Lepidoptera: Pyralidac). *Entomol* 23 (2): 153-156.

- Talukder, D., Khan, A. R., Hasan, M. (1989). *thuringiensis* Var. Kurstaki. *Entomophaga*. 34(4):587-589. Growth of *Diacrisia obliqua* [Lepidoptera:Arctidae] with low doses of *Bacillus*.
- Venkateswarlu, P. Sharma, C.M. and Singh, R.S.J. (1993). Effect of neem oil on growth and development of mustard aphid, *Lipaphis erysimi* (Kalt.). *Bioved.* 4 (2): 139-142.
- Waterhouse, D.F. and Norris, K.R. (1987). Chapter 31: *Polyphagotarsonemus latus* (Banks). In: *Biological Control Pacific Prospects*. Inkata Press: Melbourne.
- Zhu, S., Xing, G.Z., Jin, D., Xu., Chang, Y. and Liu, F. (2004). Control effects and biological activity of azadirachtin on rice stem borer, *Chilo suppressalis* in paddy fields. *Chinese J. Rice. Sci.* 18 (6): 551-55.